

Putting Beta-Diversity on the Map: Broad-Scale Congruence and Coincidence in the Extremes

Meghan W. McKnight^{1*}, Peter S. White², Robert I. McDonald³, John F. Lamoreux^{4,5}, Wes Sechrest⁶, Robert S. Ridgely⁷, Simon N. Stuart⁴

1 Curriculum in Ecology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, **2** Department of Biology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, **3** Graduate School of Design, Harvard University, Cambridge, Massachusetts, United States of America, **4** IUCN/SSC-CI/CABS Biodiversity Assessment Unit, Conservation International, Arlington, Virginia, United States of America, **5** Department of Wildlife and Fisheries Sciences, Texas A&M University, College Station, Texas, United States of America, **6** Department of Environmental Sciences, University of Virginia, Charlottesville, Virginia, United States of America, **7** World Land Trust-US, Deerfield, New Hampshire, United States of America

Beta-diversity, the change in species composition between places, is a critical but poorly understood component of biological diversity. Patterns of beta-diversity provide information central to many ecological and evolutionary questions, as well as to conservation planning. Yet beta-diversity is rarely studied across large extents, and the degree of similarity of patterns among taxa at such scales remains untested. To our knowledge, this is the first broad-scale analysis of cross-taxon congruence in beta-diversity, and introduces a new method to map beta-diversity continuously across regions. Congruence between amphibian, bird, and mammal beta-diversity in the Western Hemisphere varies with both geographic location and spatial extent. We demonstrate that areas of high beta-diversity for the three taxa largely coincide, but areas of low beta-diversity exhibit little overlap. These findings suggest that similar processes lead to high levels of differentiation in amphibian, bird, and mammal assemblages, while the ecological and biogeographic factors influencing homogeneity in vertebrate assemblages vary. Knowledge of beta-diversity congruence can help formulate hypotheses about the mechanisms governing regional diversity patterns and should inform conservation, especially as threat from global climate change increases.

Citation: McKnight MW, White PS, McDonald RI, Lamoreux JF, Sechrest W, et al. (2007) Putting beta-diversity on the map: Broad-scale congruence and coincidence in the extremes. *PLoS Biol* 5(10): e272. doi:10.1371/journal.pbio.0050272

Introduction

Beta-diversity, the change in species composition between places, represents the differentiation component of diversity, as opposed to the inventory component, which describes the species composition of a single place [1–3]. Although beta-diversity was originally defined as the differentiation of communities along environmental gradients [1], the concept applies more widely to the phenomenon of species compositional change at any scale, regardless of mechanism [2–7]. Beta-diversity *sensu lato* is determined through a complex array of processes relating to the interaction of species traits (e.g., vagility and niche width) and characteristics of the physical landscape (e.g., environmental dissimilarity, topographic complexity, and isolation) over time [3,8–11]. Geographic variation in beta-diversity, from gradual changes to abrupt transitions, reflects past and present differences in environment, ecological interactions, and biogeographic history, including barriers to dispersal [4,7,9–15].

As beta-diversity quantifies the change, or turnover, in species across space, it is central to a wide array of ecological and evolutionary topics, such as the scaling of diversity [16–19], the delineation of biotic regions or biotic transitions [20,21], and the mechanisms through which regional biotas are formed [15,20–22]. Beta-diversity also provides information critical to conservation planning, which strives to represent all biodiversity within practical constraints such as area and cost [11,14,16,23,24]. While the total number of species, endemic

species, or threatened species often contributes to the relative importance of an area [25–29], it is the rate of species turnover between sites that dictates the optimal spatial arrangement of conservation areas [10,11,16]. Although the principles behind most approaches to systematic planning, such as complementarity, are driven by patterns of beta-diversity [23,30], few methods make explicit use of turnover measures [6,31]. Directly incorporating beta-diversity patterns into priority setting, however, benefits conservation efforts. For example, modeling compositional dissimilarity to develop surrogates for data-poor regions can improve biodiversity representation [6,30,32]. Moreover, including turnover estimates in area selection algorithms captures variation in species assemblages, which helps to preserve ecological and evolutionary processes as well as underlying environmental heterogeneity necessary for long-term persistence [28,31].

Despite the importance of beta-diversity, relatively little is known about diversity's “other component,” particularly at

Academic Editor: Georgina M. Mace, Imperial College London, United Kingdom

Received May 15, 2006; **Accepted** August 17, 2007; **Published** October 9, 2007

Copyright: © 2007 McKnight et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abbreviations: ess, estimated sample size

* To whom correspondence should be addressed. E-mail: meghan.mcknight@gmail.com

Author Summary

Beta-diversity—how species composition varies from place to place—is a fundamental attribute of biodiversity. However, despite its recognized importance, beta-diversity is rarely studied across large spatial scales. Here we use a new method to compare amphibian, bird, and mammal beta-diversity across large regions within the Western Hemisphere. We show that although the areas of low beta-diversity are different for the three groups, areas of high beta-diversity largely coincide. Moreover, we find that the degree to which the groups exhibit similar patterns of beta-diversity depends on the geographic location and extent at which it is measured. Beta-diversity is high where species are most susceptible to climate change, such as in areas with complex topography or high environmental variation. Identifying where areas of high beta-diversity coincide for different species groups is essential to the design of effective protected area networks.

broad scales. This is largely because measures of beta-diversity require knowledge of species identities rather than just species counts. Recent advances in species distributional data have made beta-diversity analyses possible at large extents [17,20,21], but these studies have been limited to single taxa. Cross-taxon congruence in beta-diversity has been tested only at small scales, with varying methods and results [14,22,32,33], in contrast to the wide range of scales at which concordance in both species richness and endemism has been studied [34–38]. Here, we present what is to our knowledge the first analysis of beta-diversity congruence across large spatial scales, based on distributional data for three groups of terrestrial vertebrates in the continental Western Hemisphere.

Beta-diversity of amphibians ($n = 2,174$) [39], breeding birds ($n = 3,882$) [40], and mammals ($n = 1,611$) [41] was estimated as a function of the distance decay of similarity—the decrease in compositional similarity with increasing geographic distance between sites [4,7,10,14]. We modeled distance decay from each 100 km × 100 km grid cell, and used these models to calculate our measure of beta-diversity, $\beta_{\text{sim-d}}$: the estimated proportional turnover in species composition at a distance of 100 km (see Materials and Methods). This individual-cell-based technique accounts for the considerable geographic variation in the rate at which similarity decays, and can be used to produce a continuous layer of compositional change similar to past grid-based neighborhood analyses of broad-scale beta-diversity (e.g., [18,20,21]). Considering comparisons over a range of distances reduces possible bias in similarity levels that could arise from the differences in centroid to centroid distance and in shared perimeter length that occur between orthogonal and diagonal neighbors of a rectangular grid. The smoothing that results from the distance decay regressions also limits the influence of artifacts due to small-scale errors in range map boundary placement.

Our approach to quantifying the distance decay relationship makes several improvements to methods used in previous studies [4,7,10,14]. For instance, we modeled distance decay using logistic regression, which has advantages over linear or log-linear ordinary least-square regressions [4,7,14], particularly for proportional data [42]. Furthermore, following Lennon et al. [18], we measured similarity with a metric shown to be independent of differences in species richness between grid cells in order to isolate change due to species replacement [43] (see Materials and Methods).

We tested congruence in $\beta_{\text{sim-d}}$ for the three taxa using two different approaches. With the first, we measured congruence in overall $\beta_{\text{sim-d}}$ patterns and examined whether congruence levels were consistent across multiple spatial extents and among different geographic locations. In the second approach, we quantified spatial overlap in the extremes of $\beta_{\text{sim-d}}$. We report that the strength of congruence depends on the location and extent at which it is measured, and that overlap in high $\beta_{\text{sim-d}}$ is much greater than in low $\beta_{\text{sim-d}}$. Furthermore, the pairs of taxa varied substantially in level of congruence and degree of overlap.

Results/Discussion

$\beta_{\text{sim-d}}$ Patterns

Amphibian, bird, and mammal $\beta_{\text{sim-d}}$ mapped at this scale (Figure 1) provide a striking contrast to well-known patterns of broad-scale species richness for these vertebrate groups. Whereas high richness is generally concentrated in the tropics and decreases towards both poles [44], $\beta_{\text{sim-d}}$ of all levels is found across a wide range of latitudes. High $\beta_{\text{sim-d}}$ stretches along the mountainous Pacific edge of the continents, while low $\beta_{\text{sim-d}}$ is found within more environmentally uniform portions of northern South America and boreal North America. Accordingly, $\beta_{\text{sim-d}}$ has a positive relationship with both elevation and number of biome boundaries ($\beta_{\text{sim-d}}$ and elevation: Spearman rank $\rho = 0.219\text{--}0.427$, $p < 0.05$ for amphibian $\beta_{\text{sim-d}}$, $p < 0.001$ for other taxa; $\beta_{\text{sim-d}}$ and biome edge: $\rho = 0.295\text{--}0.320$, $p < 0.001$ for all; Table S1; see Materials and Methods). Although the variables show considerable spread (Figure S1), high $\beta_{\text{sim-d}}$ grid cells of all three groups occur at significantly higher elevations and on a greater number of biome edges than expected by chance alone, while low $\beta_{\text{sim-d}}$ grid cells have significantly lower elevations and fewer biome edges than expected by chance (Table S2; 10,000 random sets, $p < 0.05$ for elevation in amphibian low $\beta_{\text{sim-d}}$ grid cells, $p < 0.001$ for all others; see Materials and Methods). The weaker significance for elevation in amphibian low $\beta_{\text{sim-d}}$ grid cells is likely due to the wood frog (*Rana sylvatica*) being the only amphibian species to occur throughout much of the boreal region, including high-altitude areas such as the Alaska panhandle [45]. This amphibian homogeneity differs greatly from the high $\beta_{\text{sim-d}}$ of birds at northern latitudes, which captures the presence of a strong Holarctic element in the avifauna along the arctic coast [46]. Such differences in $\beta_{\text{sim-d}}$ reveal the individual biogeographic histories of the taxa and may arise from variation in dispersal ability, particularly in relation to historical factors such as glaciation and faunal interchange [45,47]. For instance, the elevated mammal $\beta_{\text{sim-d}}$ in South America's southern cone reflects a transition in the region's diverse mammal lineages, notably the radiation of narrowly ranging hystricognath rodents [48], while the high amphibian $\beta_{\text{sim-d}}$ of the southern Appalachian Mountains results from the diversification of salamanders within this area's stable, moist environments [45].

Congruence in Overall $\beta_{\text{sim-d}}$ Patterns

Pair-wise correlations of amphibian, bird, and mammal $\beta_{\text{sim-d}}$ across the Western Hemisphere were positive and significant ($\rho = 0.340\text{--}0.553$, $p < 0.001$ for all; see Materials and Methods) (Table 1; Figure 2). When measured at the extent of a single biogeographic realm, however, we found

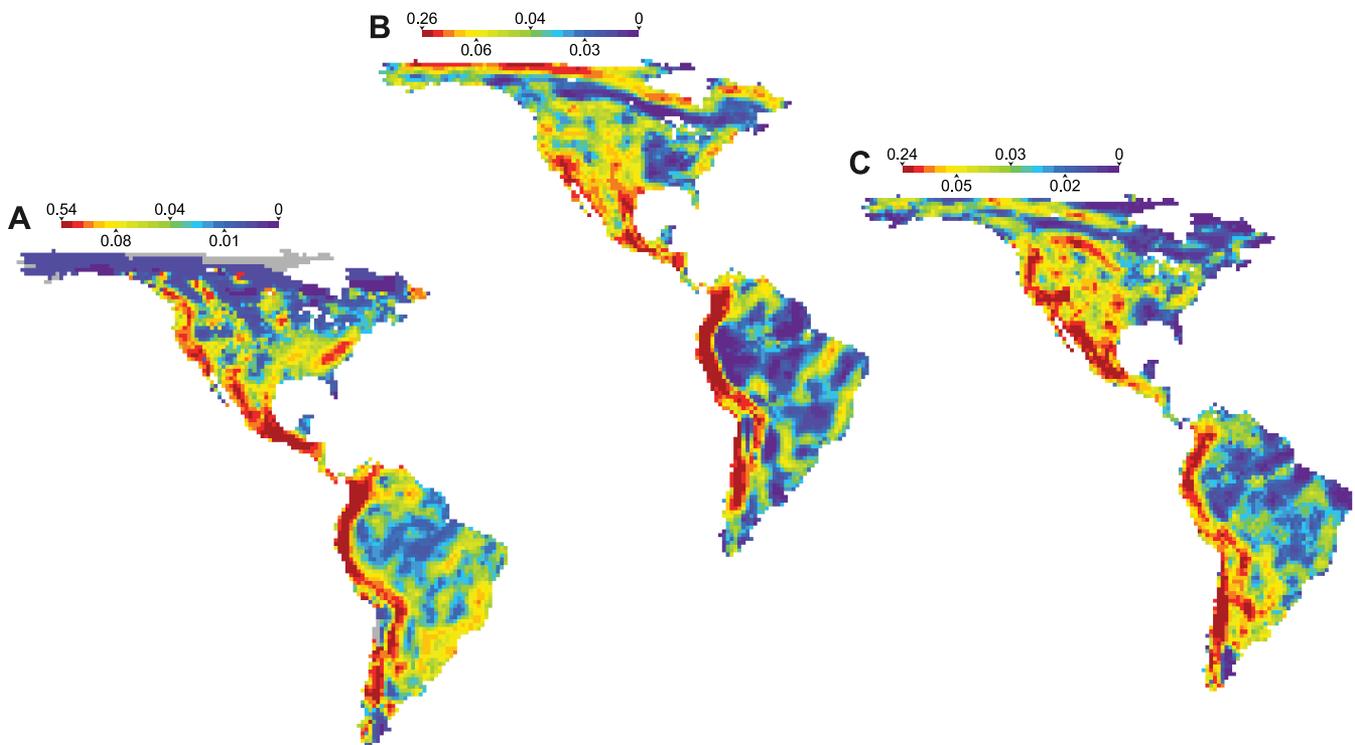


Figure 1. Beta-Diversity of Amphibians, Birds, and Mammals Mapped Continuously across the Continental Western Hemisphere

Beta-diversity (β_{sim-d}) values for each taxon are divided into 20 quantiles, represented by warm (higher β_{sim-d}) to cool (lower β_{sim-d}) colors. The scale accompanying the color ramp for each taxon shows minimum, first quartile, median, third quartile, and maximum values of β_{sim-d} . Gray grid cells do not contain amphibian species. (A) Amphibians. (B) Birds. (C) Mammals.
doi:10.1371/journal.pbio.0050272.g001

that pair-wise congruence was greater within the Neotropics ($\rho = 0.636\text{--}0.695$, $p < 0.001$ for all) than at the hemisphere extent, but was comparatively weak within the Nearctic (amphibians and mammals: $\rho = 0.390$, $p < 0.05$; birds and mammals: $\rho = 0.405$, $p < 0.001$) or even lacking (amphibians and birds: $\rho = 0.032$, not significant) (Table 1; Figure 2). The disparity in congruence strength between the realms indicates that congruence measured across large regions can hide incongruities that manifest at reduced spatial extents [35,36].

To examine congruence at even smaller extents, we used a moving-window algorithm that calculated the correlation in β_{sim-d} between each pair of taxa within a 350-km radius of each grid cell (see Materials and Methods). Composite maps of the resulting correlation coefficients for the pairs revealed considerable geographic variation in congruence (Figure 3). Although the majority of correlations were strongly positive,

others were weak or strongly negative. The latter were most apparent in the Nearctic realm for correlations with amphibians. Understanding the dependence of diversity relationships on observational scale is of pressing concern for ecology, biogeography, and conservation planning [10,18,23,36]. Our analyses demonstrate that both the geographic location and the spatial extent of analysis affect the level of congruence observed in β_{sim-d} , and emphasize the need for tests across multiple scales and regions in order to make objective comparisons among ecological studies.

Spatial Overlap in High and Low β_{sim-d}

Correlations across all grid cells do not necessarily indicate the level of cross-taxon spatial coincidence in areas of highest or lowest β_{sim-d} —a more useful measure for conservation planning and biogeographic delineation [34,35,49]. Congruence in the extremes of diversity is frequently measured as

Table 1. Correlations in Beta-Diversity (β_{sim-d}) within the Western Hemisphere, Nearctic Realm, and Neotropical Realm

Extent	Amphibians and Birds			Amphibians and Mammals			Birds and Mammals		
	ρ	<i>n</i>	ess	ρ	<i>n</i>	ess	ρ	<i>n</i>	ess
Western Hemisphere	0.340**	3,693	224.88	0.499**	3,693	104.68	0.553**	3,821	117.48
Nearctic realm	0.032	1,744	122.68	0.390*	1,744	43.81	0.405**	1,862	68.94
Neotropical realm	0.695**	1,878	86.44	0.636**	1,878	51.55	0.662**	1,888	52.44

Spearman rank correlation coefficients (ρ), number of grid cells (*n*), and corrected sample size (ess) for each pair-wise comparison are shown.

*, $p < 0.05$; **, $p < 0.001$.

doi:10.1371/journal.pbio.0050272.t001

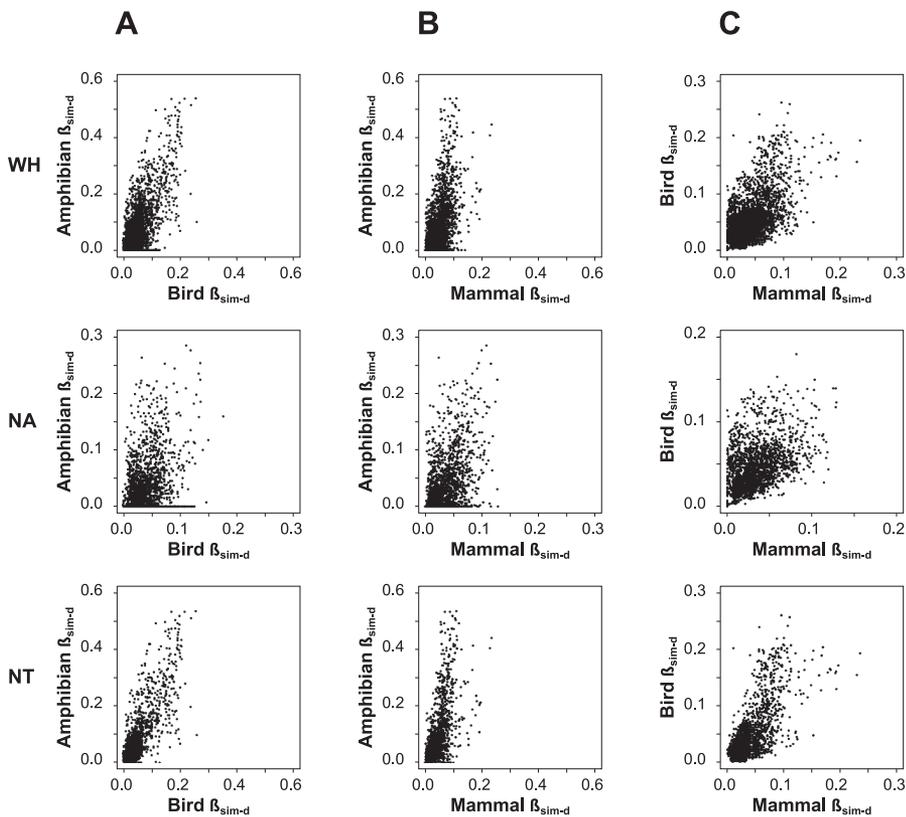


Figure 2. Cross-Taxon Relationships in Beta-Diversity of Amphibians, Birds, and Mammals

Scatter plots show the relationships between $\beta_{\text{sim-d}}$ for each pair of taxa within the Western Hemisphere (WH, top row), the Nearctic realm (NA, middle row), and the Neotropical realm (NT, bottom row). The axes for each plot are scaled according to the maximum $\beta_{\text{sim-d}}$ value of the two taxa within the extent specified. Note that maximum values are much greater for amphibians than for either birds or mammals and that all three taxa reach higher rates of assemblage change in the Neotropics than in the Nearctic. (A) Amphibians and birds. (B) Amphibians and mammals. (C) Birds and mammals. doi:10.1371/journal.pbio.0050272.g002

the degree of overlap in matching percentage sets of two groups [34,50]. We evaluated high and low $\beta_{\text{sim-d}}$ congruence for the pairs of taxa and between all three groups as the proportion of maximum possible overlap [34] in matching percentage sets of the highest 2.5% and lowest 2.5% of each taxon's $\beta_{\text{sim-d}}$ grid cells (see Materials and Methods).

Spatial coincidence in high $\beta_{\text{sim-d}}$ was greatest between amphibians and birds (51.6%). These taxa showed lower, but similar levels of overlap in high $\beta_{\text{sim-d}}$ with mammals (21.5% and 29.2%, respectively), and coincidence between all three groups was minimal (15.1%). Grid cells with overlapping high $\beta_{\text{sim-d}}$ primarily occurred in the northern and southern Andes (Figure 4), consistent with the former as a center of endemism for all three taxa and with the extreme climatic gradient within the latter [44,45]. A substantial proportion of grid cells were found only in the high $\beta_{\text{sim-d}}$ percentage sets of one taxon. For example, 41.9% of amphibian high $\beta_{\text{sim-d}}$ grid cells were unique, as were 35.4% of bird high $\beta_{\text{sim-d}}$ grid cells and 64.6% of mammal high $\beta_{\text{sim-d}}$ grid cells. The distribution of these grid cells reflects the specific biogeographies of each taxon. Whereas unique grid cells were predominantly located in the northern Andes for birds and in the Central American highlands for amphibians, unique mammal grid cells were largely outside the tropics (Figure 4).

There was comparatively little spatial coincidence in the lowest 2.5% of $\beta_{\text{sim-d}}$. Low $\beta_{\text{sim-d}}$ of birds and mammals showed the most overlap, at only 11.5%. Coincidence was

negligible for the other two pairs of taxa (amphibians and mammals, 5.4%; amphibians and birds, 2.2%), and there was no overlap among all three groups. Accordingly, the majority of grid cells in the low $\beta_{\text{sim-d}}$ percentage sets were restricted to one taxon (83.3%–92.5%). These grid cells were located mainly in the boreal and arctic regions of the Nearctic realm for amphibians and mammals, respectively (Figure 4). Conversely, most unique bird grid cells occurred in the Neotropics within several biomes, including a substantial number in the Amazon Basin (Figure 4).

The degree of overlap in matching percentage sets, however, does not provide a complete picture of spatial coincidence in the extremes of $\beta_{\text{sim-d}}$. In fact, the majority of highest $\beta_{\text{sim-d}}$ grid cells for all three taxa actually had relatively high levels of $\beta_{\text{sim-d}}$ for the other groups (Figure 5), indicating that areas of high beta-diversity largely coincide. On average, more than two-thirds of grid cells in the highest 2.5% of one taxon's $\beta_{\text{sim-d}}$ grid cells were also in the highest 10% of $\beta_{\text{sim-d}}$ for the other taxa ($70.0\% \pm 8.7\%$, range = 61.5%–81.7%). This was not true for low $\beta_{\text{sim-d}}$. Low $\beta_{\text{sim-d}}$ grid cell sets exhibited greater variation in $\beta_{\text{sim-d}}$ values for the other taxa than did the high $\beta_{\text{sim-d}}$ sets. Moreover, less than one-quarter of the lowest 2.5% of one taxon's $\beta_{\text{sim-d}}$ grid cells were in the lowest 10% of $\beta_{\text{sim-d}}$ for the other taxa ($21.9\% \pm 14.6\%$, range = 2.9%–40.6%)—further evidence that areas of low $\beta_{\text{sim-d}}$ are spatially distinct (Figure 5).

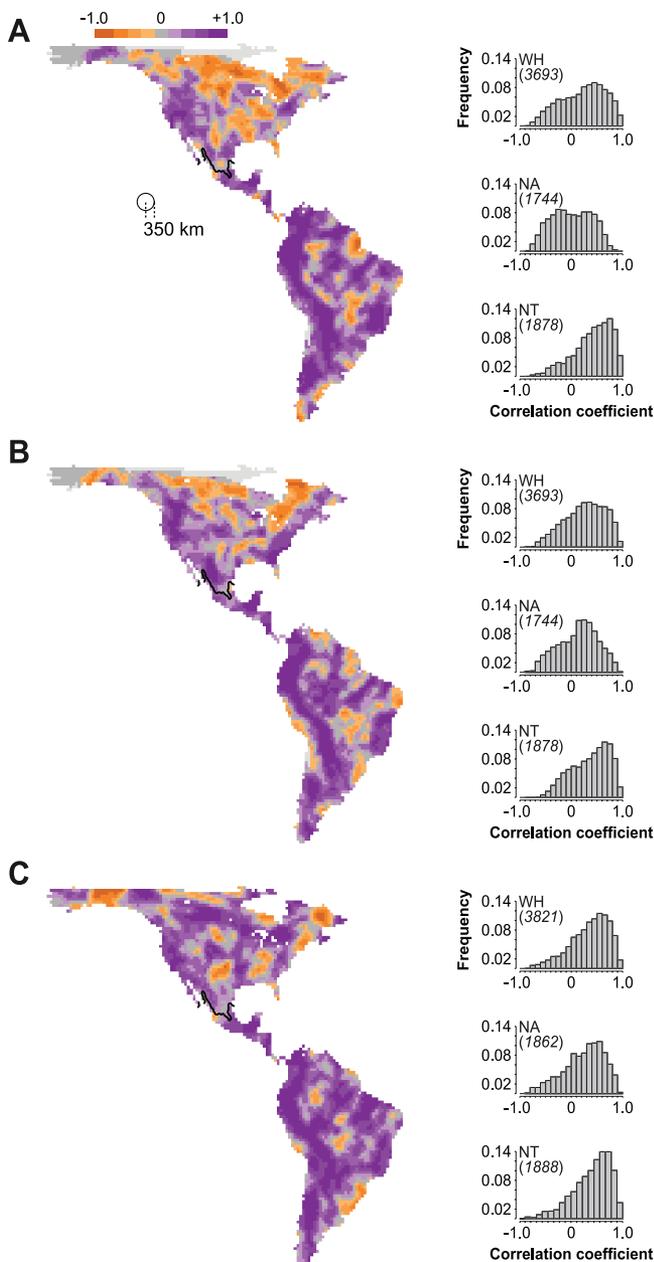


Figure 3. Geographic Variation in Beta-Diversity Congruence of Amphibians, Birds, and Mammals at Small Spatial Extents

The color of each grid cell indicates the strength of beta-diversity (β_{sim-d}) congruence calculated within a 350-km-radius window around that grid cell. Orange shades represent strong (darkest) to weak (lightest) negative correlations. Purple shades show strong (darkest) to weak (lightest) positive correlations. Dark gray indicates very weak correlations of either sign, or no correlation. Light gray grid cells do not contain amphibian species. Shown to the right of each map are frequency distributions of correlation coefficients for windows located within the entire Western Hemisphere (WH), the Nearctic realm (NA), and the Neotropical realm (NT), which are consistent with the overall level of congruence measured at these extents. The black line marks the boundary between the two realms. (A) Amphibians and birds. (B) Amphibians and mammals. (C) Birds and mammals.
doi:10.1371/journal.pbio.0050272.g003

Conclusions

Congruence in beta-diversity of three groups of terrestrial vertebrates is highly dependent on the geographic location and extent of analysis, reflecting taxonomic and regional variation

in the influence of large-scale historical processes and environmental factors [4,7,10,14,15]. Our results show that although correlations in amphibian, bird, and mammal β_{sim-d} measured at small extents vary in strength throughout the Western Hemisphere, congruence is generally stronger within the Neotropical realm than within the Nearctic. This difference may be part of a broader asymmetry in biodiversity patterns between the Northern Hemisphere and the Southern Hemisphere [51,52]. The weak pair-wise correlations within the Nearctic realm, as well as the minimal overlap in both high and low β_{sim-d} , could result from differing responses of amphibians, birds, and mammals to the realm's climatic and geologic history [45,47]. In contrast, the comparatively strong β_{sim-d} congruence in the Neotropics is indicative of common patterns of speciation and extinction histories. This is particularly apparent within the Neotropical mountains, where the substantial overlap in high β_{sim-d} among the three groups underscores the importance of this region in generating diversity. Variation in β_{sim-d} congruence also has implications for conservation, because the efficacy of conservation surrogates and efforts to model overall biodiversity distribution depend on taxa having concordant patterns of compositional change [30]. Our results largely support these approaches, but it is important to recognize limitations that may arise from differing congruence levels among biogeographic realms.

Regions of rapid species turnover require increased attention to the placement and size of conservation areas in order to protect biodiversity. Spatial coincidence in areas of high β_{sim-d} is therefore encouraging, as successful conservation strategies in these places may be resource intensive. Conservation planning, of course, must occur across hierarchical scales in order to ensure adequate representation [23,28]. Broad-scale analyses of β_{sim-d} highlight regions where protected areas should be closely spaced to effectively conserve biodiversity; however, the optimal configuration for conservation networks will depend on finer-scale beta-diversity patterns [53]. Mapping broad-scale β_{sim-d} can also identify areas where species face increasing threat to persistence. For example, because β_{sim-d} is high where species' ranges are particularly susceptible to climatic variability, such as at steep environmental gradients and centers of endemism [54–56], or at biome transitions where range shifts are most noticeable [54,55], we suggest that areas of high β_{sim-d} are likely to be especially vulnerable to climate change.

The unique biogeography of the Western Hemisphere—the great variation in the effects of Pleistocene glaciation, the complex of mountain chains along much of the western coast, and the relative isolation of the continents—has played a major role in shaping the distribution and evolution of biodiversity. More work is needed to determine if our findings will extend to other parts of the world with different geologic and environmental histories. Furthermore, the relative contribution of historical factors and current ecological interactions in determining beta-diversity patterns and congruence in beta-diversity across taxa is an important area of inquiry.

Our results describe patterns of species turnover at a 100 km \times 100 km resolution. As comprehensive finer-resolution data become available, further analyses will confirm whether the levels of beta-diversity and congruence we found are consistent with those measured at smaller grain sizes. Future research is also needed to ascertain the degree to which our results can be generalized to other taxa, especially

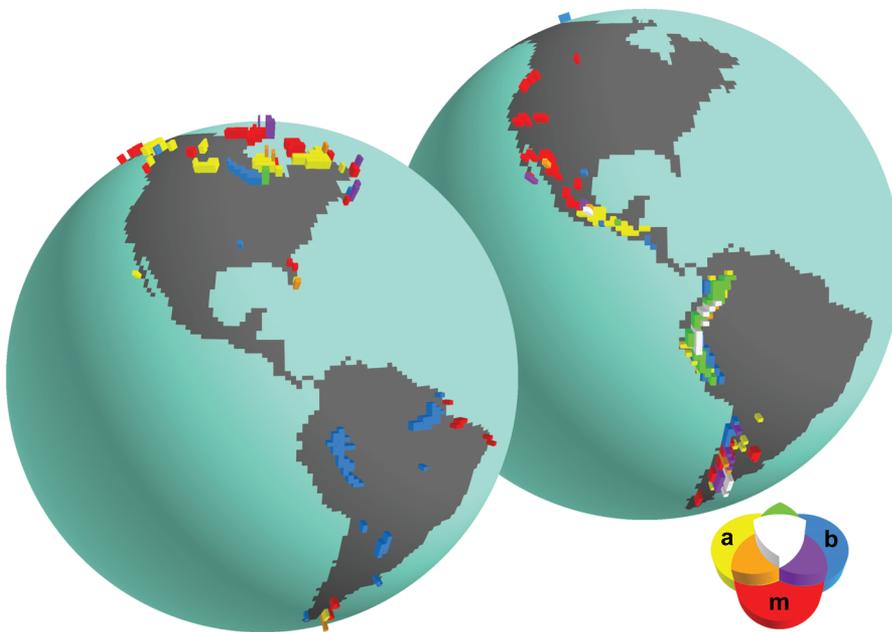


Figure 4. Geographic Distribution of Amphibian, Bird, and Mammal High and Low Beta-Diversity Overlap

Spatial overlap in beta-diversity (β_{sim-d}) for percentage sets of each taxon's lowest (left) and highest (right) 2.5% of β_{sim-d} grid cells is shown. Primary colors indicate grid cells unique to one taxon (yellow, amphibians; blue, birds; red, mammals), secondary colors indicate overlap between two groups, and white indicates overlap of all three groups. The height of the grid cells reflects the number of overlapping groups. Note the greater degree of spatial coincidence in high β_{sim-d} than in low β_{sim-d} . doi:10.1371/journal.pbio.0050272.g004

more distantly related groups or those that show large variation in dispersal ability. For instance, taxa with poor dispersal and low rates of gene flow are apt to exhibit higher beta-diversity than those groups that have high dispersal and high rates of gene flow. However, we believe that some of our findings, such as the strong relationship between topography and beta-diversity congruence, will prove true for most taxa.

Materials and Methods

Data. Analyses were based on range data for extant species of amphibians ($n=2,174$), breeding birds ($n=3,882$), and mammals ($n=1,611$) in the Western Hemisphere [39–41]. The range maps used for this study were obtained as digital vector files (ArcView format) from the Web sites indicated by [39–41], where one can also find information on updates, detailed descriptions of the production process, and complete lists of

sources. Note that these datasets are periodically updated, and the files used for these analyses may differ from the most recent versions available from [39–41]. We confined our analyses to terrestrial breeding birds, and we provide a map of bird β_{sim-d} based on both breeding and non-breeding ranges of all terrestrial birds ($n = 3,890$) for comparison. β_{sim-d} for all birds (Figure S2) was highly correlated with β_{sim-d} for breeding birds (Figure 1) ($\rho = 0.954$, estimated sample size [ess] = 249.12, $p < 0.001$). The number of species in these vertebrate groups is not static, as new species, especially of amphibians, continue to be discovered [57]. However, the areas from which species are most often described tend to be the same and will likely accentuate the patterns we present [58]. In relation to this point, systematic bias in the data may result from differences in sampling efforts, as the distributions of certain groups (e.g., birds) or geographic areas (e.g., temperate regions) for which sampling efforts have been intense will be more reliable than those that are undersampled (e.g., amphibians or tropical regions). As a precaution against such bias, we excluded from the analyses the 630 amphibian

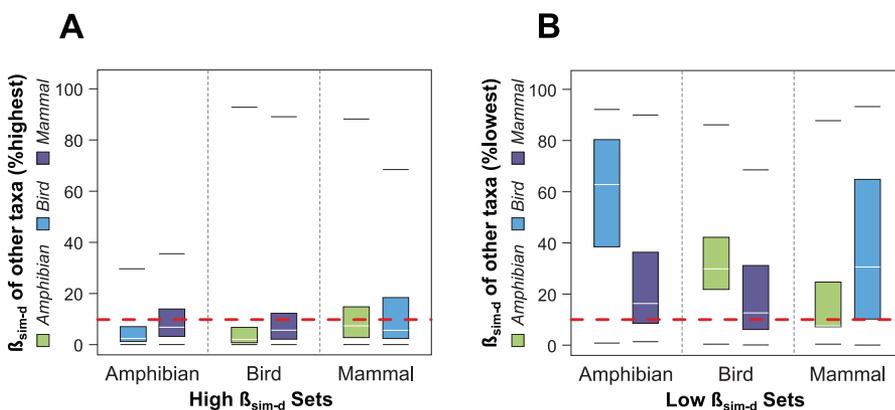


Figure 5. Levels of Beta-Diversity for Vertebrate Taxa within Areas of High and Low Beta-Diversity of Amphibians, Birds, and Mammals

Percentage sets of the highest (A) and lowest (B) 2.5% of beta-diversity (β_{sim-d}) grid cells for one taxon (x -axis) contain a range of β_{sim-d} levels for the other taxa (y -axis), as shown by the box plots (white lines within boxes indicate the median; top and bottom box edges indicate first and third quartiles; black lines indicate minimum and maximum percentage rank of β_{sim-d}). The red dashed line indicates the highest or lowest 10% of β_{sim-d} . doi:10.1371/journal.pbio.0050272.g005

species with an IUCN Red List category of “data deficient” (<http://www.redlist.org/>) because of the unreliability of their range maps. The exclusion of these species did not substantially affect our results (correlation between amphibian $\beta_{\text{sim-d}}$ using all mapped species and amphibian $\beta_{\text{sim-d}}$ excluding “data deficient” species; $\rho = 0.993$, $\text{ess} = 158.6$, $p < 0.001$).

We recorded the presence/absence of each species in 100 km \times 100 km equal-area grid cells, roughly equivalent to $1^\circ \times 1^\circ$ at the equator (Behrmann projection, WGS84 datum); a species was considered present if any portion of its range (exclusive of polygons coded as introduced, migratory, or vagrant) occurred within the continental land area of the grid cell. However, the range maps used approximate the extent of occurrence of a species, rather than its area of occupancy, and therefore the species may not be found in every grid cell that falls within the mapped range [59,60]. Such biases are inherent in range data compiled across large regions, and remind us that while the patterns they show inform us about species turnover at broad scales, they are not a replacement for finer-scale distributional information.

Grid cells on the perimeter of the continents vary considerably in the amount of land they contain, particularly those along the narrow Isthmus of Panama. To avoid potential effects of species-area relationships or errors from range map boundary placement, only grid cells containing $\geq 40\%$ of continental land were included in the analyses (grid cells: $n = 3,693$ for amphibians; $n = 3,821$ for birds and mammals). Estimates of $\beta_{\text{sim-d}}$ using this cutoff were not appreciably different from those based on a more conservative cutoff of 75% land area, but allowed for the inclusion of additional species. Grid cells were classified as either Nearctic ($n = 1,744$ for amphibians; $n = 1,862$ for birds and mammals), Neotropical ($n = 1,878$, amphibians; $n = 1,888$, birds and mammals), or transitional between the two biogeographic realms ($n = 71$ for all taxa) [61]. Transitional grid cells were not included in analyses at the realm extent.

Analyses. We used a moving-window algorithm to model the distance decay of similarity from each individual grid cell, and used the resulting regression parameters to calculate a value of beta-diversity, $\beta_{\text{sim-d}}$, as the estimated proportional turnover from that grid cell at a distance of 100 km. Considering comparisons between grid cells over a range of distances helps alleviate concerns typical of gridded nearest-neighbor analyses of large-scale species distributions. For example, artifacts may arise from the small-scale errors that can occur in range boundary placement when converting polygon maps into gridded data, as well as from the discrepancy in centroid to centroid distance and shared perimeter length between orthogonal and diagonal neighbors in a rectangular grid.

Similarity (S) between two grid cells was calculated as the complement of β_{sim} , a dissimilarity metric that isolates change due to species replacement from differences in species richness:

$$\beta_{\text{sim}} = \frac{\min(b, c)}{a + \min(b, c)},$$

where a is the number of species shared, b is the number of species found only in the second grid cell, and c is the number of species found only in the first grid cell, making $\min(b, c)$ the number of unshared species in the more depauperate grid cell [18,43]. Therefore, as the complement of β_{sim} (i.e., $1 - \beta_{\text{sim}}$),

$$S = 1 - \beta_{\text{sim}} = \frac{a}{a + \min(b, c)},$$

or the proportion of species in the more depauperate grid cell that also occur in the other grid cell.

Note that $S/1 - S$ is a transformation of the ratio of shared species to unshared species in the more depauperate grid cell, or $a/\min(b, c)$. This enables us to model distance decay using a logistic regression functional form defined such that

$$\ln\left(\frac{a}{\min(b, c)}\right) = I + r \times \ln(d),$$

where d is the centroid to centroid distance, and I and r are fitted intercept and slope coefficients. This functional form has an advantage over linear and log-linear forms, in that \hat{S} is bound between zero and one, as is appropriate for a similarity index. The observed data are counts, and we use a binomial error distribution for our regression. This has several advantages over linear and log-linear regressions with a normal error distribution, resulting in a better empirical fit than other techniques [42]. First, the special cases where $S=0$ (i.e., $a=0$) and $S=1$ (i.e., $\min(b, c)=0$) do not cause problems in the estimation process, as these are valid possibilities under the binomial distribution. Second, the binomial error distribution accounts for the greater variance in $a/\min(b, c)$ (and hence S) at low species numbers.

The distance decay regression at each grid window was built using between-grid-cell comparisons of the focal grid cell and all grid cells within a ≤ 500 -km centroid to centroid radius. Thus, unlike most published distance decay regressions, which compute a rate of change based on comparisons between all samples within a region, our regressions are based on comparisons only with the focal grid cell and therefore reflect the change from a particular point (i.e., grid cell). The arbitrary distance of 500 km was chosen after experimenting with several other maximum distances (350, 1,000, 1,500, 2,000, and 3,000 km) because it provided a sufficient total number of between-grid comparisons (i.e., sample size), spread over a range of distances, to ensure a robust distance decay relationship, but did not result in an over-smoothed beta-diversity surface, as occurred with greater maximum distances (as judged by visual comparisons of the maps). By transforming the above defined regression equation as

$$1 - \hat{S}_d = \frac{1}{e^{I+r \times \ln(d)} + 1},$$

it is possible to use any set of distance decay regression coefficients (I and r) to predict the proportional dissimilarity at any distance (d). Thus, we used the coefficients from the distance decay regression for each grid cell to estimate our measure of beta-diversity, $\beta_{\text{sim-d}}$, as $(1 - \hat{S}_d)$ for $d = 100$ km, or the estimated proportional turnover in composition from that grid cell at a distance of 100 km.

The predicted degree of dissimilarity at a given distance, $\beta_{\text{sim-d}}$, differs from the average observed dissimilarity (β_{sim}) at the same distance because it accounts for the rate at which dissimilarity changes with increasing distance (i.e., the effect of extent). At the same time, $\beta_{\text{sim-d}}$ differs from the rate of distance decay in the following ways. (i) Estimates of $\beta_{\text{sim-d}}$ depend on both the intercept and slope parameters of the distance decay relationship. The former, as initial similarity level, reflects dissimilarity at near distances and the latter, as the rate of distance decay, captures dependency of dissimilarity on extent [62,63]. (ii) $\beta_{\text{sim-d}}$ is the estimated dissimilarity at a specified distance (i.e., 100 km)—predictions at other distances (e.g., 50 or 300 km) would result in different values, reflecting the effect of spatial extent on compositional change. Turnover at this distance, which is the minimum distance between adjacent grid cells, is more intuitive than that between distant grid cells for discussion and graphical representation of beta-diversity as a continuous surface, and makes it easier to compare our results to other broad-scale diversity analyses.

Although the number of grid cells included in a regression model decreased with increasing proximity to the coast (including major interior water bodies), graphical examination of scatter plots and the resulting maps showed that coastal effects were negligible for amphibians and mammals and varied geographically for birds. The elevated bird $\beta_{\text{sim-d}}$ on some coastal sections likely has a biological rather than methodological basis [18]. It is important to remember that $\beta_{\text{sim-d}}$ quantifies change in species composition between 100 km \times 100 km grid cells, and therefore does not reflect the level of heterogeneity within a grid cell. Furthermore, $\beta_{\text{sim-d}}$ is a measure of proportional species turnover and does not represent the absolute number of species gained or lost between grid cells. Lastly, while the smooth surface that results from modeling the effect of distance on similarity reduces the effect of potential errors in gridded large-scale range data, extremely abrupt transitions may be attenuated. However, the major patterns found for $\beta_{\text{sim-d}}$ were also apparent in maps of average nearest-neighbor beta-diversity (the average dissimilarity [β_{sim}] of a focal grid cell and its orthogonal and diagonal neighbors) (Figure S3). Further, a comparison of Table 1 with pair-wise correlations of average β_{sim} (Table S3) shows that the congruence levels we report are not artifacts of the smoothing process.

We tested whether grid cells containing high $\beta_{\text{sim-d}}$ or those with low $\beta_{\text{sim-d}}$ differed significantly in elevation or were found on a greater number of biome edges than could be expected by chance [64]. To do this, we selected sets of grid cells containing the highest 2.5% and the lowest 2.5% of $\beta_{\text{sim-d}}$ values for each taxon (2.5% = 93 grid cells for amphibians, 96 grid cells for birds and mammals), and calculated the mean elevation and mean number of biome edges for each set. We then compared these values to distributions of values for the mean elevation and mean number of biome edges, respectively, calculated for 10,000 sets of randomly selected grid cells (grid cells per random set: $n = 93$ for amphibians; $n = 96$ for birds and mammals). For each comparison, we computed a one-tailed p -value by counting the number of values in the random distribution greater than or equal to the value of a high $\beta_{\text{sim-d}}$ set—or less than or equal to the value of a low $\beta_{\text{sim-d}}$ set. Elevation was measured as the mean elevation within a grid cell from a digital elevation model of approximately 1 km \times 1 km resolution (the Global 30 Arc Second Elevation Data Set, [PLoS Biology | www.plosbiology.org](http://www1.</p>
</div>
<div data-bbox=)

gsi.go.jp/geowww/globalmap-gsi/gtopo30/gtopo30.html). Following van Rensburg et al. [49], we considered a grid cell to be on a biome edge if a biome (as delineated by Olson et al. [61]) covering $\geq 5\%$ of that grid cell also covered $< 5\%$ of any of the neighboring grid cells. The number of biome edges was then calculated as the number of biomes in that grid cell meeting this definition.

To evaluate the overall relationships between $\beta_{\text{sim-d}}$ and elevation and between $\beta_{\text{sim-d}}$ and number of biome boundaries within a grid cell, we calculated the correlation between $\beta_{\text{sim-d}}$ for the three taxa and each environmental variable. Correlations were calculated with Spearman rank correlation coefficients to accommodate the non-normal distributions of $\beta_{\text{sim-d}}$. Standard significance tests are not appropriate for autocorrelated data because the assumption of independence is violated; therefore, we tested for significance using a method developed by Clifford et al. [65] that corrects the sample size of two variables based on the level of the spatial dependency in and between them [18]. We calculated the ess for each pair of variables using the PASSAGE software package [66], and then used the corrected degrees of freedom to test the significance of each correlation.

Pair-wise congruence at the hemisphere and biogeographic realm extents was measured as the correlation in $\beta_{\text{sim-d}}$ values for each pair of taxa, and significance was tested using the method described above. To examine congruence at extents smaller than a biogeographic realm, we calculated the correlation in $\beta_{\text{sim-d}}$ values within a ≤ 350 -km-radius window (centroid to centroid distance) around each grid cell. We used this window size because it provided a better representation of the geographic variation in $\beta_{\text{sim-d}}$ at small extents than the other window sizes we experimented with (radii of 150, 250, and 450 km). The same overall pattern was also apparent using larger windows but became increasingly muted as the extent widened. Moreover, larger windows had a greater discrepancy in the number of grid cells occurring within windows around coastal versus inland grid cells, while smaller windows considerably decreased the number of grid cells across which congruence was measured. The ≤ 350 -km window was not substantially affected by either of these issues, and differences that did exist in the number of grid cells within coastal and interior windows did not appear to influence the geographical variation in congruence.

Spatial overlap between matching percentage sets of the highest 2.5% and lowest 2.5% of $\beta_{\text{sim-d}}$ grid cells for each pair of taxa and for all three groups was calculated as the maximum overlap possible [34]: N_c/N_t , where N_c is the number of grid cells common to the sets and N_t is the total number of grid cells in the smallest set (amphibians have slightly fewer grid cells than birds or mammals).

Supporting Information

Figure S1. Scatter Plots Showing Relationships between Beta-Diversity and Two Environmental Variables (Elevation and Number of Biome Edges within Grid Cells)

For each panel, untransformed (left plots) and transformed (right plots) values of $\beta_{\text{sim-d}}$ (y-axis) against either grid cell elevation (x-axis, upper plots) or number of biome edges within grid cell (x-axis, lower plots). In each plot, the red dots represent the highest 2.5% of $\beta_{\text{sim-d}}$ grid cells, and the purple dots show the lowest 2.5% of $\beta_{\text{sim-d}}$ grid cells. (A) Amphibians. (B) Birds. (C) Mammals.

Found at doi:10.1371/journal.pbio.0050272.sg001 (472 KB PDF).

References

- Whittaker RH (1960) Vegetation of the Siskiyou Mountains, Oregon and California. *Ecol Monogr* 30: 279–338.
- Whittaker RH (1972) Evolution and measurement of species diversity. *Taxon* 21: 213–251.
- Harrison S, Ross SJ, Lawton JH (1992) Beta-diversity on geographic gradients in Britain. *J Anim Ecol* 61: 151–158.
- Condit R, Pitman N, Leigh EG Jr, Chave J, Terborgh J, et al. (2002) Beta-diversity in tropical forest trees. *Science* 295: 666–669.
- Koleff P, Lennon JJ, Gaston KJ (2003) Are there latitudinal gradients in species turnover? *Global Ecol Biogeogr* 12: 483–498.
- Ferrier S, Powell GVN, Richardson KS, Manion G, Overton JM, et al. (2004) Mapping more of terrestrial biodiversity for global conservation assessment. *BioScience* 54: 1101–1109.
- Qian H, Ricklefs RE, White PS (2005) Beta diversity of angiosperms in temperate floras of eastern Asia and eastern North America. *Ecol Lett* 8: 15–22.
- Shmida A, Wilson MV (1985) Biological determinants of species diversity. *J Biogeogr* 12: 1–20.
- Cody ML (1986) Diversity, rarity, and conservation in Mediterranean-climate ecosystems. In: Soulé ME, editor. *Conservation biology: The science of scarcity and diversity*. Sunderland (Massachusetts): Sinauer Associates. pp. 122–152.

Figure S2. Bird Beta-Diversity Based on Both Breeding and Non-Breeding Ranges

Beta-diversity ($\beta_{\text{sim-d}}$) values are divided into 20 quantiles, represented by warm (higher $\beta_{\text{sim-d}}$) to cool (lower $\beta_{\text{sim-d}}$) colors. The scale accompanying the color ramp shows minimum, first quartile, median, third quartile, and maximum values of $\beta_{\text{sim-d}}$.

Found at doi:10.1371/journal.pbio.0050272.sg002 (210 KB PDF).

Figure S3. Average Nearest-Neighbor Beta-Diversity of Amphibians, Birds, and Mammals Mapped Continuously across the Continental Western Hemisphere

Average nearest-neighbor beta-diversity (β_{sim}) values are divided into 20 quantiles, represented by warm (higher β_{sim}) to cool (lower β_{sim}) colors. The scale accompanying the color ramp shows minimum, first quartile, median, third quartile, and maximum values of β_{sim} . Gray grid cells do not contain amphibian species. (A) Amphibians. (B) Birds. (C) Mammals.

Found at doi:10.1371/journal.pbio.0050272.sg003 (503 KB PDF).

Table S1. Correlations between Beta-Diversity ($\beta_{\text{sim-d}}$) and Two Environmental Variables (Elevation and Number of Biome Edges of Grid Cells)

Found at doi:10.1371/journal.pbio.0050272.st001 (81 KB PDF).

Table S2. Mean Elevation and Mean Number of Biome Edges for Sets of the Highest 2.5% and Lowest 2.5% of Beta-Diversity Grid Cells

Found at doi:10.1371/journal.pbio.0050272.st002 (56 KB PDF).

Table S3. Correlations in Average Nearest-Neighbor Beta-Diversity (β_{sim}) within the Western Hemisphere, Nearctic Realm, and Neotropical Realm

Found at doi:10.1371/journal.pbio.0050272.st003 (81 KB PDF).

Acknowledgments

We thank Thomas Brooks, Jack Lennon, David Orme, Jake Overton, John Terborgh, Dean Urban, and an anonymous reviewer for comments on the manuscript; John Bruno, Kevin Gaston, Aaron Moody, and Robert Peet for collaboration and discussion; and Gerardo Ceballos, Bruce Patterson, Marcelo Tognelli, James Zook, The Nature Conservancy, Conservation International, World Wildlife Fund, and Environment Canada–WILDSPACE for contributions to the compilation of bird and mammal range maps. Amphibian data were developed as part of the Global Amphibian Assessment and were provided by IUCN, Conservation International, and NatureServe.

Author contributions. MWM and PSW conceived and designed the study. RIM contributed new analytical tools. MWM, RIM, and JFL analyzed the data. WS, RSR, and SNS were responsible for the collection, organization, and verification of the species range maps. MWM, PSW, and JFL wrote the paper, with extensive input from RIM, WS, RSR, and SNS.

Funding. MWM thanks the US National Science Foundation Graduate Research Fellowship Program for financial support.

Competing interests. The authors have declared that no competing interests exist.

- Nekola JC, White PS (1999) The distance decay of similarity in biogeography and ecology. *J Biogeogr* 26: 867–878.
- Nekola JC, White PS (2002) Conservation, the two pillars of ecological explanation, and the paradigm of distance. *Nat Areas J* 22: 305–310.
- Gascon C, Malcolm JR, Patton JL, da Silva MNF, Bogart JP, et al. (2000) Riverine barriers and the geographic distribution of Amazonian species. *Proc Natl Acad Sci U S A* 97: 13672–13677.
- Hubbell SP (2001) *The unified neutral theory of biodiversity and biogeography*. Princeton (New Jersey): Princeton University Press. 375 p.
- Tuomisto H, Ruokolainen K, Yli-Halla M (2003) Dispersal, environment, and floristic variation of western Amazonian forests. *Science* 299: 241–244.
- Graham CH, Moritz C, Williams SE (2006) Habitat history improves prediction of biodiversity in rainforest fauna. *Proc Natl Acad Sci U S A* 103: 632–636.
- Pimm SL, Gittleman JL (1992) Biological diversity—Where is it? *Science* 255: 940.
- Blackburn TM, Gaston KJ (1996) The distribution of bird species in the New World: Patterns in species turnover. *Oikos* 77: 146–152.
- Lennon JJ, Koleff P, Greenwood JJD, Gaston KJ (2001) The geographical structure of British bird distributions: Diversity, spatial turnover and scale. *J Anim Ecol* 70: 966–979.

19. Drakare S, Lennon JJ, Hillebrand H (2006) The imprint of the geographical, evolutionary and ecological context on species-area relationships. *Ecol Lett* 9: 215–227.
20. Williams PH (1996) Mapping variations in the strength and breadth of biogeographic transition zones using species turnover. *Proc R Soc Lond B Biol Sci* 263: 579–588.
21. Williams PH, de Klerk HM, Crowe TM (1999) Interpreting biogeographical boundaries among Afrotropical birds: Spatial patterns in richness gradients and species replacement. *J Biogeogr* 26: 459–474.
22. Moritz C, Richardson KS, Ferrier S, Monteith GB, Stanicic J, et al. (2001) Biogeographical concordance and efficiency of taxon indicators for establishing conservation priority in a tropical rainforest biota. *Proc R Soc Lond B Biol Sci* 268: 1875–1881.
23. Pressey RL, Humphries CJ, Margules CR, Vane-Wright RI, Williams PH (1993) Beyond opportunism: Key principles for systematic reserve selection. *Trends Ecol Evol* 8: 124–128.
24. Sarkar S (2006) Ecological diversity and biodiversity as concepts for conservation planning: Comments on Ricotta. *Acta Biotheor* 54: 133–140.
25. Reid WV (1998) Biodiversity hotspots. *Trends Ecol Evol* 13: 275–280.
26. Ricketts TH, Dinerstein E, Boucher T, Brooks TM, Butchart SHM, et al. (2005) Pinpointing and preventing imminent extinctions. *Proc Natl Acad Sci U S A* 102: 18497–18501.
27. Stuart SN, Chanson JS, Cox NA, Young BE, Rodrigues ASL, et al. (2004) Status and trends of amphibian declines and extinctions worldwide. *Science* 306: 1783–1786.
28. Margules CR, Pressey RL (2000) Systematic conservation planning. *Nature* 405: 243–253.
29. Williams PH (1998) Key sites for conservation: Area-selection methods for biodiversity. In: Mace GM, Balmford A, Ginsberg JR, editors. *Conservation in a changing world*. Cambridge (United Kingdom): Cambridge University Press. pp. 211–250.
30. Ferrier S (2002) Mapping spatial pattern in biodiversity for regional conservation planning: Where to from here? *Syst Biol* 51: 331–363.
31. Fairbanks DHK, Reyers B, van Jaarsveld AS (2001) Species and environment representation: Selecting reserves for the retention of avian diversity in KwaZulu-Natal, South Africa. *Biol Conserv* 98: 365–379.
32. Steinitz O, Heller J, Tsoar A, Rotem D, Kadmon R (2005) Predicting regional patterns of similarity in species composition for conservation planning. *Conserv Biol* 19: 1978–1988.
33. Su JC, Debinski DM, Jakubauskas ME, Kindscher K (2004) Beyond species richness: Community similarity as a measure of cross-taxon congruence for coarse-filter conservation. *Conserv Biol* 18: 167–173.
34. Prendergast JR, Quinn RM, Lawton JH, Eversham BC, Gibbons DW (1993) Rare species, the coincidence of diversity hotspots and conservation strategies. *Nature* 365: 335–337.
35. Gaston KJ (1996) Biodiversity—Congruence. *Prog Phys Geogr* 20: 105–112.
36. Prendergast JR (1997) Species richness covariance in higher taxa: Empirical tests of the biodiversity indicator concept. *Ecography* 20: 210–216.
37. Grenyer R, Orme CDL, Jackson SF, Thomas GH, Davies RG, et al. (2006) Global distribution and conservation of rare and threatened vertebrates. *Nature* 444: 93–96.
38. Lamoreux JF, Morrison JM, Ricketts TH, Olson DM, Dinerstein E, et al. (2006) Global tests of biodiversity concordance and the importance of endemism. *Nature* 440: 212–214.
39. IUCN, Conservation International, NatureServe (2004) Digital distribution maps of the world's amphibians, version 1.1 [database]. Washington (D. C.) and Arlington (Virginia): IUCN, Conservation International, NatureServe. Available: <http://www.natureserve.org/getData/amphibianMaps.jsp>. Accessed 29 October 2005.
40. Ridgely RS, Allnutt TF, Brooks T, McNicol DK, Mehlman DW, et al. (2003) Digital distribution maps of the birds of the Western Hemisphere, version 1.0 [database]. Arlington (Virginia): NatureServe. Available: <http://www.natureserve.org/getData/birdMaps.jsp>. Accessed 29 January 2004.
41. Patterson BD, Ceballos G, Sechrest W, Tognelli MF, Brooks T, et al. (2003) Digital distribution maps of the mammals of the Western Hemisphere, version 1.0 [database]. Arlington (Virginia): NatureServe. Available: <http://www.natureserve.org/getData/mammalMaps.jsp>. Accessed 29 January 2004.
42. Ferrier S, Drielsma M, Manion G, Watson G (2002) Extended statistical approaches to modelling spatial pattern in biodiversity in northeast New South Wales. II. Community-level modelling. *Biodivers Conserv* 11: 2309–2338.
43. Koleff P, Gaston KJ, Lennon JJ (2003) Measuring beta diversity for presence-absence data. *J Anim Ecol* 72: 367–382.
44. Baillie JEM, Hilton-Taylor C, Stuart SN (2004) 2004 IUCN red list of threatened species: A global species assessment. Gland (Switzerland): The IUCN Species Program. Available: http://www.iucn.org/themes/ssc/red_list_2004/GSA_book/Red_List_2004_book.pdf. Accessed 28 August 2007.
45. Duellman WE (1999) Patterns of distribution of amphibians: A global perspective. Baltimore (Maryland): The Johns Hopkins University Press. 633 p.
46. Mayr E (1946) History of the North American bird fauna. *Wilson Bull* 58: 3–41.
47. Hawkins BA, Porter EE (2003) Relative influences of current and historical factors on mammal and bird diversity patterns in deglaciated North America. *Global Ecol Biogeogr* 12: 475–481.
48. Hershkovitz P (1972) The recent mammals of the Neotropical region: A zoogeographic and ecological review. In: Keast A, Erk FC, Glass B, editors. *Evolution, mammals, and southern continents*. Albany (New York): State University of New York Press. pp. 311–431.
49. van Rensburg BJ, Koleff P, Gaston KJ, Chown SL (2004) Spatial congruence of ecological transition at the regional scale in South Africa. *J Biogeogr* 31: 843–854.
50. Orme CDL, Davies RG, Burgess M, Eigenbrod F, Pickup N, et al. (2005) Global hotspots of species richness are not congruent with endemism or threat. *Nature* 436: 1016–1019.
51. Chown SL, Sinclair BJ, Leinaas HP, Gaston KJ (2004) Hemispheric asymmetries in biodiversity—A serious matter for ecology. *PLoS Biol* 2: e406.
52. Orme CDL, Davies RG, Olson VA, Thomas GH, Ding TS, et al. (2006) Global patterns of geographic range size in birds. *PLoS Biol* 4: e208.
53. Kattan GH, Franco P, Saavedra-Rodríguez CA, Valderrama C, Rojas V, et al. (2006) Spatial components of bird diversity in the Andes of Colombia: Implications for designing a regional reserve system. *Conserv Biol* 20: 1203–1211.
54. Bush MB (2002) Distributional change and conservation on the Andean flank: A palaeoecological perspective. *Global Ecol Biogeogr* 11: 463–473.
55. Hannah L, Midgley GF, Millar D (2002) Climate change-integrated conservation strategies. *Global Ecol Biogeogr* 11: 485–495.
56. Pounds JA, Bustamante MR, Coloma LA, Consuegra JA, Fogden MPL, et al. (2006) Widespread amphibian extinctions from epidemic disease driven by global warming. *Nature* 439: 161–167.
57. Collins JP, Halliday T (2005) Forecasting changes in amphibian biodiversity: Aiming at a moving target. *Philos Trans R Soc Lond B Biol Sci* 360: 309–314.
58. Watson DM (2005) Diagnosable versus distinct: Evaluating species limits in birds. *BioScience* 55: 60–68.
59. Moore JL, Balmford A, Brooks T, Burgess ND, Hansen LA, et al. (2003) Performance of sub-Saharan vertebrates as indicator groups for identifying priority areas for conservation. *Conserv Biol* 17: 207–218.
60. Ceballos G, Ehrlich PR (2006) Global mammal distributions, biodiversity hotspots, and conservation. *Proc Natl Acad Sci U S A* 103: 19374–19379.
61. Olson DM, Dinerstein E, Wikramanayake ED, Burgess ND, Powell GVN, et al. (2001) Terrestrial ecoregions of the world: A new map of life on earth. *BioScience* 51: 933–938.
62. McKnight MW (2007) Broad-scale patterns and determinants of beta-diversity [dissertation]. Chapel Hill (North Carolina): University of North Carolina at Chapel Hill. 161 p.
63. Soininen J, McDonald R, Hillebrand H (2007) The distance decay of similarity in ecological communities. *Ecography* 30: 3–12.
64. Manly BFJ (1997) Randomization, bootstrap and Monte Carlo methods in biology. London: Chapman and Hall. 399 p.
65. Clifford P, Richardson S, Hemon D (1989) Assessing the significance of the correlation between two spatial processes. *Biometrics* 45: 123–134.
66. Rosenberg MS (2001) PASSAGE (Pattern Analysis, Spatial Statistics, and Geographic Exegesis), version 1.1.2.3 [computer program]. Tempe (Arizona): Arizona State University.