

INDIVIDUALISTIC AND PHYLOGENETIC PERSPECTIVES
ON PLANT COMMUNITY PATTERNS

Jeffrey E. Ott

A dissertation submitted to the faculty of the University of North Carolina at Chapel Hill
in partial fulfillment of the requirements for the degree of Doctor of Philosophy
in the Department of Biology

Chapel Hill
2010

Approved by:

Robert K. Peet

Peter S. White

Todd J. Vision

Aaron Moody

Paul S. Manos

©2010
Jeffrey E. Ott
ALL RIGHTS RESERVED

ABSTRACT

Jeffrey E. Ott

**Individualistic and Phylogenetic Perspectives on Plant Community Patterns
(Under the direction of Robert K. Peet)**

Plant communities have traditionally been viewed as spatially discrete units structured by dominant species, and methods for characterizing community patterns have reflected this perspective. In this dissertation, I adopt an alternative, individualistic community characterization approach that does not assume discreteness or dominant species importance *a priori* (Chapter 2). This approach was used to characterize plant community patterns and their relationship with environmental variables at Zion National Park, Utah, providing details and insights that were missed or obscure in previous vegetation characterizations of the area.

I also examined community patterns at Zion National Park from a phylogenetic perspective (Chapter 3), under the assumption that species sharing common ancestry should be ecologically similar and hence be co-distributed in predictable ways. I predicted that related species would be aggregated into similar habitats because of phylogenetically-conserved niche affinities, yet segregated into different plots because of competitive interactions. However, I also suspected that these patterns would vary between different lineages and at different levels of the phylogenetic hierarchy (phylogenetic scales). I examined aggregation and segregation in relation to null models for each pair of species within genera and each sister pair of a genus-level vascular plant

supertree. Some pairs confirmed predictions, but many others did not, suggesting niche divergence rather than niche conservatism.

In the final chapter, I discuss community characterization from a phylogenetic perspective, exploring the possibility of using phylogenetic units in lieu of species in community analysis. I consider scenarios where species may not be optimal units of analysis, such as broad-scale community studies spanning species range limits. In such scenarios, species sharing common ancestry could potentially be merged and treated as a single unit. I present a method for identifying such species that I developed by adding a phylogenetic dimension to species clustering. This method is demonstrated through an analysis of bog and rock outcrop plant communities of the Southern Appalachian Mountains.

To my grandfather James. A. Ott,
scientific thinker and admirer of Southern Utah's natural wonders

ACKNOWLEDGEMENTS

My graduate work at the University of North Carolina would not have been possible without the support of individuals and institutions on both sides of the country. On the North Carolina side, I have had enjoyed working with the UNC Plant Ecology Lab, where many students, staff and post-docs who have provided friendship, technical assistance and a stimulating intellectual atmosphere. My graduate advisor Bob Peet and committee members Peter White, Todd Vision, Aaron Moody and Paul Manos have been helpful, supportive and patient as my research has slowly evolved. I am indebted to numerous field and data workers who have contributed to the Carolina Vegetation Survey, especially Susan Wiser and Brenda Wichmann for their work on Southern Appalachian rock outcrop and bog communities, respectively. I have received financial support from the UNC Graduate School in the form of the Graduate Merit Fellowship and Dissertation Completion Fellowship, as well as from the UNC Biology Department through the Mrs. W.C. Coker Fellowship, the Dr. W.C. Coker Fellowship in Botany, and the Alma Holland Beers Scholarship. In addition, I have held numerous teaching and research assistantships and express appreciation for my mentors Pat Gensel, Jim Massey, Seth Reice, Forbes Boyle, Charles Mitchell, Maria Servedio and Corey Johnson. Beyond professional circles, John and Heather Scott and the local Latter-day Saint community deserve mention for their role in welcoming me and helping me feel at home in North Carolina.

On the Utah side, I owe much to my long-time mentors E. Durant McArthur and Stewart Sanderson at the Rocky Mountain Research Station Shrub Sciences Laboratory of the U.S. Forest Service, as well as Brigham Young University/Utah Valley University professor Kimball Harper. This trio provided inspiration, encouragement and guidance toward my decision to pursue a doctorate degree. They were also the architects and builders of the 1987-89 Zion National Park vegetation survey that ultimately became a centerpiece of this dissertation. I acknowledge the many workers who contributed to this survey (Harper 1993) as well as the more recent USGS-NPS survey (Cogan et al. 2004) which also proved extremely useful. Zion National Park personnel, notably Elena Robisch, Leslie Courtright, Cheryl Decker and Dave Sharow, provided assistance and feedback during various stages of my research. Portions of this dissertation were funded through the Rocky Mountain Research Station, inasmuch as I laid the groundwork for Chapters 2 and 4 during the course of summer employment there. My parents C. Eric and Marilyn Ott graciously accommodated me during my frequent visits to my home state, and I wish to pay them special tribute for their unfailing love and support.

TABLE OF CONTENTS

CHAPTER 1: INTRODUCTION.....	1
REFERENCES.....	6
CHAPTER 2: INDIVIDUALISTIC SPECIES GROUPS OF ZION NATIONAL PARK.....	9
ABSTRACT.....	9
INTRODUCTION.....	10
BACKGROUND.....	15
Study Area	15
BYU-RMRS Vegetation Characterization (Harper et al. 2001).....	18
USGS-NPS Vegetation Characterization and Mapping (Cogan et al. 2004)	20
INDIVIDUALISTIC COMMUNITY CHARACTERIZATION: CONCEPTS AND METHODS	22
Conceptual Overview	22
Coalition Clustering and Affinity Calculations (Individualistic Fidelity Analysis)...	24
Affiliation Calculations (Individualistic Constancy and Dominance Analysis)	27
Environmental Modeling of Coalition Species Group Affinities.....	28
Quantifying Correspondence with Previously-described Vegetation Units	32
RESULTS.....	33
Coalition Species Groups and Their Environmental Affinities.....	33
Affiliate Species of Coalition Groups	41
Correspondence with Previously-described Vegetation Units	43

DISCUSSION	48
Community Continuity and Discreteness	49
Coalition Clustering Effects	52
Sampling Issues.....	54
Dominant Species, Floristic Patterns and Environmental Interpretations	58
REFERENCES.....	65
CHAPTER 3: PHYLOGENETIC NICHE OVERLAP IN PLANT COMMUNITIES OF ZION NATIONAL PARK.....	96
ABSTRACT	96
INTRODUCTION.....	97
METHODS.....	100
Study Area and Plant Community Data	100
Taxonomic/phylogenetic Data Synthesis	102
Niche Overlap Calculations	103
Significance Tests	104
RESULTS AND DISCUSSION	106
CONCLUSIONS	109
REFERENCES.....	111
CHAPTER 4: SHARPENING THE FOCUS OF COMMUNITY PATTERNS BY ADJUSTING PHYLOGENETIC SCALE.....	128
ABSTRACT	128
INTRODUCTION.....	129
1. SCALE CONCEPTS: EXTENT, GRAIN AND FOCUS	132
2. PROCESSES LINKED TO SPATIAL AND PHYLOGENETIC SCALE	135

3. REFOCUSING THE PHYLOGENETIC SCALE OF COMMUNITY PATTERNS	137
4. AN EXPLORATORY APPROACH FOR PHYLOGENETIC REFOCUSING ..	139
5. APPLICATION OF PHYLOGENETIC SCALE REFOCUSING FOR SOUTHERN APPALACHIAN PLANT COMMUNITIES	143
CONCLUSION	144
REFERENCES	148

LIST OF TABLES

Table 2.1. Geologic substrate classes used for environmental modeling of plant species group affinities at Zion National Park	84
Table 2.2. Coalition species groups and affiliate species of Zion National Park, based on coalition clustering of 1987-89 vegetation survey data	85
A. Streambank.....	85
B. High Plateau	86
C. Crevice Canyon	87
D. Slickrock.....	88
E. Upland Sands.....	89
F. Rocky Slopes.....	90
G. Arid Lowlands.....	91
H. Lowland Flats	92
I. Mesic Mesa Top	93
J. Xeric Mesa Top	94
Table 2.3. Affinity correspondence between vegetation units of Harper et al. (2001) and coalition species groups of the current study at Zion National Park	95
Table 2.4. Affinity correspondence between National Vegetation Classification System formation classes of Cogan et al. (2004) and coalition species groups of the current study at Zion National Park	95
Table 2.5. Affinity correspondence between lumped vegetation map units of Cogan et al. (2004) and coalition species groups of the current study at Zion National Park	95
Table 3.1. Ecological divisions used to group alliances of the U.S. National Vegetation Classification at Zion National Park	125
Table 3.2. References used to resolve phylogenetic relationships within families for the genus-level vascular plant supertree used in the current study	126
Table 3.3. Summary of niche overlap tests for congeneric pairs and phylogenetic sister groups (genus and above) of vascular plant species at Zion National Park.....	127

Table 4.1. Multilevel Coalition Clusters obtained by applying the MOCC algorithm to a plant community dataset of 219 100 m² vegetation plots taken from the Carolina Vegetation Survey, representing high elevation rock outcrops and mountain bogs.....**161**

LIST OF FIGURES

Figure 2.1. Zion National Park, Utah in relation to topography, showing plot locations of the 1987-89 vegetation survey	73
Figure 2.2. Plot locations of two vegetation survey datasets at Zion National Park	73
Figure 2.3. Abiotic environmental variable layers derived from a 10-m resolution digital elevation model of Zion National Park.....	74
Figure 2.4. Geologic substrate classes derived from a 1:24,000 scale digital geologic map of Zion National Park and vicinity.....	74
Figure 2.5. Regression tree model and map for the Streambank coalition group.....	75
Figure 2.6. Regression tree model and map for the High Plateau coalition group	75
Figure 2.7. Regression tree model and map for the Crevice Canyon coalition group	76
Figure 2.8. Regression tree model and map for the Slickrock coalition group.....	76
Figure 2.9. Regression tree model and map for the Upland Sands coalition group.....	77
Figure 2.10. Regression tree model and map for the Rocky Slopes coalition group.....	77
Figure 2.11. Regression tree model and map for the Arid Lowlands coalition group.....	78
Figure 2.12. Regression tree model and map for the Lowland Flats coalition group	78
Figure 2.13. Regression tree model and map for the Mesic Mesa Top coalition group ...	79
Figure 2.14. Regression tree model and map for the Xeric Mesa Top coalition group....	79
Figure 2.15. Cluster dendrogram of National Vegetation Classification System associations of Zion National Park based on compositional affinities to coalition species groups presented in this paper.....	80
Figure 2.16. Cluster dendrogram of Zion National Park vegetation map units based on pixel-averaged affinities to coalition species groups presented in this paper	81
Figure 2.17. Two Arid Lowlands species groups of Zion National Park obtained through coalition clustering in RCLUS at a threshold affinity value of phi=0.2	82
Figure 2.18. Histograms of proportions of background pixels representing four topographic variables at Zion National Park against pixels represented by plots for the BYU-RMRS dataset and USGS-NPS dataset.....	83

Figure 2.19. Histograms of proportions of background pixels in different geologic substrate classes at Zion National Park against pixels represented by plots for the BYU-RMRS dataset and USGS-NPS dataset.....	83
Figure 3.1. Genus-level phylogenetic supertree of vascular plants reported in the USGS-NPS vegetation survey of Zion National Park	119
Figure 3.2. Hypothetical scenario illustrating procedure for aggregating cover values of multiple species (or strata) within plots	120
Figure 3.3. Results of beta niche overlap tests based on presence/absence, mapped onto a genus-level phylogeny of vascular plants of Zion National Park	121
Figure 3.4. Results of beta niche overlap tests based on cover, mapped onto a genus-level phylogeny of vascular plants of Zion National Park	122
Figure 3.5. Results of alpha niche overlap tests based on presence/absence, mapped onto a genus-level phylogeny of vascular plants of Zion National Park	123
Figure 3.6. Results of alpha niche overlap tests based on cover, mapped onto a genus-level phylogeny of vascular plants of Zion National Park.....	124
Figure 4.1. Conceptual parallels between spatial scale and phylogenetic scale of community studies: extent and grain	155
Figure 4.2. Conceptual parallels between spatial scale and phylogenetic scale of community studies: focus	156
Figure 4.3. Conceptual model of expected aggregation and segregation patterns of closely related species under different scenarios of spatial and ecological divergence ..	157
Figure 4.4. Hypothetical scenario illustrating phylogenetic refocusing of community patterns.....	158
Figure 4.5. Expansion of a species by species pairwise association matrix to include higher taxa	159
Figure 4.6. Location of high elevation rock outcrop and mountain bog vegetation survey plots in the Southern Appalachian Mountains, USA, analyzed using Multilevel Ordered Coalition Clustering	160

LIST OF APPENDICES

APPENDIX 2.1. Taxa recorded by the 1999-2000 USGS-NPS vegetation survey of Zion National Park, Utah (Cogan et al. 2004), following nomenclature of Kartesz (1999), with translations to equivalent taxonomic concepts of the 1987-89 BYU-RMRS survey (Harper et al. 2001), updated to nomenclature of Welsh et al. (2003)	162
APPENDIX 2.2. Vascular plant species/taxa recorded by the 1987-89 vegetation survey of Zion National Park, Utah, and their affinities to coalition species groups	173
APPENDIX 2.3. Modal species of vegetation types described by Harper et al. (2001) for Zion National Park, Utah and their affinities to coalition species groups of the current study.....	190
APPENDIX 2.4. U.S. National Vegetation Classification System associations of Zion National Park, Utah as recognized by Cogan et al. (2004) with their compositional affinities to coalition species groups of the current study.....	195
APPENDIX 2.5. Vegetation map units of Zion National Park, Utah as mapped by Cogan et al. (2004) (rows) with their spatial correspondence to coalition species groups of the current study.....	201
APPENDIX 3.1. Vascular plant genera and species included in niche overlap analyses for plant communities of Zion National Park, Utah, showing correspondence between different taxonomic concepts	204
APPENDIX 3.2. Vascular plant genera and species included in niche overlap analyses for plant communities of Zion National Park, Utah, showing correspondence between different taxonomic concepts. Results of niche overlap significance tests comparing congeneric species of vascular plants recorded in the USGS-NPS vegetation characterization of Zion National Park, Utah	215
APPENDIX 3.3. Results of niche overlap significance tests comparing phylogenetic sister groups (genus and above) for vascular plants recorded in the USGS-NPS vegetation characterization of Zion National Park, Utah	223

CHAPTER 1

INTRODUCTION

This dissertation deals with plant communities: how they are conceptualized, sampled and characterized; and the environmental, evolutionary and historical factors influencing their composition and distribution. Describing and understanding plant communities is an important component of the discipline known as plant community ecology or vegetation science (Mueller-Dombois and Ellenberg 1974, Kent and Coker 1991). Building on this research tradition, I have sought to characterize and interpret plant community patterns for a specific area of interest: Zion National Park, Utah. I have also sought to contribute to the conceptual and methodological advancement of this field by synthesizing concepts, offering new perspectives and developing new approaches for community analysis.

The main chapters of this dissertation revolve around two themes, (1) the individualistic concept of the plant community and (2) phylogenetic community organization. The purpose of this introductory chapter is to briefly explain how these seemingly disparate concepts relate to one another in the context of community characterization.

Plant communities have traditionally been characterized and mapped as discrete units differentiated based on composition, structure or physiognomy. Floristic community classifications systems such as those developed by the Zurich-Montpellier

school of phytosociology (Braun-Blanquet 1965, Ewald 2003) or the U.S. National Vegetation Classification (Jennings et al. 2009) use taxonomic composition, generally at the species level, as the primary criteria for differentiating community units. Plant communities are thus commonly portrayed as species assemblages occurring within discretely-defined spatial areas. From an individualistic perspective, this portrayal is problematic because species may not be co-distributed in a discrete manner, whereas from a phylogenetic perspective, the species emphasis conceals evolutionary relationships that may be relevant to community organization.

The individualistic concept as developed by Gleason (1926, 1939) addresses the variable nature of species distributions and environmental responses. Gleason pointed out that community boundaries may not be easily identified because of this individualistic variability. Subsequent authors influenced by Gleason's work developed alternatives to community classification (e.g., ordination) that do not require discrete community classes (Whittaker 1967, Mueller-Dombois and Ellenberg 1974, Kent and Coker 1992, McCune and Grace 2002). Others have used the individualistic concept to justify species-specific as opposed to community-level environmental modeling (e.g. Evans and Cushman 2009). The individualistic concept has been interpreted by some to mean that individual species distributions bear no relation to one another and that community units are entirely arbitrary constructs (Shipley and Keddy 1987, Leibold and Mikkelsen 2002). However, Gleason's (1926, 1939) view was more balanced in that he acknowledged the existence of recurring community patterns, including species sharing sufficiently similar environmental responses to justify their recognition as ecological species groups. This balanced perspective between species individualism and similarity underlies the

“individualistic species group” approach that I use to characterize plant community patterns of Zion National Park (Chapter 2). Instead of assuming that species are organized into discrete community units, I assume that some species are more similarly distributed along environmental gradients than others, and use cluster analysis to identify such species groups. Since species groups are spatially-diffuse constructs, this approach leads to continuous-scale representations of spatial community variability.

The notion that all species are distributed independently of each other can be countered on ecological and evolutionary grounds. Plant species interact, often through competition or other negative interactions that limit their ability to coexist (Silvertown 2004), but also through positive interactions such as facilitation that promote coexistence (Callaway 1997). Species attributes related to local coexistence comprise what has become known as the alpha niche, as contrasted with beta niche attributes related to species responses along environmental gradients (Pickett and Bazazz 1978, Silvertown et al. 2006). Alpha and beta niche attributes are likely to be similar in species sharing recent common ancestry because of phylogenetic niche conservatism, or the tendency for niche attributes to be retained over evolutionary time (Harvey and Pagel 1991, Webb et al. 2002). Species with conserved alpha niches might be expected to compete for resources and thus be segregated at local scales, while species with conserved beta niches might be expected to be aggregated with respect to environmental gradients (Webb et al. 2002, Silvertown et al. 2006, Cavender-Bares et al. 2009)—hypotheses that I examine for plants of Zion National Park in Chapter 3. Phylogenetic niche conservatism thus provides reason to expect non-random, non-individualistic distribution patterns among at least some species in communities.

Whereas proponents of the individualistic concept may prefer that species be treated as independent units in community studies, proponents of the phylogenetic perspective may argue that species are not statistically independent and should be examined only in a phylogenetic context (Harvey and Pagel 1991, Harvey et al. 1995). The phylogenetic perspective has fueled the recent development of metrics for measuring phylogenetic relatedness and phylogenetic diversity of communities (Webb 2000, Webb et al. 2002, Hardy and Senterre 2007, Helmus et al. 2007, Graham and Fine 2008, Cadotte et al. 2010). These metrics offer phylogenetic alternatives to the species-based metrics commonly used in community characterization and analysis (e.g., McCune and Grace 2002). In many respects, this emerging field of phylogenetic community analysis parallels the earlier emergence of continuum analyses such as ordination. Phylogeny transforms taxonomic units into continuous fields of relatedness, just as ordination transforms community units into continuous fields of compositional similarity. In both cases, continuous representations of community patterns may be useful but also carry certain drawbacks. For phylogenetic community analysis, a major drawback is that the presumed correlation between phylogenetic relatedness and niche similarity may not be consistent across lineages with different evolutionary histories (Giannini 2003, Losos 2008). In some lineages niche conservatism may be essentially overridden by rapid niche divergence, as in adaptive radiations (Levin 2005, Ackerly 2009). Even in lineages exhibiting niche conservatism, the degree of conservatism may differ for different niche attributes. Some types of niche conservatism may only be detectable or relevant at narrow phylogenetic scales (e.g. between closely-related species within a genus).

In light of these drawbacks of strictly continuous phylogenetic community analysis, I propose an alternative approach that incorporates phylogenetic information but recognizes discontinuities of phylogenetic niche conservatism. Instead of dissolving taxonomic units into measures of phylogenetic relatedness, I advocate an approach that maintains unit structure but merges units when niche conservatism is evident. Species are merged into broader phylogenetic units if they show similar habitat (beta niche) affinities, as inferred from their association patterns with other species (or other merged phylogenetic units). I implement this approach through an algorithm derived from the species clustering algorithm used to identify species groups in Chapter 2. The new algorithm (multilevel ordered coalition clustering) is presented in Chapter 4, along with an explanation of its conceptual basis and applicability. Reasoning that this approach may be most useful for community studies spanning large areas (with high spatial species turnover), I use plant community data drawn from the Southern Appalachian region to demonstrate the new algorithm.

Chapter 4, which caps this dissertation, also includes a discussion of parallels between spatial scale and phylogenetic scale in community studies. Following previous treatments of spatial scale (Levin 1992, Scheiner et al. 2000), I introduce grain, extent and focus as components of phylogenetic scale.

REFERENCES

- Ackerly, D. 2009. Conservatism and diversification of plant functional traits: Evolutionary rates versus phylogenetic signal. *Proceedings of the National Academy of Sciences of the United States of America* 106:19699-19706.
- Braun-Blanquet, J. 1965. *Plant Sociology: The Study of Plant Communities*. Hafner Pub. Co., New York.
- Cadotte, M. W., T. J. Davies, J. Regetz, S. W. Kembel, E. Cleland, and T. H. Oakley. 2010. Phylogenetic diversity metrics for ecological communities: Integrating species richness, abundance and evolutionary history. *Ecology Letters* 13:96-105.
- Callaway, R. M. 1997. Positive interactions in plant communities and the individualistic-continuum concept. *Oecologia* 112:143-149.
- Cavender-Bares, J., K. H. Kozak, P. V. A. Fine, and S. W. Kembel. 2009. The merging of community ecology and phylogenetic biology. *Ecology Letters* 12:693-715.
- Evans, J. S., and S. A. Cushman. 2009. Gradient modeling of conifer species using random forests. *Landscape Ecology* 24:673-683.
- Ewald, J. 2003. A critique for phytosociology. *Journal of Vegetation Science* 14:291-296.
- Giannini, N. P. 2003. Canonical phylogenetic ordination. *Systematic Biology* 52:684-695.
- Gleason, H. A. 1926. The individualistic concept of the plant association. *Bulletin of the Torrey Botanical Club* 53:7-26.
- Gleason, H. A. 1939. The individualistic concept of the plant association. *American Midland Naturalist* 21:92-110.
- Graham, C. H., and P. V. A. Fine. 2008. Phylogenetic beta diversity: Linking ecological and evolutionary processes across space in time. *Ecology Letters* 11:1265-1277.
- Hardy, O. J., and B. Senterre. 2007. Characterizing the phylogenetic structure of communities by an additive partitioning of phylogenetic diversity. *Journal of Ecology* 95:493-506.
- Harvey, P. H., and M. D. Pagel. 1991. *The Comparative Method in Evolutionary Biology*. Oxford University Press, Oxford; New York.
- Harvey, P. H., A. F. Read, and S. Nee. 1995. Why ecologists need to be phylogenetically challenged. *Journal of Ecology* 83:535-536.

- Helmus, M. R., T. J. Bland, C. K. Williams, and A. R. Ives. 2007. Phylogenetic measures of biodiversity. *American Naturalist* 169:E68-E83.
- Jennings, M. D., D. Faber-Langendoen, O. L. Loucks, R. K. Peet, and D. Roberts. 2009. Standards for associations and alliances of the US National Vegetation Classification. *Ecological Monographs* 79:173-199.
- Kent, M., and P. Coker. 1992. *Vegetation Description and Analysis: A Practical Approach*. CRC Press, Boca Raton.
- Leibold, M. A., and G. M. Mikkelson. 2002. Coherence, species turnover, and boundary clumping: elements of meta-community structure. *Oikos* 97:237-250.
- Levin, D. A. 2005. Niche shifts: the primary driver of novelty within angiosperm genera. *Systematic Botany* 30:9-15.
- Levin, S. A. 1992. The problem of pattern and scale in ecology. *Ecology* 73:1943-1967.
- Losos, J. B. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. *Ecology Letters* 11:995-1003.
- McCune, B., and J. B. Grace. 2002. *Analysis of Ecological Communities*. MjM Software Design, Gleneden Beach, OR.
- Mueller-Dombois, D., and H. Ellenberg. 1974. *Aims and Methods of Vegetation Ecology*. Wiley, New York.
- Pickett, S. T. A., and F. A. Bazzaz. 1978. Organization of an assemblage of early successional species on a soil-moisture gradient. *Ecology* 59:1248-1255.
- Scheiner, S. M., S. B. Cox, M. Willig, G. G. Mittelbach, C. Osenberg, and M. Kaspari. 2000. Species richness, species-area curves and Simpson's paradox. *Evolutionary Ecology Research* 2:791-802.
- Shipley, B., and P. A. Keddy. 1987. The individualistic and community-unit concepts as falsifiable hypotheses. *Vegetatio* 69:47-55.
- Silvertown, J. 2004. Plant coexistence and the niche. *Trends in Ecology and Evolution* 19:605-611.
- Silvertown, J., M. Dodd, D. Gowing, C. Lawson, and K. Mcconway. 2006. Phylogeny and the hierarchical organization of plant diversity. *Ecology* 87:S39-S49.
- Webb, C.O. 2000. Exploring the phylogenetic structure of ecological communities: An example for rain forest trees. *American Naturalist* 156:145-155.

Webb, C. O., D. D. Ackerly, M. A. Mcpeak, and M. J. Donoghue. 2002. Phylogenies and community ecology. Annual Review of Ecology and Systematics 33:475-505.

Whittaker, R. H. 1967. Gradient analysis of vegetation. Biological Reviews of the Cambridge Philosophical Society 42:207-264.

CHAPTER 2

INDIVIDUALISTIC PLANT SPECIES GROUPS OF ZION NATIONAL PARK

ABSTRACT

Questions: Can plant community characterization be carried out in a manner consistent with Gleason's individualistic concept? What new insights can be obtained by redefining communities in terms of individualistic species groups rather than discrete vegetation units?

Location: Zion National Park, Utah, U.S.A.

Methods: Individualistic community characterization is demonstrated through re-analysis of a systematically-sampled vegetation dataset. Overlapping species groups identified through coalition clustering ('coalition groups') were treated as individualistic equivalents of faithful species, while 'affiliate' species having high co-occurrence and high cover with respect to these groups were treated as constant and dominant species, respectively. Environmental affinities of each coalition group were modeled and mapped through regression tree analysis. Results were compared with two previous vegetation characterizations of the park.

Results: Coalition groups overlapped spatially and compositionally, especially in terms of affiliate species, but each represented a unique pattern with interpretable environmental relationships. A mesic gradient from low-elevation riparian zones through mid-elevation narrow canyons to high-elevation environments was represented by three

overlapping coalition groups. Two groups occupying slickrock and sand environments were detected on the Navajo Sandstone, as well as two on mesa tops above it. At lower elevations, three intergrading xeric coalition groups were distinguished. These patterns had been obscured in previous vegetation characterizations that had demanded discrete units, emphasized dominant species, applied a physiognomic classification hierarchy and/or sampled non-systematically.

Conclusions: The coalition clustering approach achieved community characterization objectives without assuming spatial discreteness or dominant species preeminence. The individualistic concept is not only compatible with community characterization, but provides a solid conceptual framework for interpreting community patterns.

INTRODUCTION

Gleason's (1926, 1939) individualistic concept, though widely recognized and accepted by contemporary plant ecologists, remains at the margins of plant community characterization activities. The individualistic concept emphasizes the unique distribution patterns and ecological requirements of individual species, and therefore questions the assumption that communities are discrete, integrated units (Gleason 1926, 1939). Gleason's criticisms of discrete community classification were initially dismissed, but influenced the later development of continuous community modeling approaches such as ordination (McIntosh 1975). The individualistic concept also eventually became more widely favored than Clements' (1916) organismal concept, which views communities as integrated entities structured by their dominant plant species (Clements 1916, 1936, McIntosh 1975, Eliot 2007). Despite these developments, discrete

vegetation classification emphasizing structural dominants continues to be a popular approach for characterizing plant communities. The U.S. National Vegetation Classification System (NVCS), for example, seeks to define spatially discrete units that can be recognized by their dominant species, structure and physiognomy (Grossman et al. 1998, FGDC 2008, Jennings et al. 2009).

The continued practice of discrete vegetation classification has been justified on the grounds of convenience and utility. Discrete community models are relatively easy to describe and map because of their well-defined boundaries. Proponents of the NVCS acknowledge that community boundaries may not be as sharp as their models depict, but argue that they provide useful, if sometimes arbitrary, partitions of community variation (Grossman et al. 1998, Comer et al. 2003). Even Curtis (1959), widely regarded as a pioneer in ordination approaches, recognized the practical value of dividing continuous community variation into discrete units. Working with these units, Curtis (1959) was able to characterize community features such as prevalent species (commonly encountered within a community) and modal species (concentrated within a community), much as others have done elsewhere using the essentially equivalent terms *constant species* and *faithful species* (Barkman 1989, Kent and Coker 1992). Constancy and fidelity are useful concepts for conveying ecological information about individual species and allowing community units to be identified in the field. Species that are both constant and abundant (dominant species) are especially useful for visualizing and identifying community units, whether from the ground or from an aerial view. Signatures of community dominants in aerial or satellite imagery are now routinely used in vegetation mapping applications, including those related to the NVCS (e.g., Cogan et al. 2004,

Faber-Langendoen et al. 2007). Ancillary environmental data are also often integrated with community data so that environmental factors responsible for community distributions can be inferred (Mueller-Dombois and Ellenberg 1974, Kent and Coker 1992, McCune and Grace 2002). In short, discrete vegetation classification is a well-established, practical means for condensing and presenting valuable ecological information.

Can the useful products of discrete vegetation classification—compositional, spatial and environmental representations of plant community patterns—be obtained without dismissing the individualistic concept? What are the benefits of aligning community classification practices more closely with individualistic principles? This paper addresses these questions by formulating and applying individualistic alternatives to conventional vegetation mapping, environmental modeling and compositional characterization (including faithful, constant and dominant species). Using novel extensions of existing techniques such as coalition clustering (Sanderson et al. 2006) and regression tree modeling (Breiman 1993), I demonstrate an ‘individualistic species group approach’ that characterizes communities without requiring discrete community units or over-emphasizing dominant species.

The ‘individualistic principles’ that guide this approach are found in the original writings of Gleason. Gleason stressed the fact that each plant species carries its own history and is to some degree ecologically unique:

In conclusion, it may be said that every species of plant is a law unto itself, the distribution of which in space depends upon its individual peculiarities of migration and environmental requirements (Gleason 1926, p. 26).

By emphasizing individualistic variability in plant species distributions due to different migration histories and environmental affinities, Gleason countered the notion that plant

associations (community units) are sufficiently distinct, discrete and integrated to be unambiguously classified on the basis of species composition or vegetation structure. He did not, however, deny that recurring patterns approximating discrete communities can often be observed in nature (Gleason 1926, p. 25-26). Gleason recognized that species individualism (spatial and ecological uniqueness) is itself a matter of degree, with some species demonstrating a greater degree of similarity than others:

The [individualistic] concept is by no means opposed to the recognition of the synusia, or union, defining it as a group of plants whose physiological demands are so similar that they are regularly selected by the same environment and consequently regularly live together (Gleason 1939, p. 108).

Hence an individualistic approach to community characterization permits the dual objective of identifying ecologically similar species, while also recognizing and quantifying their differences.

The “unions” mentioned by Gleason later came to be referred to as “ecological species groups” by plant ecologists in Europe and elsewhere (Mueller-Dombois and Ellenberg 1974, Abella and Shelburne 2004). Analogous “sociological species groups” defined by overlapping distribution patterns were promoted by Doing (1969). Relaxing the requirement that species within a group belong to the same life form leads to the definition of “species group” often used in contemporary community studies: a set of species with similar distributions among community samples, typically identified through cluster analysis (Austin and Belbin 1982, Ferrier et al. 2002, McCune and Grace 2002). The “individualistic species groups” presented in this paper have the additional distinction of being quasi-independent, potentially-overlapping species sets. They also serve as substitutes for faithful species and provide starting points for community characterizations that are spatially continuous and not biased toward dominant species.

The individualistic species group approach is demonstrated for plant communities of Zion National Park (Zion), an important conservation area in Utah, U.S.A. with pronounced topographic, edaphic, and hydrological variation. Relatively abrupt transitions between geologic substrates, xeric and mesic zones, exposed and shaded slopes, and lower and upper elevations are noticeable in some areas of Zion, but in other areas these environmental gradients are more subtle and gradual (Biek et al. 2003, Sharro 2007a, 2007b). Zion houses a rich flora including over 900 native species that are differentiated among the park's environments (Fertig and Alexander 2009), yet relatively few plant species are dominants (Harper 1993, Harper et al. 2001, Cogan et al. 2004, Fertig and Alexander 2009). These characteristics of Zion suggest that an examination of community patterns without bias toward discrete zonation or dominant species would be an appropriate compliment to previous studies.

Specifically, this paper presents a re-analysis of the dataset that Harper (1993) and Harper et al. (2001) used to characterize the vegetation of Zion. In the previous analysis, vegetation survey plots were classified into vegetation types whose constant and faithful species were described using Curtis' (1959) prevalent and modal species concepts (Harper 1993, Harper et al. 2001). In the current analysis, faithful species take the form of individualistic species groups derived directly from association patterns through cluster analysis, and constant species are defined as species that tend to co-occur with these species groups. Ten individualistic species groups are presented along with their co-occurring species, environmental relationships and distribution. These results are compared with the previous vegetation characterizations of Harper et al. (2001) as well as those of Cogan et al. (2004) who applied the NVCS at Zion.

BACKGROUND

Study Area

Zion National Park (Zion), Utah, U.S.A. was established in 1919 to protect the striking scenery of Zion Canyon, where massive walls of the Navajo Sandstone formation tower above the Virgin River (NPS 2009). Subsequent expansion to include surrounding canyons, mesas and lowlands brought the park to its current ca. 590 km² (Cogan et al. 2004, NPS 2009). Zion is comprised of two roughly rectangular sections (Fig. 2.1), a large southeastern section that includes Zion Canyon and Coalpits Wash (lowest elevation in Zion, 1128 m) and a smaller northwestern section that includes the Kolob Canyons and Horse Pasture Plateau (highest elevation in Zion, 2660 m) (Cogan et al. 2004).

Zion lies at the western edge of the Colorado Plateau physiographic province, bordering the Basin and Range province to the west at the Hurricane fault escarpment (Biek et al. 2003). The Basin and Range can be further divided into the higher-elevation Great Basin northwest of Zion and the lower-elevation Mojave Desert to the southwest. The Colorado Plateau, Great Basin and Mojave Desert have been recognized as distinct floristic regions (McLaughlin 1986, 1989); and Zion, because of its position near their intersection, contains floristic elements characteristic of each (Meyer 1978, Cogan et al. 2004, Fertig and Alexander 2009).

Zion has a semiarid climate with seasonal temperature changes and precipitation concentrated in winter/spring frontal storms and late-summer monsoons (Woodbury 1933, Mortensen et al. 1977, Sharro 2007b). During the period 1959-1989 (i.e., 30 years preceding the 1987-89 vegetation survey discussed below), mean annual

precipitation at the Zion Canyon weather station (1234 m) was 388 mm and mean annual temperatures were 6.3° C in January and 34.5° C in July (UCC 2010). Local climate varies along the 1128–2660 m elevation gradient (Cogan et al. 2004) and is further modified by topographic position and exposure. Moisture is concentrated in seeps and springs that emerge from porous substrates, and in permanent and intermittent streams that are periodically exposed to flash flood disturbance (Sharrow 2007a).

Horizontally-aligned exposures of sedimentary rocks, including both cliff and bench-forming strata, are prominent at Zion. The oldest (Permian to Early Jurassic) strata occur at low elevations on the southwestern side of the park and on slightly higher elevations on the northwestern side where the Taylor Creek thrust fault disrupts their stratigraphic continuity (Biek et al. 2003, O'Meara 2006). These strata, which include the Kiabab, Moenkopi, Chinle, Moenhave and Kayenta formations, contain alternating bands of mudstone, siltstone, sandstone, conglomerate, limestone and gypsum (Biek et al. 2003). The badlands, slopes, and small cliffs of these lower formations at Zion are dwarfed by the massive cliffs and ‘slickrock’ exposures of the overlying Jurassic Navajo Sandstone (Biek et al. 2003). The stratigraphic sequence is topped by Middle Jurassic and Cretaceous sandstone, limestone, shale and gypsum deposits of the Temple Cap, Carmel and Cedar Mountain formations (Biek et al. 2003, O'Meara 2006). The entire sequence is in turn partially overlain by Quaternary deposits including volcanic basalt flows and cinder cones, alluvium, lacustrine, landslide, talus, residual and eolian deposits (Biek et al. 2003, O'Meara 2006, Sharrow 2007a).

Soil development has been inhibited in many areas of Zion because of rapid erosion by water, wind and gravity, leading to exposed rock outcrops in some areas and

recent deposits with limited soil profile development (entisols) in others (Mortensen et al. 1977, Biek et al. 2003, Sharro 2007a). Aridisols and mollisols are also present in Zion at lower and higher elevations, respectively; and alfisols can be found at the highest elevations (above 1830 m) (Mortensen et al. 1977).

Many of Zion's geologic substrates and their derivative soils have distinctive physical and chemical properties that pose challenges to plants, as exemplified by the bentonite shrink-swell clays of the Petrified Forest member of the Chinle Formation and the solid slickrock surfaces of the Navajo Sandstone (Biek et al. 2003, Sharro 2007a, Cogan et al. 2004). In other instances substrate and hydrology combine to favor plant growth, as in the hanging gardens that occur on seepage areas of canyon walls (Malanson and Kay 1980, Welsh 1989).

Substrate variation and gradients of moisture and temperature related to hydrology, topographic position, exposure and elevation have produced a variety of different vegetation types at Zion (Woodbury 1933, Harper 1993, Harper et al. 1992, 2001, 2003, Cogan et al. 2004). As a general pattern, desert shrublands and grasslands at lower elevation, more exposed sites give way to coniferous woodlands, mountain brush, and montane forests and grasslands at higher elevation, less exposed sites (Harper 1993, Cogan et al. 2004). Riparian areas and wetlands, which are most extensive at lower elevations, are occupied by various forest, woodland, shrubland and herbaceous vegetation types (Harper et al. 1992, Cogan et al. 2004).

Although most of Zion currently has an undeveloped wilderness character, the park is not free of human disturbance. Euro-American settlement beginning in the mid-1800s led to agricultural clearing in low-elevation floodplains and widespread livestock

introductions throughout the Zion area (Woodbury 1933, Madany and West 1983). Heavy livestock grazing in the early part of the twentieth century followed by fire suppression later in the century favored an increase of woody plants at the expense of grasses and forbs (Madany and West 1983, Jeffries and Klopatek 1987). Isolated mesas in the central portion of the park escaped grazing impacts because of their inaccessibility (Madany and West 1983, 1984). More recently, livestock have been removed but mule deer (*Odocoileus hemionus*) have increased in areas of high human visitation, resulting in declines in cottonwood (*Populus fremontii*) and other riparian species (Ripple and Beschta 2006). The vegetation survey data used here record a snapshot of conditions following 30+ years of livestock exclusion. The troublesome invasive species cheatgrass (*Bromus tectorum*), red brome (*B. rubens*), ripgut brome (*B. diandrus*) and tamarisk (*Tamarix* spp.) were already well-established in Zion at the time of sampling.

BYU-RMRS Vegetation Characterization (Harper et al. 2001)

The first park-wide, plot-based vegetation survey of Zion was carried out in 1987-1989 by workers affiliated with Brigham Young University (BYU) and the Rocky Mountain Research Station (RMRS, formerly the Intermountain Research Station) Shrub Sciences Laboratory of the U.S. Forest Service. This survey, which I refer to by the acronym BYU-RMRS, is the primary source of species occurrence data for the current study. The survey has previously been described in a series of reports and manuscripts (Harper 1993, Harper et al. 1992, 2001, 2003). Data from the BYU-RMRS survey will soon be archived in the VegBank repository (ESA 2010).

The BYU-RMRS survey implemented a systematic sampling strategy in which U.S. Geological Survey (USGS) cadastral section corners and corresponding park boundary markers served as sampling targets (Fig. 2.1-2.2). In addition to 270 plots located at or near these markers, 18 plots were placed using subjective criteria in order to capture spatially-restricted plant communities (riparian areas, hanging gardens, and old fields). The standard plot was a 100 m² circle with the exception of six equivalently-sized plots that were rectangular or square. Data collected within plots included vascular plant species composition following Welsh et al. (1987), cover classes for each species according to the Braun-Blanquet system (Braun-Blanquet 1965), and percent cover of bare soil, rock, litter and cryptogams (Harper 1993, Harper et al. 2001).

Harper (1993) used the BYU-RMRS survey data to classify the vegetation of Zion into ten discrete vegetation types that were concisely summarized in a subsequent manuscript (Harper et al. 2001). The vegetation types were recognized on account of being “commonly used by land managers in the area” (Harper et al. 2001) and each plot was intuitively assigned to a vegetation type based on dominant species and environmental context. Plot data were then used to characterize attributes of each vegetation type including prevalent and modal species following Curtis (1959). Prevalent species are defined as the most frequently encountered species (occurring in the most plots) within a community or vegetation type, where the number of prevalent species is equal to the species density (average number of species per plot) for the community (Curtis 1959, Harper et al. 2001). Modal species are defined as prevalent species that reach their greatest frequency in a given community relative to other communities under

study. In Harper et al. (2001), modal species were identified for each vegetation type relative to the others described at Zion.

The Hanging Gardens vegetation type described by Harper et al. (2001) incorporated additional data from published studies to supplement the hanging garden plots captured by the BYU-RMRS survey. Twenty-one hanging gardens described by Malanson and Kay (1980) and Welsh (1989) were treated as plots for characterizing this vegetation type (Harper 1993, Harper et al. 2001). These hanging garden data were excluded from the current analysis, which used only the 288 standard-sized plots of the BYU-RMRS survey.

USGS-NPS Vegetation Characterization and Mapping (Cogan et al. 2004)

Another round of extensive vegetation sampling at Zion took place in 1999-2000 through a partnership of the USGS, U.S. National Park Service (NPS), U.S. Bureau of Reclamation and NatureServe (Cogan et al. 2004). New vegetation plot data were deemed necessary to meet the standards of the USGS-NPS Vegetation Mapping Program which aims to apply the U.S. National Vegetation Classification System (NVCS) to all U.S. National Park units (USGS 2010). Zion was among the first parks to be targeted by this program because the program's products were in demand by park managers (Cogan et al. 2004). The plot data, vegetation descriptions, GIS layers and other products from this program (USGS 2004) proved useful in the current study for comparative and interpretive purposes.

The USGS-NPS vegetation survey sampled 346 circular, square or rectangular plots (Fig. 2.2) ranging from 100 m² to 400 m² where plant species were identified

following Kartesz (1999) (USGS 2004, Cogan et al. 2004). A gradsect strategy (Gillison and Brewer 1985) was used to divide Zion into biophysical units (BPUs) that were mapped and targeted for field sampling. Each of the 70 BPUs mapped at Zion was defined by a unique combination of categorical environmental variables (two hydrology, two fire history, four solar insolation and nine geology/elevation categories) developed as GIS layers specifically for this purpose (Cogan et al. 2004). Accessible polygons of each BPU were targeted by field survey teams who ultimately placed plots on sites deemed representative of major vegetation types and relatively homogeneous in terms of vegetation structure and composition (Cogan et al. 2004). GPS coordinates were recorded for each plot.

Cogan et al. (2004) used the USGS-NPS plots to classify and map the vegetation of Zion following the NVCS. Multivariate analysis of species data, including cover class by strata recorded for each species, helped guide the classification process (Cogan et al. 2004). Each plot was assigned to an association (the standard lowest level of the NVCS hierarchy) nested within an alliance and other higher hierarchical levels (formation, subgroup, group, subclass, and class). Association descriptions (Cogan et al. 2004, Appendix F) include references to “characteristic species” and “associated species” that can be interpreted as constant and/or faithful species, although the basis for their derivation is not explicitly explained.

In conjunction with plot sampling and classification, aerial photo interpretation techniques were used to map vegetation types across the entire park, guided by the areal orthophoto signatures associated with each plot (Cogan et al. 2004). Mappers sought to follow the NVCS with as much resolution as possible but not all vegetation was

discernable or separable at the association level. The final vegetation map, published in digital format, contains 76 map units: 23 associations (including one association separated into two phases), six alliances, two mosaics (associations too intermixed to separate), 16 complexes (associations too similar to distinguish), and 29 miscellaneous units (unclassified vegetation and non-vegetated areas) (Cogan et al. 2004). This discrete vegetation map provided a framework for comparison with the individualistic species group environmental affinity maps of the current study.

INDIVIDUALISTIC COMMUNITY CHARACTERIZATION: CONCEPTS AND METHODS

Conceptual Overview

The objective of the present analysis is to re-characterize plant communities at Zion National Park through an individualistic approach based on spatially-diffuse species groups rather than discrete vegetation units. I begin my description of this approach with a brief conceptual summary.

Species groups, defined broadly as species belonging to multiple life forms with overlapping distributions across community samples, are conceptually similar to faithful species. Members of a species group are ‘faithful’ to each other across an entire study area, but not necessarily faithful to discretely-defined communities within the area. This conceptual parallel provides the rationale for substituting faithful species with species groups in the individualistic approach presented here. The approach exchanges the need for identifying discrete community units with the need for discrete species groups, but also acknowledges individualistic variation of species by quantifying their affinity (degree of association) relative to a group. *Affinity* to a species group replaces *fidelity* to

a community unit in the individualistic species group approach. Furthermore, the approach allows species to belong to and/or have high affinities to more than one species group, which differs somewhat from conventional thinking about community fidelity but is compatible with the individualistic concept.

The individualistic species group approach also incorporates a conceptual equivalent of constant species. Whereas *constancy* is normally quantified as the frequency of *occurrence* of a species within a *community unit*, the analogous *affiliation* value presented here is the frequency of *co-occurrence* of a species with members of a *species group*. Constancy is an inherently individualistic measure, in the sense that it is calculated for each community unit independently and hence a species can be constant in multiple communities. In the same way, species can be independently affiliated with multiple species groups in the approach followed here.

An important feature of the individualistic species group approach is that it allows spatial community variation to be represented as continuous fields of species group affinity. Because species groups are spatially-diffuse patterns of multiple species across multiple sites, individual sites (e.g. community sample units) generally do not correspond exactly to any given species group. A given site may house many members a given species group, or few members, or a mixture of species with different degrees of affinity to the group. Such differences can be quantified by averaging the affinity values of the species present at a site (essentially a form of weighted averaging; Kent and Coker 1992, McCune and Grace 2002). Community sample units can thus be placed along a continuum of affinity values for each species group, congruent with Gleason's (1926, 1939) individualistic concept in relation to sites. This approach opens the door for

quantitative spatial and environmental analyses of species group affinities. In the application presented here, relationships between site-averaged species group affinities and environmental variables are modeled using regression tree analysis (Breiman 1993). This leads to descriptions of *environmental affinities* of species groups, in which the term *affinity* again carries a specific quantitative meaning, in this case denoting the environmental conditions of sites where species group affinity is comparatively high.

A final point is that individualistic species groups can be compared with discrete community units wherever both have been characterized for the same study area. One way of quantifying the correspondence between a species group and a community unit is by averaging the group affinity values of species attributed to the unit (an extension of the weighted averaging technique described above). Affinity can be averaged for all species occurring in plots belonging to the unit, or more specifically for the unit's faithful species. Spatial models of species groups and community units can also be compared. A combination of these procedures was used in the comparisons presented here.

Coalition Clustering and Affinity Calculations (Individualistic Fidelity Analysis)

Coalition clustering, a non-hierarchical agglomerative clustering algorithm included in the program RCLUS (Sanderson et al. 2006) provided a means for identifying the individualistic equivalent of faithful species at Zion without having to define community units. RCLUS was developed as a tool for R-mode (species-based) clustering of community datasets. The name *coalition clustering* underscores the individualistic variability of species within the clusters ('coalitions') that the algorithm builds. Coalition clustering generates clusters of positively associated species here referred to as *coalition*

species groups or simply *coalition groups*. The term *core species*, used by Sanderson et al. (2006) to refer to species belonging to a group, is here replaced by the term *coalition species*.

Different coalition clustering solutions can be obtained by varying three user-defined parameters of the algorithm: association coefficient, threshold association value, and minimum number of occurrences for species inclusion in groups (see below). In addition, clustering solutions may vary from one run of the algorithm to another because of the random nature of the agglomerative process. The clustering solution presented here was selected from among a small number of variants obtained using the phi coefficient at a threshold value of 0.15, and allowing species occurring in at least three plots to be included in groups. These parameter values yielded eleven coalition groups, providing a numerically-congruent alternative to the ten vegetation types previously described for Zion by Harper et al. (2001). Dr. S.C. Sanderson, author of the RCLUS program and former field crew leader for the BYU-RMRS survey, played a key role in analyzing and selecting this clustering solution.

Coalition Clustering in RCLUS begins with a matrix of association between species pairs. Association coefficients measure the degree to which two species show the same pattern of presence and absence in a dataset, calculated from the cells of a two-way presence-absence contingency table:

		Species 1	
		Present	Absent
Species 2	Present	a	b
	Absent	c	d

The phi coefficient incorporates all four cells of the table according to the formula:

$$\text{phi coefficient} = \frac{ad - bc}{\sqrt{(a+b)(a+c)(b+d)(c+d)}}$$

Phi ranges from -1 (negative association) to +1 (positive association) and is independent of the total number of occurrences in the table (Jackson et al. 1989, Tichý and Chytrý 2006). This is a useful property for measuring association, but makes the phi coefficient ineffective at measuring statistical significance (unlike the related chi-square). If one or both species has low occurrence in the dataset, the statistical significance of the association may be in question, but association is still quantifiable. The RCLUS user's option of excluding species of low frequency from cluster analysis is important for this reason.

Clustering proceeds by agglomerating species whose mean association is higher than the user-defined threshold value. That is, when pairwise association values are averaged for all species pairs in a group, the average must exceed the threshold for the group to be accepted. The algorithm initiates the first group with a randomly-selected species and then randomly cycles through the remaining species, adding them to existing groups or initiating new groups as appropriate. Coalition Clustering allows species to belong to multiple groups with the exception that species already present in an existing group cannot initiate a new group. The algorithm undergoes several cycles of adding and removing species allowing the composition of species groups to stabilize. By defining a critical threshold for mean association within groups, the user determines how loose or tight the groups are allowed to be. High thresholds tend to produce many small groups in contrast to few large groups at low thresholds (Sanderson et al. 2006).

Once the Coalition Clustering algorithm is complete, RCLUS saves the mean association value of each species to each group (Sanderson et al. 2006). These mean

association values, or *affinity values*, provide a quantitative measure of the relationship between individual species and coalition groups.

Affiliation Calculations (Individualistic Constancy and Dominance Analysis)

RCLUS also contains a function that calculates *percent co-occurrence* (previously named *mean percent*) of each species to each coalition group. Percent co-occurrence is calculated by summing joint presences across all pairings of a focal species with the species comprising a coalition group and then standardizing by the maximum possible number of joint presences. Using the notation of presence-absence frequencies, this can be represented as follows:

$$\begin{array}{ccccc}
 & & \text{Focal Species} & & \\
 & & \textit{Present} & \textit{Absent} & \\
 \text{Coalition Species} & \textit{Present} & a & b & \\
 \end{array}$$

$$\text{percent co - occurrence} = \frac{\sum_{sp1}^{spN} a}{\sum_{sp1}^{spN} (a+b)}$$

Restated using this notation, percent co-occurrence of the focal species to the coalition group is the sum of joint presences between the focal species and each of the coalition species in the group ($sp1$ to spN), standardized by the total number of occurrences of all the coalition species.

The intuitive term *affiliation* is here used interchangeably with percent co-occurrence. Species belonging to a group are naturally likely to have high affiliation with their own group. However, correspondence between high group affinity and high affiliation need not be exact because the former measures specificity (fidelity) while the

latter measures commonness (constancy). A widespread generalist species can be highly affiliated with multiple groups without demonstrating high affinity for any of them.

Building on Sanderson's RCLUS analysis of the BYU-RMRS dataset, I added a step to extract a set of highly affiliated species ('affiliate species') for each coalition group. Since Curtis (1959) used species density, the mean number of species per plot in a community, to determine how many species to include in prevalent species lists, I followed an analogous procedure for determining the number of affiliate species for each coalition group. The species density of a coalition group was calculated as the mean number of species occurring in plots where coalition species occurred, weighting plots by how many coalition species they contained. Defining S as the species density of a coalition group, affiliate species are the S species with highest affiliation to the group.

Extending the affiliation concept further to include abundance as well as occurrence, I calculated the abundance of each affiliate species as its mean cover across plots containing coalition species, weighted once again by the number of coalition species per plot. Arithmetic midpoints of cover classes were used in these calculations. In this way I was able to identify the dominant species associated with each coalition group.

Environmental Modeling of Coalition Species Group Affinities

For each plot used in coalition clustering, RCLUS calculates the plot's affinity to each coalition species group as the average affinity value of the species recorded in the plot. Both coalition and non-coalition species contribute to these averages. I took these plot-averaged affinity values and used them to model correlative relationships between

coalition groups and five environmental variables: elevation, slope, topographic exposure, topographic position, and geologic substrate (Fig. 2.3-2.4). These variables were selected because of they are easily-measured proxies for the major physical and chemical gradients affecting plant distributions at Zion. They were available or easily created as geographic information system (GIS) layers that were integrated with plot data in ArcGIS (ESRI 2003, 2004). My use of GIS-based environmental variables permitted predictive mapping of species group affinities beyond plot locations (Ferrier et al. 2006).

I manually digitized plot coordinates from the BYU-RMRS survey, originally marked on paper maps, with assistance from the primary field crew leader from the survey (S.C. Sanderson) using as references a high-resolution aerial orthophoto of Zion National Park (Cogan et al. 2004), digital USGS topographic maps and cadastral survey marker coordinates acquired through the USDI Bureau of Land Management (BLM 2006). Hawth's Tools, an ArcGIS add-in program (Beyer 2004), was then used to extract environmental data values for each plot based on its coordinates.

Two GIS layers served as sources of environmental data: a 10 m resolution digital elevation model (DEM) and a 1:24,000 scale digital geologic map built from multiple maps of Utah Geologic Survey (O'Meara 2006). These digital layers and accompanying metadata are now archived in the NPS data store (NPS 2010). They provide full coverage of Zion with the exception of the geologic map layer which omits ca. 1.3 km² along the western inner corner of the park. Although seven plots of the BYU-RMRS survey lie outside this layer, geologic units were easily assigned to these plots through visual interpolation from the edge of the geologic layer and examination of plot descriptions. These plots could thus be used in environmental modeling although the

area missed by the geologic layer was ultimately excluded from predictive maps. The geologic layer was converted to raster format at the same resolution as the DEM prior to modeling and mapping.

Elevation was taken directly from the DEM, while slope and exposure were calculated using the *slope* and *hillshade* functions, respectively, in ArcGIS 9.0 Spatial Analyst (ESRI 2004). *Hillshade* was implemented at 215° azimuth and 45° altitude such that exposure values were highest on steep southwest-facing slopes where solar heat loading is expected to be highest (cf. McCune and Keon 2002). Topographic position was defined using the Topographic Position Index (TPI) developed by Weiss (2001). TPI was calculated for each pixel of the DEM by subtracting the mean elevation of surrounding pixels (within a defined radius) from the focal pixel's elevation (Weiss 2001, Jenness 2006). Negative TPI values indicate topographic lows such as canyon bottoms and positive values indicate topographic highs such as peaks and ridges. The ArcGIS 9.0 raster calculator (ESRI 2004) was used to calculate TPI at 50 m and 100 m radius.

A categorical variable map comprised of 16 geologic substrate classes (Fig. 2.4) was created through aggregation of the ca. 100 map units occurring within Zion according to O'Meara (2006). Most of the map units of O'Meara (2006) are individual members of geologic formations or descriptive categories for local Quaternary deposits. Given the need to simplify this map for modeling purposes, but acknowledging the many possible ways of merging map units, I settled on a pragmatic approach using lithology, spatial continuity and/or topographic characteristic as grouping criteria (Table 2.1, Fig. 2.4). Exploratory analyses (boxplots of coalition group affinities by substrate type) revealed that the eight major geologic formations of the park (Moenkopi, Chinle,

Moenhave, Kayenta, Navajo, Temple Cap, and Carmel), though internally heterogeneous to various degrees, were likely to be useful categories for modeling community-environment relationships at Zion. Thus each formation was assigned its own class, except for poorly-represented ones at the lower end of the stratigraphic sequence (Kiabab) and upper end (Cedar Mountain) that were merged with the adjacent Moenkopi and Carmel formations, respectively. Exploratory analyses also supported a scheme in which Quaternary deposits were grouped according to their dominant depositional material (alluvium, colluvium, eolian, lacustrine, mass movement, residuum, and volcanics). The well-represented mass movement deposits were further split into three classes that occupy different topographic settings in the park: (1) talus, (2) slides/slumps/flows and (3) mixed (mass movement/colluvium/ alluvial pediment mantle) deposits (Table 2.1, Fig. 2.4). Map units of O'Meara (2006) included in each of these classes are shown in Table 2.1; note that some map units were not actually sampled by plots of the BYU-RMRS dataset but were assigned to the closest logical class.

Relationships between species group affinities and environmental variables were modeled using regression tree analysis (the `tree` function) in SPLUS 7.0 (Insightful Corp. 2005). Regression tree analysis generates a dichotomous tree model by splitting a dataset into progressively smaller subsets (Breiman 1993, Crawley 2007). Each split, represented by a node in the tree, segregates data points based on an ordinal relationship between a response variable and a predictor variable. The tree algorithm automatically selects the predictor variable with the strongest relationship at a given node if multiple predictor variables are supplied. Plot-averaged group affinity (in units of the phi coefficient) was the response variable and substrate class, elevation, slope, exposure, and

topographic position (TPI at radius=50 m and radius=100 m) were the predictor variables used to build regression tree models for each coalition group. Models were built using consistent splitting rules (mincut=5, minsize=10, mindev=0.1). Multiple runs of random 10-fold cross-validation (the `cv.tree` function) were applied to each model to determine the best number of terminal nodes minimizing total deviance (Crawley 2007), and models were pruned accordingly.

Quantifying Correspondence with Previously-described Vegetation Units

In order to assess the correspondence between coalition groups identified through RCLUS analysis and vegetation types reported by Harper et al. (2001), results from both analyses were merged into a common database. Modal species listed for each vegetation type in Harper et al. (2001) were joined to their group affinity values from the RCLUS output, then averaged for each vegetation type/coalition group comparison. Two modal species (*Carex curvata* and *Lobelia cardinalis*) were restricted to the supplemental hanging garden dataset used only by Harper et al. (2001) and were thus omitted from these calculations. All other species could be matched because of their common data source (the BYU-RMRS dataset).

I was also interested in quantifying the correspondence between coalition groups and NVCS associations reported by Cogan et al. (2004). Because modal species (or other types of faithful species) were not reported for these associations, I based my analysis on species composition of the raw plot data. That is, species recorded in plots of the USGS-NPS dataset were joined to coalition group affinity values, which were then averaged across all species occurrences for each association. This yielded association-averaged

affinity values similar to the plot-averaged affinity values described previously (see “Environmental Modeling of Coalition Species Group Affinities”). As part of this process, taxonomic concepts in the USGS-NPS dataset were standardized to match Welsh et al. (2003). Following taxonomic standardization, 185 taxon records (including 71 taxa identified only to the genus level) remained unmatched because they were unique to the USGS-NPS dataset (Appendix 2.1); and conversely, 237 taxa captured by the BYU-RMRS survey were not represented in the USGS-NPS dataset (Appendix 2.2).

Mean coalition group affinities were also calculated for each mapped vegetation unit of Cogan et al. (2004). Using the zonal statistics tool in ArcGIS, pixel values from each species group affinity map were averaged for each vegetation unit.

RESULTS

Coalition Species Groups and Their Environmental Affinities

Eleven coalition species groups were identified through coalition clustering at a threshold value of phi=0.15. The eleven groups contain a total of 174 species extracted from the 511 species recorded by the BYU-RMRS vegetation survey (Appendix 2.2; updated to nomenclature of Welsh et al. 2003). One group containing six species (*Artemisia dracunculus*, *Elymus spicatus*, *Sisymbrium altissimum*, *Galium aparine*, *Physalis hederifolia*, and *Tragopogon dubius*) was excluded from further analysis because of its limited distribution in disturbed low-elevation environments. The remaining ten groups containing 9-36 species each (Table 2.2) can be readily described in terms of their environmental affinities at Zion (Fig. 2.5-2.14). I have given each species group a name that describes the environmental setting for which it demonstrates greatest

affinity. Coalition species of each group are presented in descending order of affinity in Table 2.2 (left column) and an expanded table showing affinities of all species to all coalition groups is provided in Appendix 2.2. Coalition clustering's allowance for overlapping group membership resulted in 34 cases of species belonging to two groups and one case of a species (*Amelanchier utahensis*) belonging to three. Environmental affinities of coalition groups also overlap to varying degrees, as depicted in the predictive maps accompanying regression tree models in Figures 2.5-2.14. In the group descriptions that follow I highlight these cases of compositional and spatial overlap and interpret their ecological significance.

The Streambank group (Table 2.2A) contains several species, including *Baccharis emoryi*, *Salix exigua*, *Tamarix chinensis* and *Populus fremontii*, with clear affinities to low-elevation riparian zones at Zion. Regression tree analysis confirmed this inference by identifying low topographic positions [TPI (50 m radius) < -0.51] below 1356 m on alluvium or lacustrine substrates as the environmental conditions of highest affinity (Fig. 2.5). On substrates beyond alluvium and lacustrine, topographic lows continued to show higher affinities, but high elevations (>2321 m) also had relatively high affinities, presumably because mesic conditions at Zion's highest elevations are favorable for some riparian species. Indeed, the High Plateau group, which has highest affinity at elevations above 2243 m (Fig. 2.6), shares three coalition species (*Bromus carinatus*, *Elymus trachycaulus* and *Poa pratensis*) with Streambank (Table 2.2A-B). Coalition species unique to the High Plateau group include *Stellaria jamesiana*, *Lupinus sericeus*, *Prunus virginiana*, *Stipa lettermanii*, *Rosa woodsii*, and *Juniperus scopulorum*. (Table 2.2B).

The Crevice Canyon group (Table 2.2C, Fig. 2.7) occupies an environmental setting intermediate between Streambank and High Plateau, namely the bottom of narrow, mid-elevation Navajo Sandstone canyons where topographic shading and intermittent water flow produce cool mesic conditions. Sites with these conditions have extremely low TPI values; TPI (50 m radius) is less than -35 in the zone where Crevice Canyon affinities are highest (Fig. 2.7). Outside of these extreme topographic lows, the Crevice Canyon group has higher affinity for higher elevations (Fig. 2.7). This affinity pattern is mirrored by High Plateau's inverse pattern of high affinity for high elevations, intermediate affinity for narrow mid-elevation canyons (Fig. 2.6). The canopy dominant species *Abies concolor* as well as two herbs (*Thalictrum fendleri* and *Taraxacum officinale*) are coalition species of both Crevice Canyon and High Plateau (Table 2.2B-C). In addition, the Crevice Canyon group shares four species (*Acer negundo*, *Elymus canadensis*, *Equisetum hymenale* and *Agrostis exarata*) with the Streambank group (Table 2.2A) and five (*Heuchera rubescens*, *Zauschneria latifolia*, *Holodiscus dumosus*, *Selaginella underwoodii*, *Brickellia grandiflora*, and *Erigeron sionis*) with Slickrock (Table 2.2D). Crevice Canyon with 36 species is the largest of the coalition groups and many of its species were relatively uncommon with only 3-6 occurrences in the BYU-RMRS dataset (e.g., *Viola canadensis*, *Cystopteris fragilis*, *Polypodium hesperium*, *Galium triflorum*, *Rubus leucodermis*, *Mimulus guttatus*, *Disporum trachycarpum*) while others were more common with 10-29 occurrences (e.g. *Eupatorium herbaceum*, *Acer grandidentatum*, *Bromus ciliatus*, *Pseudotsuga menziesii*, *Philadelphus microphyllus*) (Table 2.2C).

Species associated with exposed Navajo Sandstone outcrops are represented in the Slickrock group which includes *Petrophytum caespitosum*, *Cercocarpus intricatus*, *Castilleja scabrida*, *Muhlenbergia thurberi* and *Ivesia sabulosa* (Table 2.2D). The Slickrock group had highest affinity on the Navajo Sandstone, especially above 1782 m, and had intermediate affinity on Carmel, Temple Cap, colluvium, residuum, and talus substrates (Fig. 2.8). Regression tree analysis further split the latter set of substrates based on topographic position revealing higher affinity at topographic lows [TPI (100 m radius) < -21] (Fig. 2.8). These topographic lows correspond to talus deposits immediately below Navajo Sandstone cliffs where sandstone substrate characteristics are to be expected. The Temple Cap formation immediately above the Navajo Sandstone is also primarily sandstone and would be expected to have similar plant species because of both substrate similarity and elevational proximity. On the other hand, the Slickrock group did not have high affinity for eolian deposits despite elevational proximity and common parent material.

Eolian deposits at Zion are composed primarily of sand and are especially extensive on the southeastern side of the park (O'Meara 2006, Fig. 2.4). Smaller pockets of sand associated with hollows and flat areas on Navajo Sandstone outcrops are scattered throughout the park. Many of these sand pockets were too small to be included on Utah Geologic Survey maps (i.e. O'Meara 2006) but were captured by plots of the BYU-RMRS survey. These plots contributed to the Upland Sands coalition group, which is characterized by sand-loving species such as *Cryptantha cinerea*, *Abronia fragrans*, *Artemisia campestris*, *Penstemon laevis*, and *Tradescantia occidentalis* (Table 2.2E). The shrub dominant *Arctostaphylos patula* is a coalition species of the Upland Sands

group, while the tree dominant *Pinus ponderosa* is a coalition species of both Upland Sands and Slickrock, along with *Arenaria fendleri*, *Chrysopsis villosa* and *Linanthastrum nuttallii* (Table 2.2D-E). The regression tree model for the Upland Sands group (Fig. 2.9) is similar to the Slickrock model (Fig. 2.8), identifying higher-elevation (>1753 m) Navajo Sandstone as the zone of highest affinity. The seemingly high overlap of these models can be clarified by examining environmental data recorded at the plot scale: among plots mapped to Navajo Sandstone, Slickrock affinity was positively correlated with percent rock cover (Spearman's rho=0.50) while Upland Sands affinity was negatively correlated (Spearman's rho=-0.15). The Upland Sands group also differed from Slickrock in its high affinity for eolian deposits and its lower affinity for canyon bottoms and cliff bases within the Navajo/Temple Cap/eolian zone (Fig. 2.8-2.9). However, below this zone on the southwestern side of Zion the Upland Sands group had higher affinity for slopes adjacent to Navajo Sandstone cliffs than more distant lowlands below 1423 m (Fig. 2.9).

The coalition group with highest affinity for the steep sloping terrain below Navajo Sandstone cliffs was given the name 'Rocky Slopes'. *Fraxinus anomala*, *Erigeron utahensis*, *Poa fendleriana*, and *Quercus turbinella* are prominent members of this group (Table 2.2F). The first split of the Rocky Slopes regression tree model identifies substrate as an important affinity predictor, with lower affinity for Carmel, alluvium, colluvium, eolian, lacustrine and residuum than for other substrates (Fig. 2.10). This data split may actually reflect the influence of slope more than substrate characteristics *per se*, given that this set of six substrates has a lower mean slope (22°) than the remaining ten substrates (61°). Rocky Slopes affinities are lowest at the park's

lowest elevations (<1327 m) and highest elevations (> 2321-2368 m) (Fig. 2.10) which suggests a mid-elevational optimum for this species group but may also reflect the fact that terrain at both these elevational extremes is relatively flat. Rocky Slopes affinity is also low in the deep canyon zone (TPI at 100 m radius < 35) (Fig. 2.10) where Crevice Canyon affinity is high (Fig. 2.7) but remains high in other topographic lows adjacent to cliffs. The highest Rocky Slopes affinities are associated with the Kayenta formation which is particularly extensive in the Taylor Creek thrust fault area of northwestern Zion (Fig. 2.10). The broad zone of relatively high affinity above, below and around the Kayenta formation can be attributed to the broad distribution of several members of the Rocky Slopes group, including *Poa fendleriana*, *Amelanchier utahensis* and *Arabis perennans*. These species occupy a wide range of environments at Zion but form a coalition with more narrowly distributed species such as *Shepherdia rotundifolia* (Table 2.2F).

The Rocky slopes group shares three coalition species (*Opuntia macrorhiza*, *Senecio multilobatus* and *Erysimum asperum*) with the Upland Sands group and three others (*Pinus monophylla*, *Juniperus osteosperma* and *Gilia inconspicua*) with the lower-elevation Arid Lowlands group (Table 2.2G). *P. monophylla* and *J. osteosperma* are well-known dominants of Great Basin woodlands and epitomize the Great Basin affinities of many of the Arid Lowlands coalition species. Other coalition species of this group, including *Coleogyne ramossima*, *Baileya multiradiata* and *Eriastrum eremicum*, have geographic affinities for the Mojave Desert and/or lower-elevation Colorado Plateau rather than the Great Basin (Welsh et al. 2003). Included in this mixture is the common Mojave Desert invasive species *Bromus rubens* as well as its Great Basin counterpart

Bromus tectorum, *Gutierrezia* spp. (*G. sarathrae* and *G. microcephala*), indicators of grazing disturbance history (Welsh et al. 2003), also have high affinity to the Arid Lowlands group (Table 2.2G).

The arid character of sites occupied by the Arid Lowlands group is verified by its regression tree model which indicates higher affinity for sites below 1596 m elevation, precluding sites such as deep canyons with TPI (100 m radius) less than -22 (Fig. 2.11). Within this low-elevation zone the Arid Lowlands group has lower affinity for substrates associated with riparian areas (alluvium and lacustrine) as well as substrates concentrated at higher elevations (Kayenta, talus, Navajo and eolian) except where these substrates are highly exposed (exposure>215). At elevations between 1596-1818 m the Arid Lowlands group likewise has higher affinity for more exposed sites (exposure>130) (Fig. 2.11) where the moisture/temperature regime is presumably similar to lower elevations.

Another low-elevation species group named ‘Lowland Flats’ had high affinities for sites below 1388 m, especially alluvium and Moenkopi substrates with TPI (50 m radius) greater than -0.5 (Fig. 2.12). In Zion Canyon these conditions correspond with floodplains and benches above the zone where the Streambank group is concentrated (Fig. 2.5). These portions of Zion Canyon have been subject to intensive agriculture in the past and bear the marks of this land use history. A combination of native species (e.g., *Lycium pallidum*, *Atriplex canescens*, *Sphaeralcea grossularifolia*, *Elymus smithii*) and exotics associated with disturbance (*Erodium circutarium*, *Tragopogon dubius*, *Lactuca serriola*) typified the Lowland Flats group (Table 2.2H). Various combinations of these species also occurred throughout the low elevation zone in Zion. Above 1388 m

elevation, higher affinities were associated with flatter areas (slope <6°) on mostly unconsolidated substrates (Fig. 2.12).

Coalition Clustering yielded two groups associated with the Carmel formation that caps the Navajo Sandstone on the eastern and northern sides of Zion. One group (Mesic Mesa Top) is interpreted as indicating slightly more mesic settings than the other group (Xeric Mesa Top). Highest affinities for both groups are found in a zone defined by the Carmel formation above 1791 m, but outside of this zone the Mesic Mesa Top group's affinities are shifted toward higher elevations than Xeric Mesa Top's (Fig. 2.13-2.14). The Mesic Mesa Top group had high affinities for the residuum deposits that in many places cap the Carmel, but lower affinities for the Navajo and eolian substrates that occur below the Carmel, and lowest affinities below 1486 m (Fig. 2.13). The Xeric Mesa Top group, in contrast, did not have exceptionally high affinity for residuum, did not have lower affinity for Navajo or eolian than for other non-Carmel substrates, and reached its lowest measurable affinity only at exposed sites (exposure>129) below 1430 m (Fig. 2.14).

Although the Mesic and Xeric Mesa Top groups share a set of prominent woody species (*Pinus edulis*, *Quercus gambelii*, *Amelanchier utahensis* and *Peraphyllum ramosissimum*) as well as two herbaceous species (*Carex rossii* and *Swertia radiata*), the affinity order of these species is almost exactly reversed for the two groups (Table 2.2I-J). *P. ramosissimum* and *S. radiata* have higher affinity for Mesic Mesa Top than *P. edulis* and *C. rossii* (Table 2.2I) while the opposite is true for Xeric Mesa Top (Table 2.2J). Other species that do not overlap these groups include *Viola purpurea*, *Cercocarpus montanus*, *Pachystima myrsinites* and *Mahonia repens* of Xeric Mesa Top

and *Lathyrus lanszwertii*, *Balsamorhiza sagittata*, *Phlox longifolia*, *Penstemon linarioides* and *Purshia tridentata* of Mesic Mesa Top (Table 2.2I-J). Only two members of these groups overlap with other groups: *Symporicarpos oreophilus* links Mesic Mesa Top to the High Plateau group (Table 2.2B) and *Amelanchier utahensis* links both Mesa Top groups to the Rocky Slopes group (Table 2.2F).

Affiliate Species of Coalition Groups

Species with the highest affiliation to coalition groups (affiliate species) corresponded only loosely to the species with highest affinity (coalition species) (Table 2.2, Appendix 2.2), reflecting the different patterns emphasized by affiliation and affinity values. Affiliation (percent co-occurrence) was much more a reflection of a species' total frequency than affinity (mean phi). With the exception of the Streambank and Rocky Slopes groups, less than half each group's affiliate species were also coalition species. The number of affiliate species (species density: a reflection of species richness in plots where coalition species are concentrated) ranged from 21 for the Xeric Mesa Top group to 27 for Crevice Canyon (Table 2.2). Although the number of affiliate species per group was in most cases higher than the number of coalition species, the total number of affiliate species (101) was lower than the coalition species total (173), reflecting the greater inter-group overlap of affiliate species.

As a general pattern, the most widely-distributed species had the highest affiliation values and were affiliated with the most coalition groups. *Poa fendleriana*, the most frequently encountered species at 190 occurrences, was an affiliate of all ten described groups. Other widely-distributed species with multiple affiliations include

Quercus gambelii, *Amelanchier utahensis*, *Bromus tectorum*, *Arabis perennans*, *Senecio multilobatus* and *Opuntia macrorhiza*. In each of these cases the species was affiliated with several additional groups beyond the one(s) where it belonged as a coalition species. Conversely, coalition species with few occurrences were generally affiliated with fewer groups, in many cases none at all. Some moderately frequent species, including *Artemisia ludoviciana*, *Artemisia tridentata*, *Elymus elymoides*, *Eriogonum racemosum*, *Machaeranthera canescens*, *Phacelia heterophylla*, *Sporobolus cryptandrus*, *Stipa comata*, *Stipa hymenoides* and *Yucca angustissima*, have the distinction of emerging as affiliate species without belonging to any of the coalition groups. The latter is also true of *Artemisia* sp., *Melilotus* sp. and *Oenothera longissima* which were excluded from entering coalition groups because of having only two occurrences, but nevertheless emerged as affiliates of the Streambank group.

Weighted mean cover ranged from a fraction of a percent for most affiliate species (Table 2.2) to 23 percent for *Quercus gambelii* with the High Plateau group (Table 2.2B), followed closely by *Bromus tectorum* with the Lowland Flats group at 18 percent (Table 2.2H). These percentages are lower than might be expected based on the maximal cover recorded for these species in these environments, but reflect average cover across plots where coalition species were concentrated. Although cover values in Table 2.2 are not strictly comparable with conventional discrete plot averages, they can be used in a similar way to quantify relative differences in species dominance. Species with a mean cover value of two percent or more could be construed as dominant species and are underlined in Table 2.2. Most of these species were also recognized as dominants in previous analyses of the BYU-RMRS dataset (Harper 1993, Harper et al.

1993, 2001, 2003), although dominant species are not as clearly segregated among coalition groups as they were for previously-defined vegetation types. For example, *Pinus ponderosa*, which defined the Ponderosa Pine vegetation type in Harper et al. (2001), emerged as a dominant affiliate species of four coalition groups (Xeric and Mesic Mesa Top, Upland Sands and Slickrock). Many of the dominant species at Zion appear to be generalists that can achieve dominance in a variety of different environments with a variety of different subordinate species.

Correspondence with Previously-described Vegetation Units

To a certain extent, the ten species groups obtained through coalition clustering mirrored the ten vegetation types previously described by Harper et al. (2001). Table 2.3 summarizes the correspondence between species groups and vegetation types in terms of group affinities of modal species described for each type (see also Appendix 2.3). In Table 2.3, vegetation types (rows) and species groups (columns) are arranged such that high values follow roughly the diagonal.

Some species groups were compositionally very similar to modal species of specific vegetation types, e.g. seven of the nine coalition species of the Lowland Flats group were modal species of the Abandoned Fields type (Table 2.3, Appendix 2.3). Other species groups and modal species sets resembled one another but the correspondence was not as strongly one-to-one; e.g. Arid Lowlands and Blackbrush, Slickrock and Rock Crevice, High Plateau and Douglas/White Fir, Streambank and Riparian (Table 2.3). The Upland Sands group had high correspondence with three vegetation types (Ponderosa Pine, Mountain Brush, and Rock Crevice) as did Crevice

Canyon (Riparian, Douglas/White Fir, and Hanging Gardens) and the Mesa Top groups (Douglas/White Fir, Ponderosa Pine and Juniper-Pinyon). The Sagebrush vegetation type had relatively weak affinities with the Arid Lowlands group compared to the stronger affinities between the latter and Blackbrush.

Few of the rarer species at Zion were listed as modals in Harper et al. (2001) because of the requirement that modal species also be prevalent. Also, the modal species concept followed by Harper et al. (2001) did not accommodate species modality in multiple vegetation types, unlike the coalition clustering approach which highlighted bimodal and (in the case of *Amelanchier utahensis*) tri-modal memberships in species groups. While these methodological details account for some of the differences between Harper et al. (2001) and the results presented here, other differences can be attributed to the fundamentally different ways in which the two approaches partitioned species distribution variation at Zion. The discrete, dominant-species based classification followed by Harper et al. (2001) masked some of the floristic distinctions and gradations captured by the species group approach, such as the subtle floristic contrast between the Xeric and Mesic Mesa Top groups and the floristic continuity of the Arid Lowlands group.

Because dominant species and growth forms such as ponderosa pine, juniper-pinyon and mountain brush occupy multiple environmental settings at Zion, their use as classification criteria by Harper et al. (2001) led to environmentally ambiguous vegetation types. Some of these ambiguities were clarified in a subsequent manuscript (Harper et al. 2003) that split the juniper-pinyon vegetation type into three subtypes: a higher elevation subtype containing *Pinus edulis* (like the Mesa Top groups), a mid-

elevation subtype with *Pinus monophylla* (like Rocky Slopes) and low-elevation subtype with *Juniperus osteosperma* but no *Pinus* (like Arid Lowlands). Cogan et al. (2004) differentiated pinyon-juniper vegetation in a similar way at the alliance level (Appendix 2.4).

The NVCS associations and alliances described for Zion by Cogan et al. (2004) are more finely divided than the vegetation types of Harper et al. (2001), although they can be lumped into a comparable number of higher units. The NVCS hierarchy provides one way of lumping associations and alliances based on a set of standardized criteria. The highest NVCS level (formation class), based on gross physiognomy, is represented at Zion by six classes: forest, woodland, shrubland, dwarf shrubland, herbaceous and sparse vegetation. As shown in Table 2.4, I used these classes to generalize compositional affinities between coalition groups and NVCS associations, which are presented in greater detail in Appendix 2.4.

Table 2.4 can be interpreted in two ways; on one hand, it indicates the relative proportions of different physiognomies associated with each coalition group, and on the other hand it summarizes the environmental affinities of the associations in each physiognomic class. From the latter perspective, the forest class has higher elevation/mesic affinities (e.g., High Plateau, Crevice Canyon, Streambank) while woodland, shrubland, dwarf-shrubland and sparse vegetation have lower elevation/xeric affinities (Rocky Slopes, Arid Lowlands, Lowlands Flats) and the affinities of herbaceous vegetation follow their own pattern (Streambank, Rocky Slope, Arid Lowlands). Upon closer examination of individual associations (i.e. Appendix 2.4), one finds that these general patterns belie considerable variation. Shrublands, for example, include high

elevation associations such as Mountain Snowberry/Kentucky Bluegrass and Strapleaf Willow/Beaked Sedge as well as the expected low elevation units such as Blackbrush and Four-wing Saltbush (Appendix 2.4).

The limited environmental interpretability of NVCS physiognomic classes at Zion led me to seek a more natural higher classification based on coalition group affinities. Using the affinity table presented in Appendix 2.4, I carried out a hierarchical cluster analysis that grouped associations in a more environmentally interpretable way. This analysis revealed clusters of associations representing certain physiognomic combinations in certain environmental settings (Fig. 2.15). The fourteen clusters highlighted in Figure 2.15 were named for their environmental (i.e. coalition group) affinities and physiognomic composition. These clusters are nested in three major groups with primary affinities for (1) Lowland Flats, Arid Lowlands, and Rocky Slopes; (2) Mesa Top, Upland Sands and Slickrock; and (3) Streambank, Crevice Canyon and High Plateau (Fig. 2.15). Some associations did not fit neatly within this classification scheme, e.g. Green Rabbitbrush/Kentucky Bluegrass Semi-natural Shrubland was a outlier to the Lowland Flats/Arid Lowlands/Rocky Slopes clusters; Needle-and-Thread Great Basin Herbaceous Vegetation was an outlier to the Crevice Canyon/High Plateau clusters, and Rocky Mountain Juniper--Gambel Oak Woodland was anomalously positioned next to the Streambank clusters (Fig. 2.15). Some of these outliers may be associations sampled by the USGS-NPS survey that were missed or poorly captured by the BYU-RMRS survey.

A reclassification exercise was also carried out for the USGS-NPS vegetation map units. Various higher classification schemes accompanied this map, including the

eight-class scheme shown in Table 2.5. These classes are more environmentally-descriptive than the NVCS formation classes because they distinguish between higher and lower elevation forests and shrublands, riparian and non-riparian zones, and slickrock as opposed to other bare soil/stone formations. Nevertheless, an even more descriptive classification (Fig. 2.16) was achieved through hierarchical clustering using affinities between vegetation map units and coalition group affinity maps (Appendix 2.5). The map unit clusters (Fig. 2.16) resembled association clusters (Fig. 2.15) in a general way, but map units were more strongly clustered by elevation and spatial proximity, as might be expected because of the spatial basis of the analysis. A cluster with High Plateau affinities (Higher Elevation Forest/Shrub/Herb) was linked to clusters with Mesa Top affinities (Higher Elevation Sparse, Higher Elevation Woodland/Shrub/Herb) rather than the cluster with Streambank affinities (Lower Elevation Riparian). Map units with Slickrock and Upland Sands affinities formed a tight cluster closely linked to a Crevice Canyon unit (the Douglas Fir Forest Alliance). The Lower Elevation Talus/Slope and Sparse/Shrub/Herb clusters parallel the Rocky Slopes and Arid Lowlands/Lowland Flats groups respectively. Lowland Flats affinities are also high for the Lower Elevation Disturbed cluster that contains a single map unit, Cheatgrass Annual Disturbed Grassland. I also assigned the name ‘Lower Elevation Sand’ to an anomalous cluster mixing riparian and upland sand substrates. Another anomaly involves Fremont Cottonwood-Velvet Ash Woodland, which was positioned near the Lower Elevation Sparse/Shrub/Herb cluster rather than the Riparian cluster, possibly because of inaccuracies in one or both of the map types being compared.

DISCUSSION

This paper has described how plant species groups were identified at Zion National Park using a clustering technique (coalition clustering) that quantifies the degree of affinity of each species to each group and allows species to belong to multiple groups. I asserted that these species groups (coalitions) are essentially faithful species according to an individualistic definition of community fidelity. Individualistic equivalents of constant species, defined as species that tend to co-occur with species groups, were also presented (affiliate species), and the relative dominance of these species was quantified. Because coalition species groups could be readily interpreted as species with similar environmental affinities at Zion, providing insights beyond previous vegetation studies, I conclude that the individualistic species group approach was an effective alternative to vegetation classification for characterizing community and environmental variation.

My discussion centers around the two main features of the individualistic species group approach that set it apart from previous plant community characterization studies at Zion. The individualistic species group approach (1) models spatial community variation continuously rather than discretely, and (2) gives species equal weight rather than weighting dominant species more heavily. Taking this approach does not preclude the possibility that relatively discrete patterns may be present, or that dominant species may be important determinants of community composition, but it does allow these possibilities to be actually examined rather than merely assumed.

Community Continuity and Discreteness

Gleason's individualistic concept (Gleason 1926, 1939) contributed to the later development of the community continuum concept by workers such as Curtis (1959), Goodall (1963), McIntosh (1967) and Whittaker (1967). Taken to its logical extreme, the continuum concept postulates that species distributions bear no relationship to one another, resulting in communities lacking any kind of natural boundaries (Leibold and Mikkelsen 2002). However, as pointed out earlier in this paper, Gleason tempered his views of community continuity with an acknowledgement that relatively discrete community patterns can often be recognized and are worthy of description and study (Gleason 1926, 1939). I have also pointed out that Curtis (1959) described communities in discrete terms for practical, heuristic reasons. The reality of (relatively) discrete community patterns and the utility of discrete community descriptions provide rationale for the individualistic species group approach followed here.

The individualistic species group approach addressed two types of patterns—species distributions and site composition—from two perspectives—discrete and continuous. Beginning with a discrete species perspective, groups of species with similar distribution patterns were identified through cluster analysis. Continuous species patterns were then added through species affinity calculations, then translated to continuous site patterns though weighted averaging at the plot scale. Finally, discrete site patterns were introduced through regression tree modeling which grouped sites sharing similar affinity values in similar environments. Spatial discreteness was thus relegated to a side effect of the final modeling step, rather than a primary feature of the community model.

What do these results tell us about community discreteness in the Zion landscape?

One would expect that the sharp transitions visible in the topography and some of the substrates would be natural boundaries for plant distributions. Regression tree models of coalition group affinities support this supposition in several instances. Narrow canyons and riparian zones define rather discrete high-affinity zones for the Crevice Canyon (Fig. 2.7) and Streambank (Fig. 2.5) coalition groups, as does the Navajo Sandstone for Slickrock and Upland Sands (Fig. 2.8-2.9). The zone of steep Navajo Sandstone cliffs appears as a dividing line in several regression tree models; for example, the elevational splitting point identified in the first data split of the Arid Lowlands coalition group (1596 m) lies at approximately the base of the lower Navajo Sandstone cliffs in the main section of the park (Fig. 2.11), while the first split of the Mesa Top groups (1791 m) lies near the top of this same set of cliffs (Fig. 2.13-2.14). This pattern breaks down, however, in the Kolob (northwestern) section of Zion where the Navajo Sandstone cliffs occur at a higher elevation than these data splitting lines. It would appear that main body of cliffs forms a natural break in the landscape detectable through regression tree modeling, but that the real factor involved is elevation, not the cliffs in and of themselves.

Austin and Smith (1989) pointed out that discrete community patterns are likely to be most evident in landscapes where extensive areas of similar environment are separated by narrow environmental transition zones—as appears to be the case for Zion. They then made a distinction between community patterns due to such landscape properties and continuum patterns due to underlying environmental gradients. If species are arranged along uniformly-scaled gradients, they argued, co-occurrence patterns should dissolve into continuous compositional gradients (Austin and Smith 1989). This

model assumes, however, that all areas of the environmental space are equally likely to be occupied, and that species are fully independent in terms of their ecological attributes. Countering this assumption, one might expect some species to be spatially clustered even in the absence of sharp environmental transitions due to phenomena such as phylogenetic niche conservatism (Webb et al. 2002) or positive ecological interactions (Callaway 1997). These phenomena can be accommodated by the individualistic concept because they fall under the category of “similarity in the contributing causes” leading to “the recurrence of a similar juxtaposition [of plant species] over tracts of measurable extent” as described by Gleason (1926, p. 25). In these cases (niche conservatism and ecological interactions) the contributing causes are species properties, not landscape properties.

Gleason (1926) invoked migration (dispersal history) and environmental selection as the primary causes of species distributions. To the extent that these causes are unique for each species, continuous community patterns will be manifest; to the extent that they are similar, discrete patterns are possible. This paper has focused on environmental causes of species distributions at Zion, but dispersal history undoubtedly also plays a role. Idiosyncrasies of dispersal can result in species being absent from portions of their optimal environment or disproportionately frequent in suboptimal environments (Shmida and Wilson 1985, Dunning et al. 1992, Nekola and White 1999). If barriers or facilitators of dispersal affect multiple species in a similar way, common distribution patterns may arise yet show little relationship to environmental variables. Such multi-species dispersal effects could have contributed to observed species association patterns at Zion, but examining such effects lies beyond the scope of this study. For current purposes,

dispersal effects are considered noise that weakens the species-environment relationships of interest.

Coalition Clustering Effects

Uncommon species that do not occupy all sites of suitable habitat are one of the primary sources of dispersal-related noise in community datasets. The coalition clustering technique partially compensates for this type of noise by linking species having overlapping distributions without requiring that all species overlap in the same way. In many cases, species belonging to the same coalition group at Zion did not actually co-occur in any plots, but the negative pairwise associations of these species were overridden by their shared positive associations with other species belonging to the group. Species occurring in many plots appeared to provide some of the “glue” that held species occurring in fewer plots together in the same group.

The phi coefficient of association likely accentuated the tendency toward diffuse overlap within coalition groups. Because phi incorporates joint absence information, it is less sensitive to differences in species frequency than presence-only coefficients such as the Jaccard index (Jackson et al. 1989, Sanderson et al. 2006). Species clustering based on presence-only coefficients tends to group species based on their frequency unless additional standardization steps are followed (Austin and Belbin 1982, Jackson et al. 1989, McCune and Grace 2002). In contrast, coalition clustering using the phi coefficient yielded species groups containing mixtures of common and rarer species. Species within coalition groups can have variable niche breadths within a diffuse zone of shared niche space. The same can be true of species that are faithful to a community unit, such as the

modal species described by Harper et al. (2001), although in this case the niche space is discretely defined.

Because coalition clustering solutions vary depending on the threshold association value defined by the analyst, multiple values may need to be explored to find a solution at the desired scale. In the current application, the selected solution contained a moderate number of clusters with moderate numbers of species each, at a scale that allowed comparison with the vegetation classification of Harper et al. (2001). A more objective method for cluster selection could be implemented by searching across multiple threshold values to find an “optimal” solution in which association is maximized within clusters and minimized between clusters (cf. Sanderson et al. 2006). Similar optimal partitioning procedures have been developed for use with standard clustering algorithms (Aho et al. 2008). Although optimal partitioning has not been tested with coalition clustering, its likely effect would be to select a solution with minimal species overlap between groups. Since overlapping composition contributes to the individualistic character of species groups, this would be an undesirable effect for current purposes. Furthermore, the idea of a single optimal classification solution runs contrary to the pluralistic spirit of the individualistic concept. The coalition groups presented here are but one of many potentially useful and informative clustering solutions.

Another reason why optimal partitioning may have limited relevance for coalition clustering is because a given threshold value does not necessarily mean the same thing for different coalition clusters, depending on their representation in community samples. The two Mesa Top groups, for example, could be differentiated at a threshold value of phi=0.15 because of the relatively large number of plots representing the Carmel

Formation zone. If there had been fewer plots in this zone, a higher threshold value may have been required to distinguish these two groups. The Arid Lowlands group provides another example of how threshold value selection influences coalition clustering results. The emergence of Arid Lowlands as a single group was somewhat surprising given its range of vegetation types and mixture of Mojave Desert and Great Basin species. Exploration of alternative clustering solutions at different threshold values helped to clarify this pattern. Figure 2.17 illustrates an alternative set of two Arid Lowland groups obtained using coalition clustering at phi=0.2. One group containing *P. monophylla* and *J. osteosperma* is recognizable as having higher elevation/Great Basin affinities while the other has lower elevation/Mojave affinities (Fig. 2.17). Nevertheless, these two groups overlap to a considerable degree and the invasive grass *B. rubens* is the highest-affinity species in both of them. Other overlapping species have mostly inverted affinity rank order in the two groups, as illustrated by the lines connecting these species in Figure 2.17. This situation is reminiscent of the inverted affinities of species in the two Mesa Top groups (Table 2.2I-J). Both Arid Lowlands and Mesa Top can thus be characterized as loosely-associated species groups that can be subdivided into overlapping subgroups. Unlike Arid Lowlands subgroups, the Mesic and Xeric Mesa Top groups do not appear to represent species with different geographic affinities, and their ecological significance at Zion deserves further study.

Sampling Issues

One of the drawbacks of coalition clustering, and species clustering in general, is that association patterns are sensitive to the area sampled and sampling strategy. Discrete

community units need not be defined, but the decision of how to define and sample the study area remains. In the current application, the use of national park boundaries to delimit the study area makes sense from a political or management perspective but not from an ecological perspective. Zion has artificial boundaries that truncate environmental gradients; the elevation gradient, for example, continues both below Zion in the Virgin River drainage and above on the Markagunt Plateau. Species whose environmental affinities lie primarily outside of such truncated gradients are incompletely characterized in analyses restricted to Zion.

However, given the fact that the National Park was chosen as the study area, the systematic sampling scheme followed by the BYU-RMRS survey was a reasonably unbiased approach for capturing environmental variation within the park. Aside from the unlikely possibility that the ca. 1.6 km distance between plots coincides with periodic environmental variation at Zion, the more-or-less evenly spaced plot arrangement of the BYU-RMRS survey (Fig. 2.2) yielded an area-proportional sample of Zion's environments. Environmental conditions that are more frequent in the landscape were captured with greater frequency by this approach, in contrast to the gradsect sampling strategy followed by the USGS-NPS survey which sought to capture the range of environmental variation without regard for frequency.

The effects of these different sampling strategies are illustrated in histograms showing the proportional correspondence between plots and pixels for the environmental variables used in this study, which corresponded more closely for the BYU-RMRS survey than the USGS-NPS survey (Fig. 2.18-2.19). Even so, the BYU-RMRS dataset had an over-representation of low elevations, low slopes, moderate exposures (Fig. 2.18)

and alluvial substrates (Fig. 2.19) that can be attributed at least in part to the non-systematic riparian and old field plots included in the survey. On the other hand, excessively steep or bare plot locations were avoided or adjusted by BYU-RMRS survey crews leading to an under-representation of high slopes and Navajo Sandstone substrates. However, these same patterns of environmental over- and under-representation were present to an even greater degree in the USGS-NPS dataset (Fig. 2.18-2.19). Consequently the BYU-RMRS dataset has a larger, comparatively more representative sample of common environments at Zion (such as the Navajo Sandstone), while the USGS-NPS dataset has better representation of rarer, more unique environments and communities (such as wetlands). In terms of ecological representation, these two datasets can be viewed as complementary.

The merits of systematic or random versus subjective community sampling have been widely debated. In a recent forum (Herben and Chytrý 2007), Lájer (2007) and Chiarucci (2007) stressed the importance of random, statistically-independent samples for rigorous statistical analysis of vegetation patterns, while other authors noted that this level of rigor is not crucial for many vegetation science objectives (Botta-Dukát et al. 2007, Diekmann et al. 2007, Lepš and Šmilauer 2007, Økland 2007, Roleček et al. 2007). A truly random plot arrangement is not only unlikely to capture community variation of interest (Diekmann et al. 2007, Lepš and Šmilauer 2007, Roleček et al. 2007), but is not truly statistically-independent because of spatial autocorrelation (Økland 2007, Ricotta 2007, Wilson 2007). Hence some form of sample stratification, whether spatial, environmental or vegetation-based, is usually required for meaningful community analysis (Økland 2007, Wilson 2007). Adding to this discussion, I note that evenly-

spaced systematic plot sampling is a form of spatial stratification that minimizes the effects of spatial autocorrelation as well as the bias of environmental/vegetation stratification schemes. Such a sampling strategy fits well with an individualistic approach that does not require *a priori* decisions concerning which environmental variables are of greatest importance in determining species distributions. Systematic sampling also captures environmental conditions that might otherwise be excluded because of their “transitional” nature, such as the vegetation transitions avoided by the USGS-NPS survey (Cogan et al. 2004).

Differences in sampling strategy between the BYU-RMRS and USGS-NPS surveys pose certain limitations when comparing the patterns they reveal. Species association patterns evident in the BYU-RMRS dataset would not be expected to be quantitatively repeated in the USGS-NPS dataset because of its different sampling distribution, not to mention differences in plot size, taxonomic resolution and the ca. 20-year interval between the two surveys. These considerations dampened the otherwise attractive possibility of using the USGS-NPS dataset to assess the accuracy of the predictive spatial models built using the BYU-RMRS dataset. Such an accuracy assessment was attempted by overlaying validation (USGS-NPS) plots on predictive affinity maps for each coalition group, then comparing map pixel values with affinities based on validation plot species composition. This exercise revealed widespread quantitative differences between validation plot- and predictive map-based affinity estimates suggesting low predictive accuracy of the regression tree models. However, an alternative interpretation is also possible: low predictive accuracy may merely indicate the failure of a subjective dataset (USGS-NPS) to predict affinity patterns in the same

way as a systematic dataset (BYU-RMRS). Edwards et al. (2006) present a study relevant to this issue; they found that classification tree models based on a ‘purposive’ sample (subjective, targeting certain species or landscape features) were less accurate in predicting distributions of four lichen species than models based on a random/systematic sample. This raises questions over the appropriateness of trying to assess the accuracy of a systematically-derived model using a subjective validation dataset; such an assessment may best be limited to a qualitative analysis. For the tree models presented here, qualitative model accuracy assessments based on ordinal relationships at each tree node revealed high correspondence between the prediction and validation datasets (i.e., predicted separation of higher vs. lower mean affinity values was replicated at 44 out of 47 regression tree nodes).

Dominant Species, Floristic Patterns and Environmental Interpretations

In addition to being associated with the continuum concept, the individualistic concept has been associated with the idea that interspecific interactions play a minimal role in determining plant community composition (Callaway 1997, Nicolson and McIntosh 2002). Gleason did not actually hold such an extreme view, as evidenced by his references to the controlling influence of dominant plant species over subordinate species (Gleason 1939, p. 104-105; cf. Nicolson and McIntosh 2002). However, Gleason challenged the notion that structural dominants are the primary factor controlling community composition at all locations, or that all species are equally affected by their influence:

Under the individualistic concept, the fundamental idea is neither extent, unit character, permanence, nor definiteness of structure. It is rather the visible expression, through the juxtaposition of individuals, of the same or different species and either *with* or *without mutual influence*, of the result of causes in continuous operation. (Gleason 1926, p. 25, emphasis added)

In contrast, the controlling influence of dominant species was a central theme of Clements' (1916, 1936) organismal community concept:

Hence, the essential unity of a climax is to be sought in its dominant species... Their reactions and coactions are the most controlling both in kind and amount, and thus they determine the conditions under which all the remaining species are associated with them. (Clements 1936, p. 255)

Given that Clements was here referring specifically to climax vegetation, and was not ignorant of other factors affecting community composition (cf. Eliot 2007), he was not suggesting that dominant species are the primary determinants of community composition under all circumstances. My purpose in quoting Clements is to portray a perspective that provides rationale for the widespread assumption that dominant species should be granted particular emphasis in plant community characterization. I use Gleason's individualistic concept as a counter perspective justifying the purely floristic emphasis of the species group approach, in which dominance is relegated to a secondary descriptor status.

The distinction between community characterization approaches that emphasize dominant species and those that do not is illustrated by Harper et al. (2001) in comparison with the current individualistic species group analysis. Harper et al. (2001) began with a set of vegetation types defined more or less by dominant species and vegetation structure. Prevalent species were then identified for each vegetation type and modal species were derived from the prevalents. The sequence of community characterization thus progressed from dominant to constant (prevalent) to faithful (modal) species. Constant and faithful species in this instance became indicators of the vegetation types, constrained by the dominant species emphasis of the classification. In contrast, the current analysis

began by identifying coalition (faithful) species, then affiliate (constant) species, and finally dominant affiliates. This reversal changed the role of faithful species from constrained indicators to unconstrained community descriptors, and changed constants and dominants to secondary descriptors. Dominant species became, in essence, potential explanatory variables for floristic community patterns. The current approach thus bears a conceptual resemblance to unconstrained ordination or indirect gradient analysis (Whittaker 1967, Kent and Coker 1992, McCune and Grace 2002), while the approach followed by Harper et al. (2001) resembles constrained ordination in the sense that the ‘environmental’ variable of species dominance was imposed *a priori*.

The classification hierarchy of the NVCS can also be viewed as a constrained model in which lower floristic classes are forced into higher physiognomic ones. The incongruence of this classification strategy has been recognized and a revised system that merges floristics and physiognomy more seamlessly has recently been proposed (Faber-Langendoen et al. 2009). A related effort has established a set of “ecological systems” and “ecological divisions” comprised of NVCS associations occupying similar environments (Comer et al. 2003, Comer and Schulz 2007). The seven ecological divisions occurring at Zion (Forest and Woodland, Herbaceous Wetland, Savanna and Shrub-steppe, Sparsely Vegetated, Upland Grassland and Herbaceous, Upland Shrubland, Woody Wetlands and Riparian) (NatureServe 2007) resemble to a certain degree the coalition-group-affinity-based ecological/physiognomic groupings presented in Fig. 2.15-2.16, but the latter provide greater specificity in terms of environments found within the park. While more ecologically-informative than the pre-existing NVCS physiognomic

hierarchy, both ecological systems and the new hierarchy are built primarily around structural dominant based descriptors.

Even at the floristic levels of the NVCS (association and alliance), vegetation classification and mapping at Zion emphasized dominant species and followed a constrained analysis approach. Cogan et al. (2004) describe a series of multivariate analyses that guided the classification process, in which species appear to have been weighted by their abundance (cover). Although all species contributed information, dominant species were awarded particular attention and served as the primary criteria for differentiating associations (Cogan et al. 2004, Appendix E). Dominant species likewise played an important role in the mapping process as the primary contributor to aerial photo vegetation signatures, although other visible features related to substrate and topography also contributed (Cogan et al. 2004). Pure floristic patterns, though embedded in the classification and mapping system, were constrained by the *a priori* emphasis on dominant species and abiotic environmental variables. In contrast, coalition species groups of the current analysis were not weighted by abundance and were related to environmental variables *a posteriori*, in accordance with the philosophy of letting the species ‘interpret’ the environment from their own perspective (Beals 1984, McCune and Grace 2002). Environmental affinity models for coalition groups demonstrated that floristic variation at Zion can be explained to a considerable degree by abiotic variables without invoking dominant species effects. This does not preclude the possibility that dominant species effects are also influential (and may co-vary with abiotic variables), but it does suggest that dominant species are not the primary determinants of broad-scale floristic patterns at Zion, at least of the kind captured by the current analysis. Further

work is needed to more fully characterize the relative importance of abiotic and biotic factors for different species and scales (cf. Chapter 3).

It should be noted that the current analysis de-emphasized dominant species because coalition clustering is an R-mode (species-based) approach and not simply because species presence-absence was used instead of abundance. Presence-absence association coefficients (e.g. phi) can be substituted with correlation coefficients (e.g. Pearson's r) that incorporate abundance information in R-mode (Ludwig and Reynolds 1998), but this does not lead to a dominant species emphasis because abundances become relativized in the process. Abundance information is not necessarily more useful than presence-absence, depending on the scale and objective of the analysis. Presence-absence information proved effective for characterizing species groups at Zion, where environmental/floristic spatial turnover is high, but might not have sufficed in a smaller or more homogeneous area.

Using presence-absence has the effect of broadening the temporal scale of the community pattern detected; in other words, presence-absence patterns are more likely to reflect long-term environmental conditions (e.g., prevailing climate and substrate) than short-term environmental fluctuations or disturbances that affect abundance (Allen and Starr 1982). Fire disturbance appears to fall into the latter category at the scale of the current analysis, inasmuch as two recently-burned (earlier in the same season) plots sampled in 1988 were compositionally, though not structurally, similar to surrounding plots with Mesa Top affinities. Most plots of the 1987-89 survey were recorded as late seral in relation to fire; hence it is difficult to extrapolate these results to early post-fire conditions in general (on the other hand, the 1987-89 survey provides a valuable baseline

for evaluating the effects of recent extensive wildfires at Zion). Fire effects are likely to vary by species and location, and although individual fires may qualify as short-term disturbances, a recurring fire regime can take on the form of a long-term environmental condition. Floyd et al. (2000), upon examining fire history of vegetation types in Mesa Verde National Park, Colorado, concluded that more frequent fire over a 150-year time scale partially explains the distribution of mountain brush vegetation (which tends to resprout after fire) in relation to pinyon-juniper vegetation where fire return intervals have been longer. Fire history may have played a similar role in structuring these same vegetation types at Zion, although its role at the floristic level is less certain.

The question of community stability over time is an important theme related to the individualistic concept, and thus deserves comment to round out this discussion. Given that the plant community patterns described in this paper are but snapshots of conditions at a fleeting period in time, to what extent can they be viewed as permanent features of the Zion landscape? Although a precise answer to this question lies beyond the scope of this study, it is clear that communities are dynamic and change of some form or another is to be expected. Plant species distributions in the Colorado Plateau region are known to have shifted during the Pleistocene (Cole 1990, Anderson et al. 1999, 2000, Coats et al. 2008) and are expected to shift in response to projected climate changes during coming decades (Krause 2009).

The general prediction that a warming, drying trend on the Colorado Plateau (Schwinning et al. 2008) should cause species distributions to shift upward in elevation is complicated by individualistic species responses and landscape configuration at Zion (cf. le Roux and McGeoch 2008). Because of the more or less horizontal arrangement of

major rock strata at Zion, an elevational climatic shift would decouple current climate-substrate combinations. Three classes of species responses under this scenario can be envisioned: (1) species sensitive to climate, but less sensitive to substrate will shift upward in elevation (dispersal permitting), (2) species sensitive to substrate, but less sensitive to climate substrate will maintain current distributions, and (3) species sensitive to both climate and substrate will disappear from the park to the degree that their required climate-substrate combination becomes unavailable (a fourth category for species responding to factors other than climate or substrate could also be included). Determining which species may fall into these different categories is difficult to judge from correlative environmental modeling, but the current study offers some clues. Species with high affinities for coalition groups that were strongly correlated with elevation (e.g., Arid Lowlands, High Plateau and the Mesa Top groups) have a greater chance of belonging to category 1, while species with high affinities for groups with strong substrate correlations (e.g. Slickrock, Upland Sands) could belong to categories 2 or 3. Since substrate specificity is a critical differentiating factor in this scenario, making species either more or less susceptible to climate change, such species may deserve particular attention and further study.

REFERENCES

- Abella, S. R., and V. B. Shelburne. 2004. Ecological species groups of South Carolina's Jocassee Gorges, Southern Appalachian Mountains. *Journal of the Torrey Botanical Society* 131:220-231.
- Aho, K., D. W. Roberts, and T. Weaver. 2008. Using geometric and non-geometric internal evaluators to compare eight vegetation classification methods. *Journal of Vegetation Science* 19:549-U513.
- Allen, T. F. H., and T. B. Starr. 1982. *Hierarchy: Perspectives for Ecological Complexity*. University of Chicago Press, Chicago, IL.
- Anderson, R. S., J. L. Betancourt, J. I. Mead, R. H. Hevly, and D. P. Adam. 2000. Middle- and Late-Wisconsin paleobotanic and paleoclimatic records from the southern Colorado Plateau, USA. *Palaeogeography Palaeoclimatology Palaeoecology* 155:31-57.
- Anderson, R. S., J. Hasbargen, P. A. Koehler, and E. J. Feiler. 1999. Late Wisconsin and Holocene subalpine forests of the Markagunt Plateau of Utah, southwestern Colorado Plateau, USA. *Arctic Antarctic and Alpine Research* 31:366-378.
- Austin, M. P., and L. Belbin. 1982. A new approach to the species classification problem in floristic analysis. *Australian Journal of Ecology* 7:75-90.
- Austin, M. P., and T. M. Smith. 1989. A new model for the continuum concept. *Vegetatio* 83:35-47.
- Barkman, J. J. 1989. Fidelity and character-species, a critical evaluation. *Vegetatio* 85:105-116.
- Beals, E. W. 1984. Bray-Curtis ordination - an effective strategy for analysis of multivariate ecological data. *Advances in Ecological Research* 14:1-55.
- Beyer, H. L. 2004. Hawth's Analysis Tools for ArcGIS.
<http://www.spatialecology.com/htools>.
- Biek, R. F., G. C. Willis, M. D. Hylland, H. H. Doelling, D. A. Sprinkel, T. C. Chidsey, Jr., and P. B. Anderson. 2003. Geology of Zion National Park, Utah. Pages 107-137 in Anonymous, editor. *Geology of Utah's Parks and Monuments*. Utah Geological Association and Bryce Canyon Natural History Association.
- BLM (USDI Bureau of Land Management). 2006. Geographic Coordinate Data Base.
<http://www.blm.gov/wo/st/en/prog/more/gcdb.html>.

- Botta-Dukát, Z., E. Kovacs-Lang, T. Redei, M. Kertesz, and J. Garadnai. 2007. Statistical and biological consequences of preferential sampling in phytosociology: Theoretical considerations and a case study. *Folia Geobotanica* 42:141-152.
- Braun-Blanquet, J. 1965. *Plant Sociology: The Study of Plant Communities*. Hafner Pub. Co., New York.
- Breiman, L. 1993. *Classification and Regression Trees*. Chapman & Hall, New York.
- Calenge, C. 2006. The package adehabitat for the R software: A tool for the analysis of space and habitat use by animals. *Ecological Modelling* 197:516-519.
- Callaway, R. M. 1997. Positive interactions in plant communities and the individualistic-continuum concept. *Oecologia* 112:143-149.
- Chiarucci, A. 2007. To sample or not to sample? That is the question... For the vegetation scientist. *Folia Geobotanica* 42:209-216.
- Clements, F. E. 1916. *Plant Succession: An Analysis of the Development of Vegetation*. Carnegie Institution of Washington, Washington.
- Clements, F. E. 1936. Nature and structure of the climax. *Journal of Ecology* 24:252-284.
- Coats, L. L., K. L. Cole, and J. I. Mead. 2008. 50,000 years of vegetation and climate history on the Colorado Plateau, Utah and Arizona, USA. *Quaternary Research* 70:322-338.
- Cogan, D., M. Reid, K. Schulz, and M. Pucherelli. 2004. Zion National Park, Utah: 1999-2003 Vegetation Mapping Program, Final Report--March 31, 2004. Report Technical Memorandum 8260-03-01, Remote Sensing and GIS Group, Technical Service Center, Bureau of Reclamation, Denver, CO.
<http://biology.usgs.gov/npsveg/zion/zionrpt.pdf>.
- Cole, K. L. 1990. Reconstruction of past desert vegetation along the Colorado River USA using packrat middens. *Palaeogeography Palaeoclimatology Palaeoecology* 76:349-366.
- Comer, P., D. Faber-Langendoen, R. Evans, S. Gawler, C. Josse, G. Kittel, S. Menard, M. Pyne, M. Reid, K. Schulz, K. Snow, and J. Teague. 2003. Ecological Systems of The United States: A Working Classification of U.S. Terrestrial Systems. NatureServe, Arlington, VA.
<http://www.natureserve.org/publications/usEcologicalsystems.jsp>.
- Comer, P. J., and K. A. Schulz. 2007. Standardized ecological classification for mesoscale mapping in the southwestern United States. *Rangeland Ecology and Management* 60:324-335.

- Crawley, M. J. 2007. *The R Book*. John Wiley & Sons Ltd., West Sussex, England.
- Curtis, J. T. 1959. *The Vegetation of Wisconsin: An Ordination of Plant Communities*. University of Wisconsin Press, Madison, WI.
- Diekmann, M., A. Kuhne, and M. Isermann. 2007. Random vs non-random sampling: Effects on patterns of species abundance, species richness and vegetation-environment relationships. *Folia Geobotanica* 42:179-190.
- Doing, H. 1969. Sociological species groups. *Acta Botanica Neerlandica* 18:398-399.
- Dunning, J. B., B. J. Danielson, and H. R. Pulliam. 1992. Ecological processes that affect populations in complex landscapes. *Oikos* 65:169-175.
- Edwards, T. C., D. R. Cutler, N. E. Zimmermann, L. Geiser, and G. G. Moisen. 2006. Effects of sample survey design on the accuracy of classification tree models in species distribution models. *Ecological Modelling* 199:132-141.
- Eliot, C. 2007. Method and metaphysics in Clements's and Gleason's ecological explanations. *Studies in History and Philosophy of Biological and Biomedical Sciences* 38:85-109.
- ESA (Ecological Society of America). 2010. VegBank vegetation plot database. <http://vegbank.org/>.
- ESRI (Environmental Systems Research Institute). 2003, 2004. ArcGIS version 8-9. <http://www.esri.com/software/arcgis/index.html>.
- Faber-Langendoen, D., N. Aaseng, K. Hop, M. Lew-Smith, and J. Drake. 2007. Vegetation classification, mapping, and monitoring at Voyageurs National Park, Minnesota: An application of the U.S. National Vegetation Classification. *Applied Vegetation Science* 10:361-U370.
- Faber-Langendoen, D., D. L. Tart, and R. H. Crawford. 2009. Contours of the revised U.S. National Vegetation Classification Standard. *Bulletin of the Ecological Society of America* 90:87-93.
- Ferrier, S., M. Drielsma, G. Manion, and G. Watson. 2002. Extended statistical approaches to modelling spatial pattern in biodiversity in Northeast New South Wales. II. Community-level modelling. *Biodiversity and Conservation* 11:2309-2338.
- Ferrier, S., and A. Guisan. 2006. Spatial modelling of biodiversity at the community level. *Journal of Applied Ecology* 43:393-404.

Fertig, W., and J. Alexander. 2009. Annotated Checklist of Vascular Flora: Zion National Park. Natural Resource Technical Report NPS/NCPN/NRTR-2009/157, USDI National Park Service, Natural Resource Program Center, Fort Collins, CO.

FGDC (Federal Geographic Data Committee). 2008. National Vegetation Classification Standard, Version 2. <http://www.fgdc.gov/standards/projects/FGDC-standards-projects/vegetation/>.

Floyd, M. L., W. H. Romme, and D. D. Hanna. 2000. Fire history and vegetation pattern in Mesa Verde National Park, Colorado, USA. *Ecological Applications* 10:1666-1680.

Gillison, A. N., and K. R. W. Brewer. 1985. The use of gradient directed transects or gradsects in natural-resource surveys. *Journal of Environmental Management* 20:103-127.

Gleason, H. A. 1926. The individualistic concept of the plant association. *Bulletin of the Torrey Botanical Club* 53:7-26.

Gleason, H. A. 1939. The individualistic concept of the plant association. *American Midland Naturalist* 21:92-110.

Goodall, D. W. 1963. The continuum and the individualistic association. *Vegetatio* 11:297-316.

Grossman, D. H., D. Faber-Langendoen, A. S. Weakley, M. Anderson, P. Bourgeron, R. Crawford, K. Goodin, S. Landaal, K. Metzler, K. Patterson, M. Pyne, M. Reid, and L. Sneddon. 1998. International Classification of Ecological Communities: Terrestrial Vegetation of the United States, Volume 1: The National Vegetation Classification System: Development, Status, and Applications. The Nature Conservancy, Arlington, VA. <http://www.natureserve.org/library/vol1.pdf>.

Harper, K. T. 1993. Zion National Park Vegetation: Summary Report. Report, Zion National Park, Springdale, UT.

Harper, K. T., S. C. Sanderson, and E. D. McArthur. 1992. Riparian ecology at Zion National Park. Pages 32-42 in Proceedings—Symposium on Ecology and Management of Riparian Shrub Communities. U.S. Department of Agriculture, Forest Service, Intermountain Research Station, Ogden, UT.

Harper, K. T., S. C. Sanderson, and E. D. McArthur. 2001. Quantifying plant diversity at Zion National Park. Pages 318-324 in Shrubland Ecosystem Genetics and Biodiversity: Proceedings. U.S. Department of Agriculture, Forest Service, Rocky Mountain Research Station, Ogden, UT.

- Harper, K. T., S. C. Sanderson, and E. D. McArthur. 2003. Pinyon-juniper woodlands in Zion National Park, Utah. *Western North American Naturalist* 63:189-202.
- Herben, T., and M. Chytrý. 2007. Analysis of non-randomly sampled data sets in vegetation ecology. *Folia Geobotanica* 42:113-113.
- Insightful Corp. 2005. S-PLUS. <http://www.insightful.com/products/splus/default.asp>.
- Jackson, D. A., K. M. Somers, and H. H. Harvey. 1989. Similarity coefficients - measures of co-occurrence and association or simply measures of occurrence? *American Naturalist* 133:436-453.
- Jeffries, D. L., and J. M. Klopatek. 1987. Effects of grazing on the vegetation of the blackbrush association. *Journal of Range Management* 40:390-392.
- Jenness, J. 2006. Topographic Position Index (TPI).
http://www.jennessent.com/downloads/TPI_Documentation_online.pdf.
- Jennings, M. D., D. Faber-Langendoen, O. L. Loucks, R. K. Peet, and D. Roberts. 2009. Standards for associations and alliances of the US National Vegetation Classification. *Ecological Monographs* 79:173-199.
- Kartesz, J. T. 1999. A Synonymized Checklist and Atlas with Biological Attributes for the Vascular Flora of the United States, Canada, and Greenland, First Edition. In J. T. Kartesz and C. A. Meacham, editors. *Synthesis of the North American Flora*, Version 1.0. North Carolina Botanical Garden, Chapel Hill, NC.
- Kent, M., and P. Coker. 1992. *Vegetation Description and Analysis: A Practical Approach*. CRC Press, Boca Raton.
- Krause, C.M. 2009. Effects of climate change on geographic distributions of endemic plants of the Colorado Plateau. Abstracts of the 94th Ecological Society of America Annual Meeting, Albuquerque, NM.
<http://eco.confex.com/eco/2009/techprogram/P17331.HTM>.
- Lájer, K. 2007. Statistical tests as inappropriate tools for data analysis performed on non-random samples of plant communities. *Folia Geobotanica* 42:115-122.
- le Roux, P. C., and M. A. McGeoch. 2008. Rapid range expansion and community reorganization in response to warming. *Global Change Biology* 14:2950-2962.
- Leibold, M. A., and G. M. Mikkelsen. 2002. Coherence, species turnover, and boundary clumping: Elements of meta-community structure. *Oikos* 97:237-250.
- Lepš, J., and P. Šmilauer. 2007. Subjectively sampled vegetation data: Don't throw out the baby with the bath water. *Folia Geobotanica* 42:169-178.

- Ludwig, J. A., and J. F. Reynolds. 1988. Statistical Ecology: A Primer on Methods and Computing. John Wiley & Sons, New York.
- Madany, M. H., and N. E. West. 1983. Livestock grazing fire regime interactions within montane forests of Zion National Park, Utah. *Ecology* 64:661-667.
- Madany, M. H., and N. E. West. 1984. Vegetation of two relict mesas in Zion National Park. *Journal of Range Management* 37:456-461.
- Malanson, G. P., and J. Kay. 1980. Flood frequency and the assemblage of dispersal types in hanging gardens of the narrows, Zion National Park, Utah USA. *Great Basin Naturalist* 40:365-371.
- McCune, B., and J. B. Grace. 2002. Analysis of Ecological Communities. MjM Software Design, Gleneden Beach, OR.
- McCune, B., and D. Keon. 2002. Equations for potential annual direct incident radiation and heat load. *Journal of Vegetation Science* 13:603-606.
- McIntosh, R. P. 1967. Continuum concept of vegetation. *Botanical Review* 33:130-&.
- McIntosh, R. P. 1975. H. A. Gleason – “individualistic ecologist” 1882-1975: His contributions to ecological theory. *Bulletin of the Torrey Botanical Club* 102:253-273.
- McLaughlin, S. P. 1986. Floristic analysis of the southwestern United States. *Great Basin Naturalist* 46:46-65.
- McLaughlin, S. P. 1989. Natural floristic areas of the western United States. *Journal of Biogeography* 16:239-248.
- Meyer, S. E. 1978. Some factors governing plant distributions in the Mojave-Intermountain transition zone. *Great Basin Naturalist Memoirs* 2:197-207.
- Mortensen, V. L. 1977. Soil Survey of Washington County Area, Utah. Dept. of Agriculture, Soil Conservation Service, Washington DC.
- Mueller-Dombois, D., and H. Ellenberg. 1974. Aims and Methods of Vegetation Ecology. Wiley, New York, NY.
- NatureServe. 2007. International Ecological Classification Standard: Terrestrial Ecological Classifications. NatureServe Central Databases. Arlington, VA. Data current as of 27 August 2007.
- Nekola, J. C., and P. S. White. 1999. The distance decay of similarity in biogeography and ecology. *Journal of Biogeography* 26:867-878.

- Nicolson, M., and R. P. McIntosh. 2002. H.A. Gleason and the individualistic hypothesis revisited. *Bulletin of the Ecological Society of America* 83:133-142.
- NPS (U.S. National Park Service). 2009. A Century of Sanctuary 1909–2009. Zion National Park Official Centennial Newspaper.
http://www.nps.gov/zion/parknews/upload/Zion%20Centennial%20Newspaper%201909-2009_7-11-09.pdf.
- NPS (U.S. National Park Service). 2010. NPS Data Store.
<http://science.nature.nps.gov/nrdata/index.cfm>.
- O'Meara, S. 2006. Geologic Map of Zion National Park and Vicinity, Utah.
<http://science.nature.nps.gov/nrdata/datastore.cfm?ID=41895>.
- Økland, R. H. 2007. Wise use of statistical tools in ecological field studies. *Folia Geobotanica* 42:123-140.
- R Development Core Team. 2008, 2010. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria.
<http://www.r-project.org/>.
- Ricotta, C. 2007. Random sampling does not exclude spatial dependence: The importance of neutral models for ecological hypothesis testing. *Folia Geobotanica* 42:153-160.
- Ripple, W. J., and R. L. Beschta. 2006. Linking a cougar decline, trophic cascade, and catastrophic regime shift in Zion National Park. *Biological Conservation* 133:397-408.
- Roleček, J., M. Chytrý, M. Hájek, S. Lvončík, and L. Tichý. 2007. Sampling design in large-scale vegetation studies: Do not sacrifice ecological thinking to statistical purism! *Folia Geobotanica* 42:199-208.
- Sanderson, S. C., J. E. Ott, E. D. McArthur, and K. T. Harper. 2006. RCLUS, a new program for clustering associated species: A demonstration using a Mojave Desert plant community dataset. *Western North American Naturalist* 66:285-297.
- Schwinning, S., J. Belnap, D. R. Bowling, and J. R. Ehleringer. 2008. Sensitivity of the Colorado Plateau to change: Climate, ecosystems, and society. *Ecology and Society* 13:20.
- Sharow, D. 2007a. Formation properties and limitations for geologic formations found in and near Zion National Park and Pipe Spring National Monument. Report, Zion National Park, Springdale, UT.

- Sharow, D. 2007b. Zion climate, hydrology and water quality. Report, Zion National Park, Springdale, UT.
- Shmida, A., and M. V. Wilson. 1985. Biological determinants of species diversity. *Journal of Biogeography* 12:1-20.
- Tichý, L., and M. Chytrý. 2006. Statistical determination of diagnostic species for site groups of unequal size. *Journal of Vegetation Science* 17:809-818.
- UCC (Utah Climate Center). 2010. Utah Climate Center Database. <http://climate.usurf.usu.edu/>.
- USGS (U.S. Geologic Survey). 2004. USGS Vegetation Characterization: Zion National Park. <http://biology.usgs.gov/npsveg/zion/index.html>.
- USGS (U.S. Geologic Survey). 2010. USGS Vegetation Characterization Program. <http://biology.usgs.gov/npsveg/about.html>.
- Webb, C. O., D. D. Ackerly, M. A. McPeek, and M. J. Donoghue. 2002. Phylogenies and community ecology. *Annual Review of Ecology and Systematics* 33:475-505.
- Weiss, A. D. 2001. Topographic position and landforms analysis. The Nature Conservancy and Jenness Enterprises, Seattle, Washington. http://www.jennessent.com/arcview/TPI_Weiss_poster.htm.
- Welsh, S. L. 1989. Hanging Gardens of Zion National Park: Final Report. Report, Endangered Plant Studies, Inc, Orem, UT.
- Welsh, S. L., N. D. Atwood, S. Goodrich, and L. C. Higgins. 1987. A Utah Flora, 1st Edition. Brigham Young University, Provo, UT.
- Welsh, S. L., N. D. Atwood, S. Goodrich, and L. C. Higgins. 2003. A Utah Flora, 3rd Edition. Brigham Young University, Provo, UT.
- Whittaker, R. H. 1967. Gradient analysis of vegetation. *Biological Reviews of the Cambridge Philosophical Society* 42:207-&.
- Wilson, J. B. 2007. Priorities in statistics, the sensitive feet of elephants, and don't transform data. *Folia Geobotanica* 42:161-167.
- Woodbury, A. M. 1933. Biotic relationships of Zion Canyon, Utah with special reference to succession. *Ecological Monographs* 3:151-245.

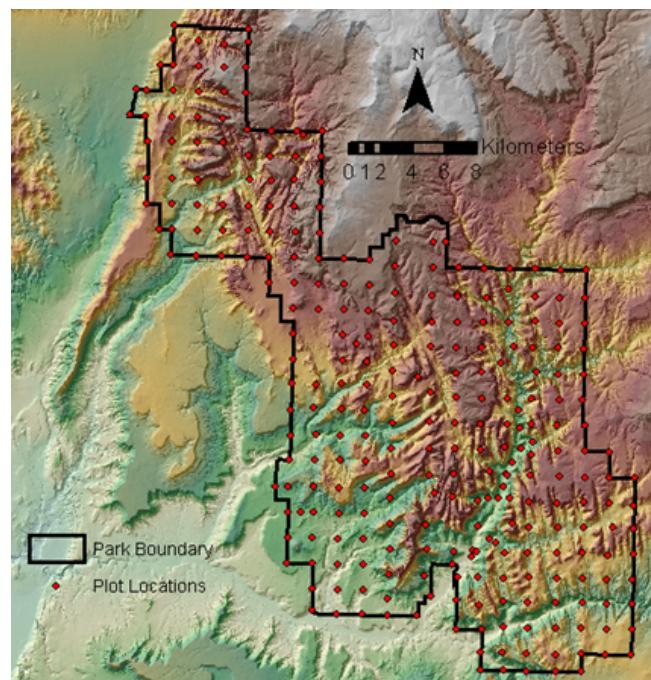


Figure 2.1. Zion National Park, Utah in relation to topography, showing plot locations of the 1987-89 vegetation survey (Harper 1993)

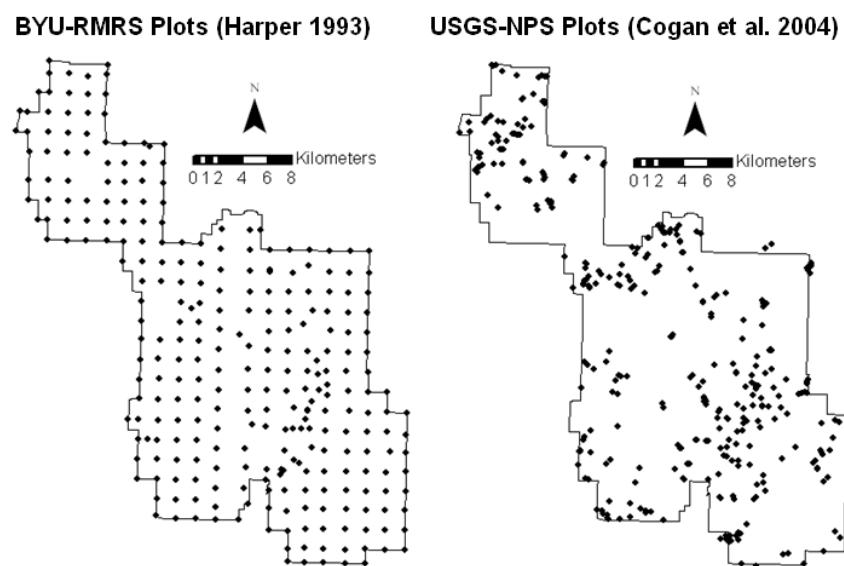
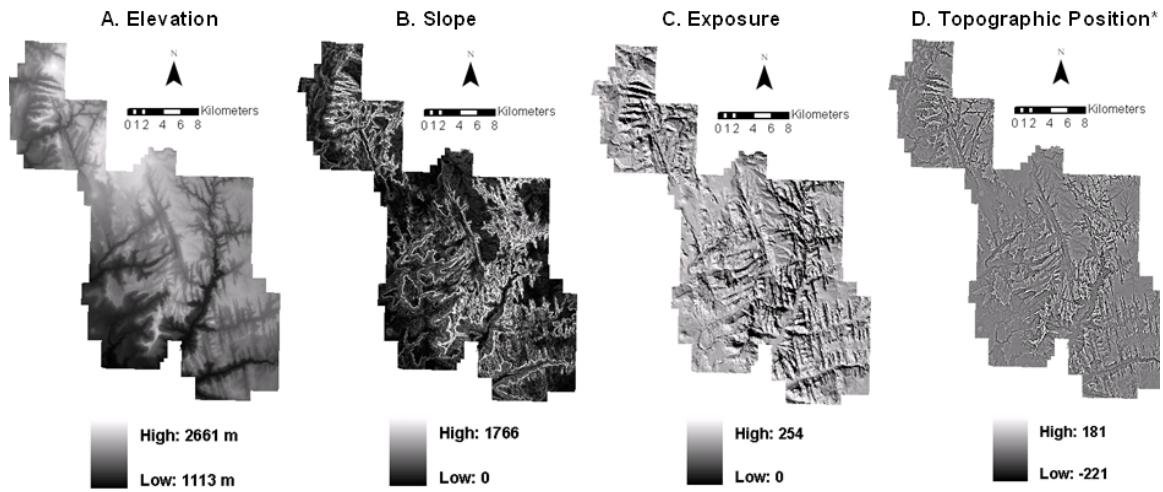


Figure 2.2. Plot locations of two vegetation survey datasets at Zion National Park



^aBased on the Topographic Position Index (TPI) at a 50 m radius (Weiss 2001)

Figure 2.3. Abiotic environmental variable layers derived from a 10-m resolution digital elevation model of Zion National Park

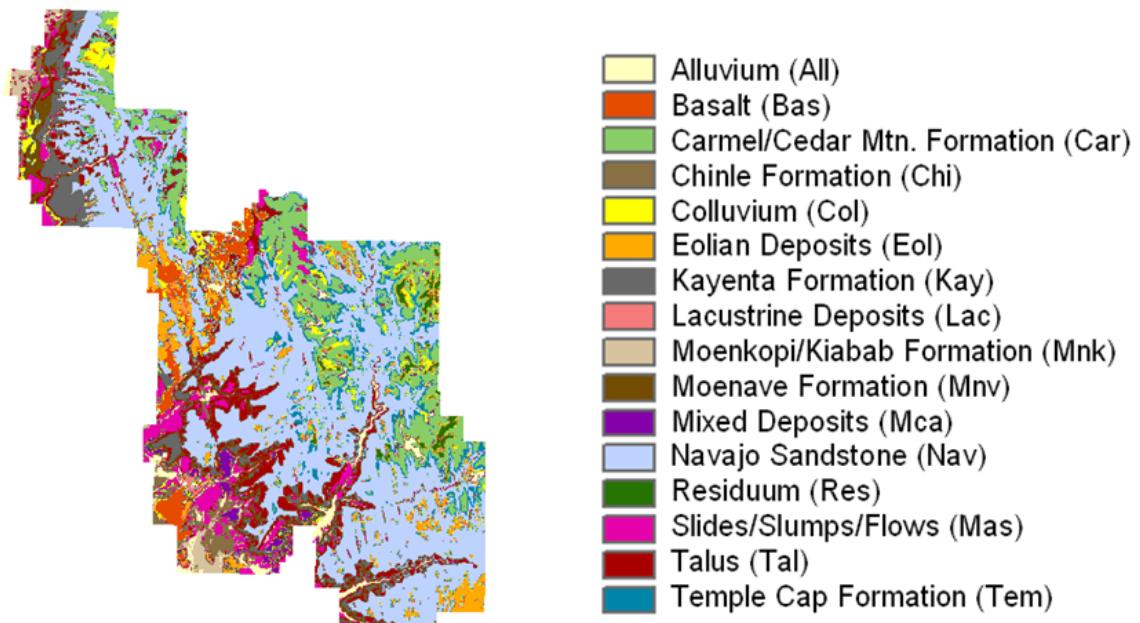


Figure 2.4. Geologic substrate classes derived from a 1:24,000 scale digital geologic map of Zion National Park and vicinity (O'Meara 2006)

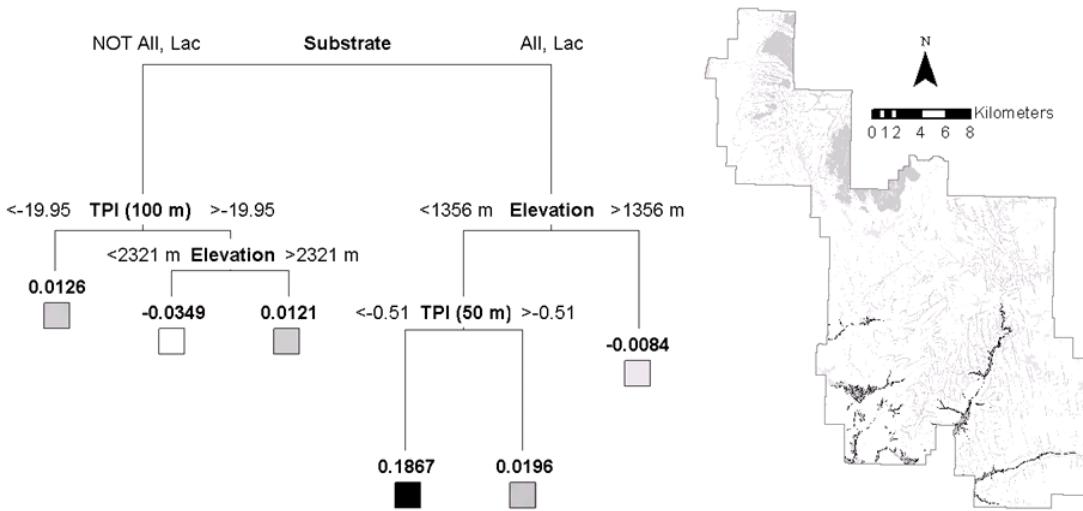


Figure 2.5. Regression tree model and map for the **Streambank** coalition group. Tree model shows relationships between plot affinities and environmental variables (*Substrate*: All=alluvium, Lac=lacustrine (see Table 2.1); *TPI* (50 or 100 m) =Topographic Position Index calculated at 50 or 100 m radius). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

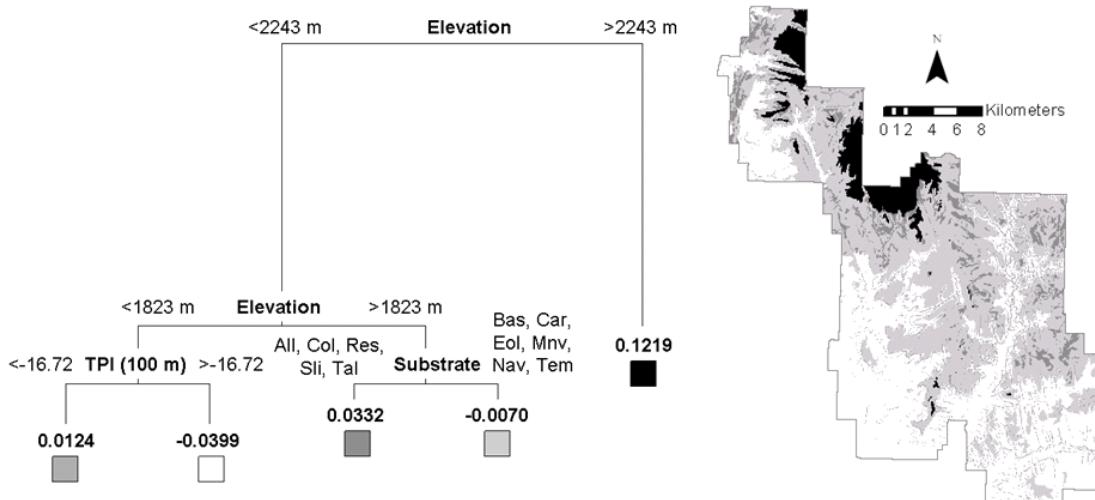


Figure 2.6. Regression tree model and map for the **High Plateau** coalition group. Tree model shows relationships between plot affinities and environmental variables (*Substrate*: see Table 2.1 for translation of abbreviations; *TPI* (100 m) =Topographic Position Index calculated at 100 m radius). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

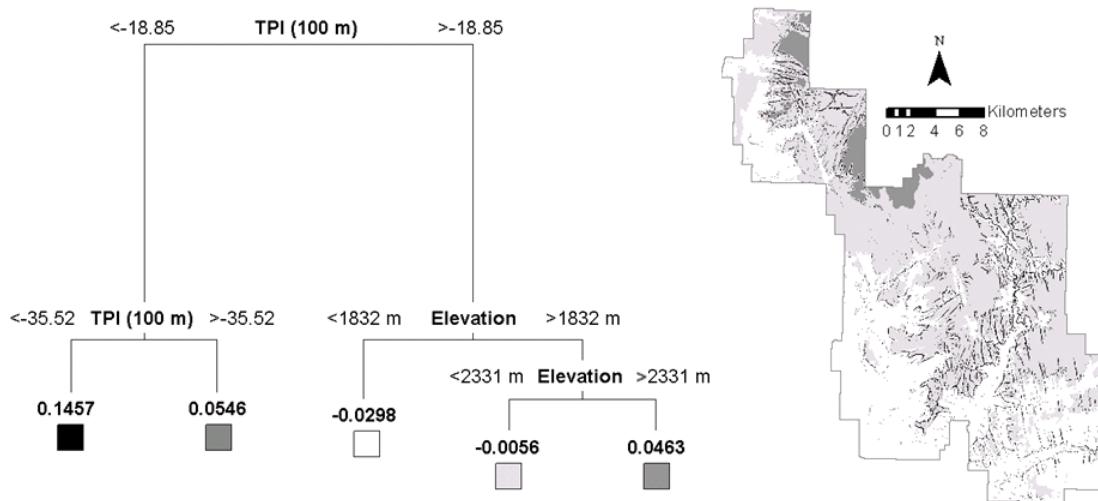


Figure 2.7. Regression tree model and map for the **Crevice Canyon** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables ($TPI (100 m)$ = Topographic Position Index calculated at 100 m radius). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

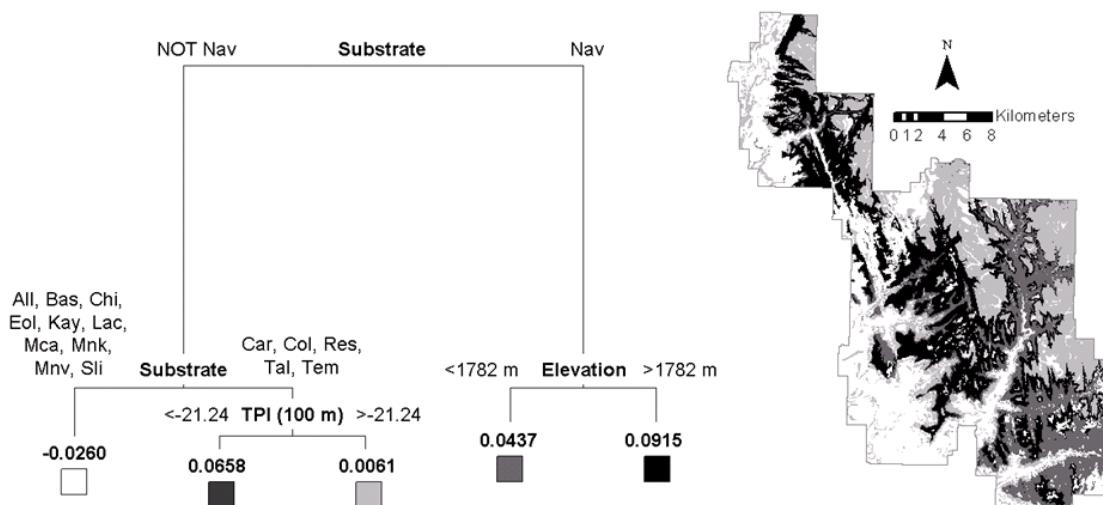


Figure 2.8. Regression tree model and map for the **Slickrock** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables (Substrate: Nav=Navajo, see Table 2.1 for translation of other abbreviations; $TPI (100 m)$ = Topographic Position Index calculated at 100 m radius). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

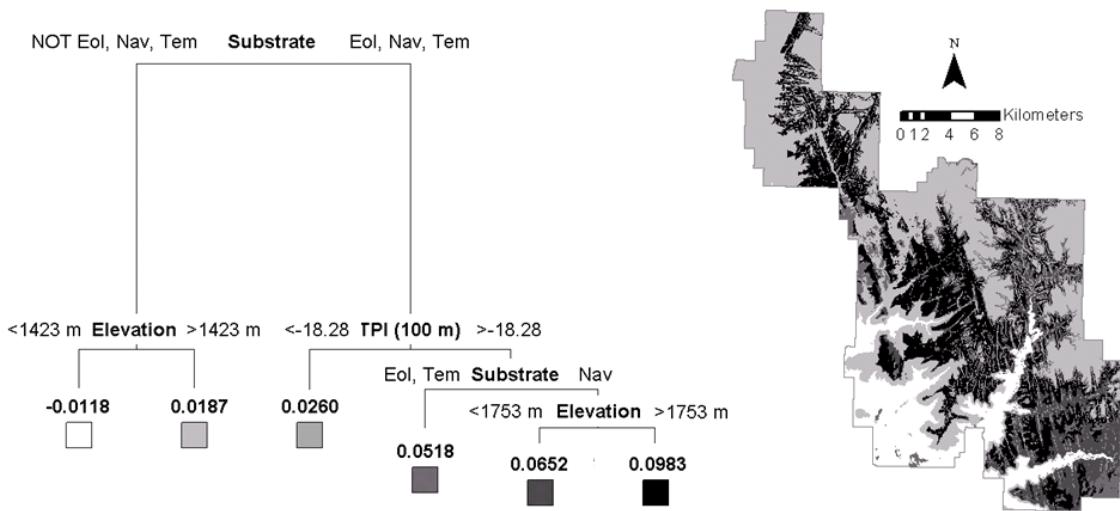


Figure 2.9. Regression tree model and map for the **Upland Sands** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables (*Substrate*: *Eol*=Eolian, *Nav*=Navajo Sandstone, *Tem*=Temple Cap, see Table 2.1; *TPI (100 m)*=Topographic Position Index calculated at 100 m radius). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

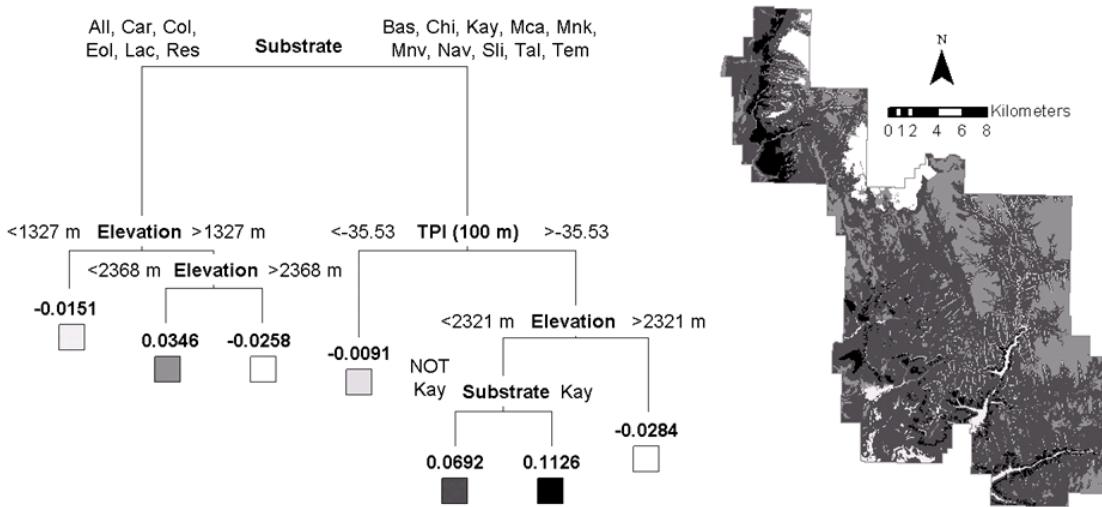


Figure 2.10. Regression tree model and map for the **Rocky Slopes** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables (*Substrate*: *Kay*=Kayenta, see Table 2.1 for translation of other abbreviations; *TPI (100 m)*=Topographic Position Index calculated at 100 m radius). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

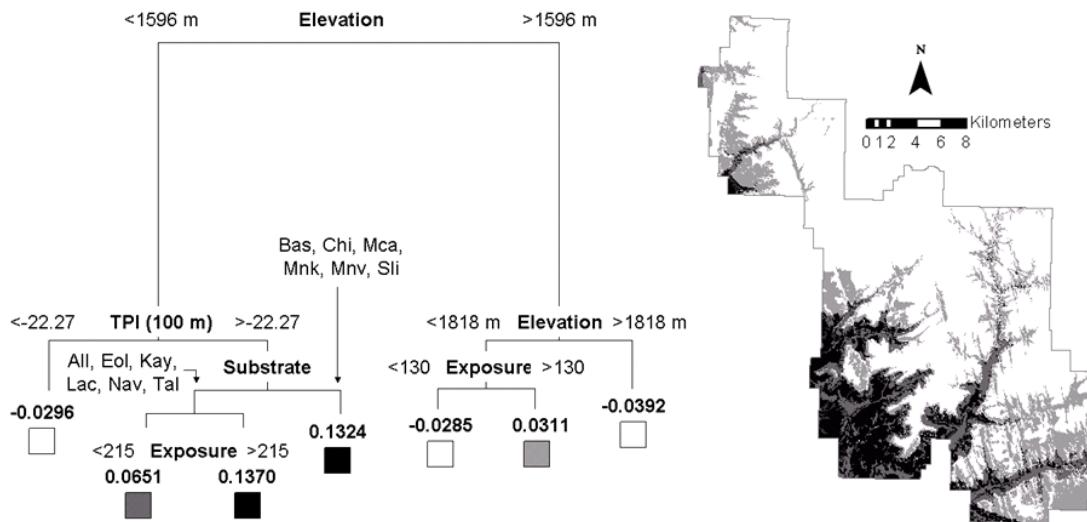


Figure 2.11. Regression tree model and map for the **Arid Lowlands** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables (Substrate: see Table 2.1 for translation of abbreviations; TPI (100 m) = Topographic Position Index calculated at 100 m radius; Exposure=solar hillshade at 215° azimuth and 45° altitude). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

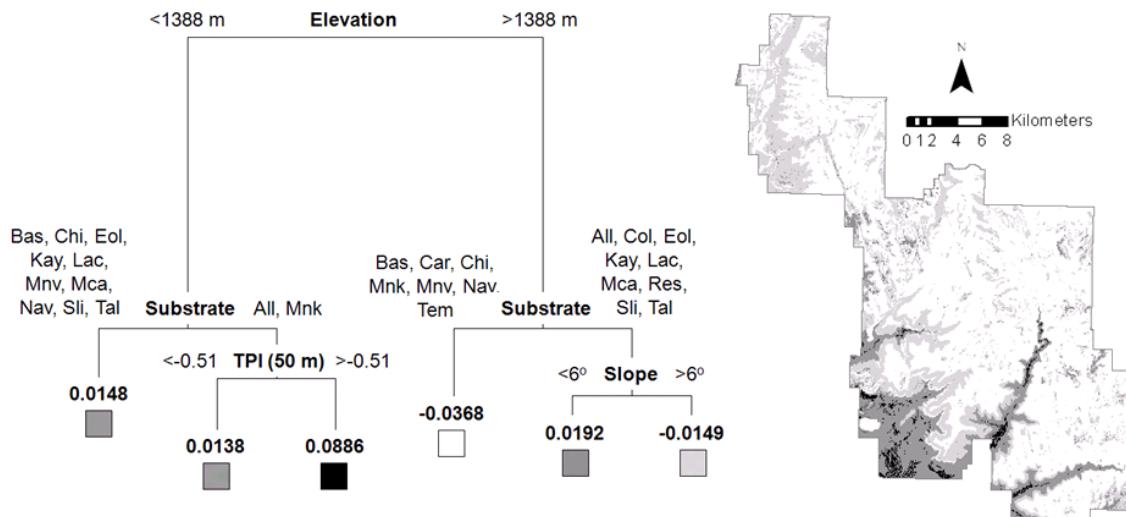


Figure 2.12. Regression tree model and map for the **Lowland Flats** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables (Substrate: see Table 2.1 for translation of abbreviations; TPI (50 m) = Topographic Position Index calculated at 50 m radius). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

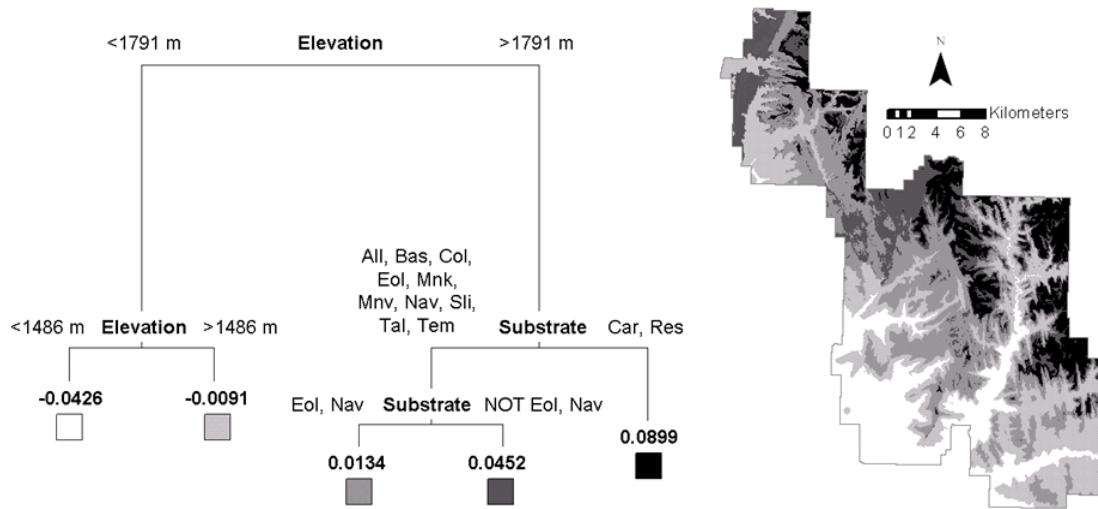


Figure 2.13. Regression tree model and map for the **Mesic Mesa Top** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables (*Substrate*: *Car*=Carmel, *Eol*=Eolian, *Nav*=Navajo, *Res*=Residuum, see Table 2.1 for translation of other abbreviations). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

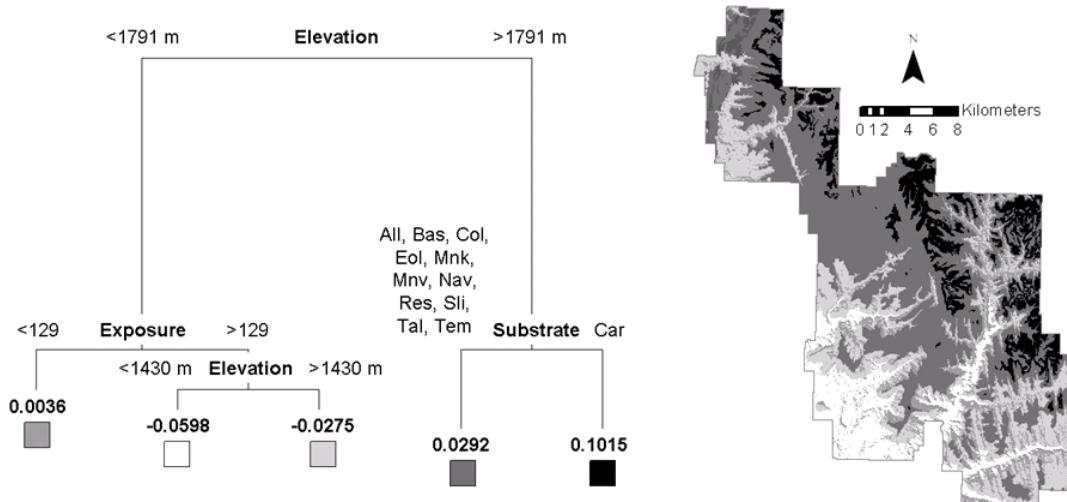


Figure 2.14. Regression tree model and map for the **Xeric Mesa Top** coalition group. Tree model shows relationships between plot affinities and abiotic environmental variables (*Substrate*: *Car*=Carmel, see Table 2.1 for translation of other abbreviations; *Exposure*=solar hillshade at 215° azimuth and 45° altitude). Mean plot affinity values at terminal nodes are in units of the phi coefficient; higher values (darker shading) = higher affinity. Predicted distribution of affinity values in Zion National Park are mapped using shading shown at tree tips.

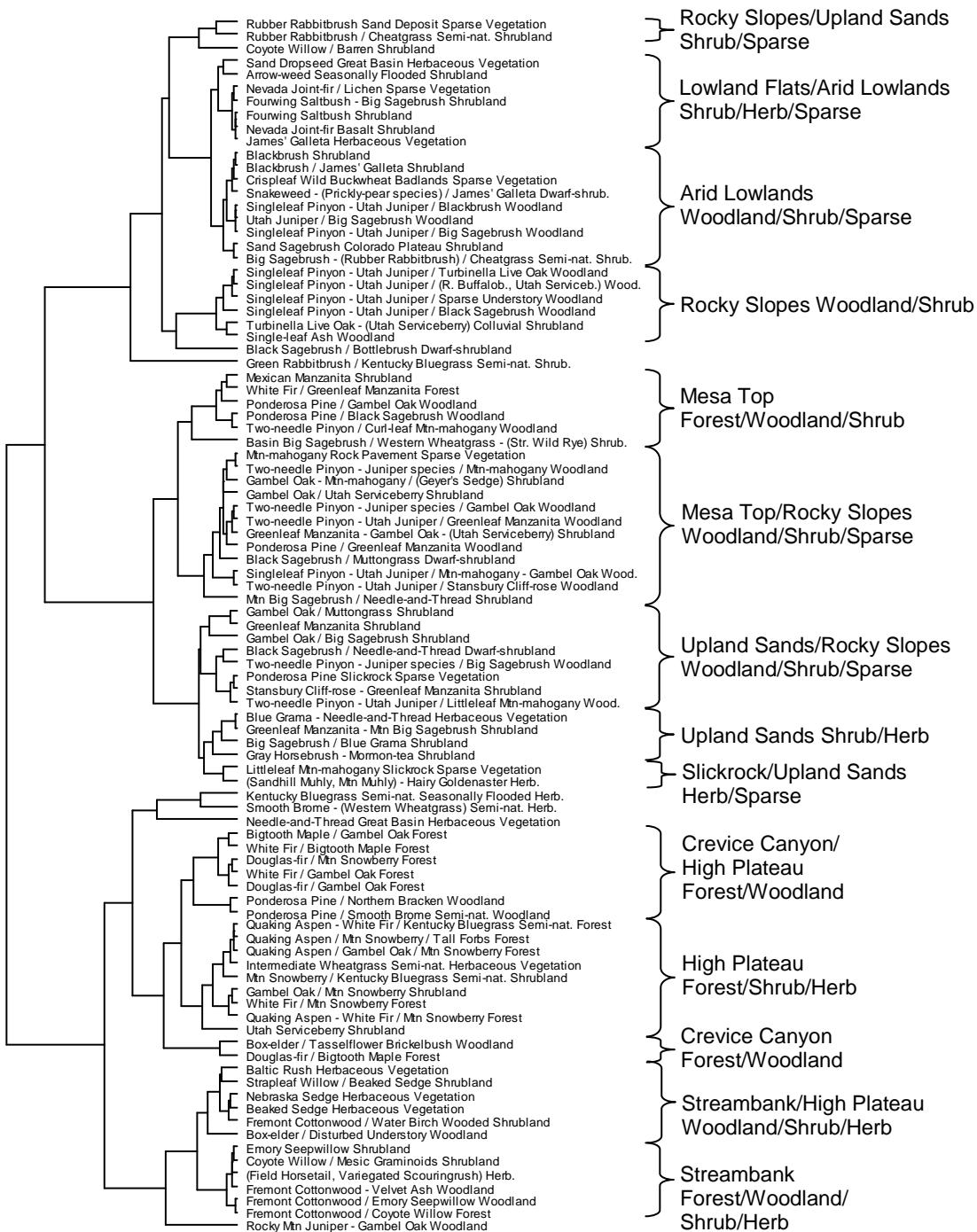


Figure 2.15. Cluster dendrogram of National Vegetation Classification System (NVCS) associations of Zion National Park (Cogan et al. 2004) based on compositional affinities to coalition species groups presented in this paper. Similarities in affinity patterns across species groups were quantified for each NVCS association pair using Pearson's correlation coefficient, then clustered using average linkage hierarchical clustering (McCune and Grace 2002) in R 2.10.1 (R Development Core Team 2010). Names assigned to major clusters (right) combine environmental and physiognomic descriptors.

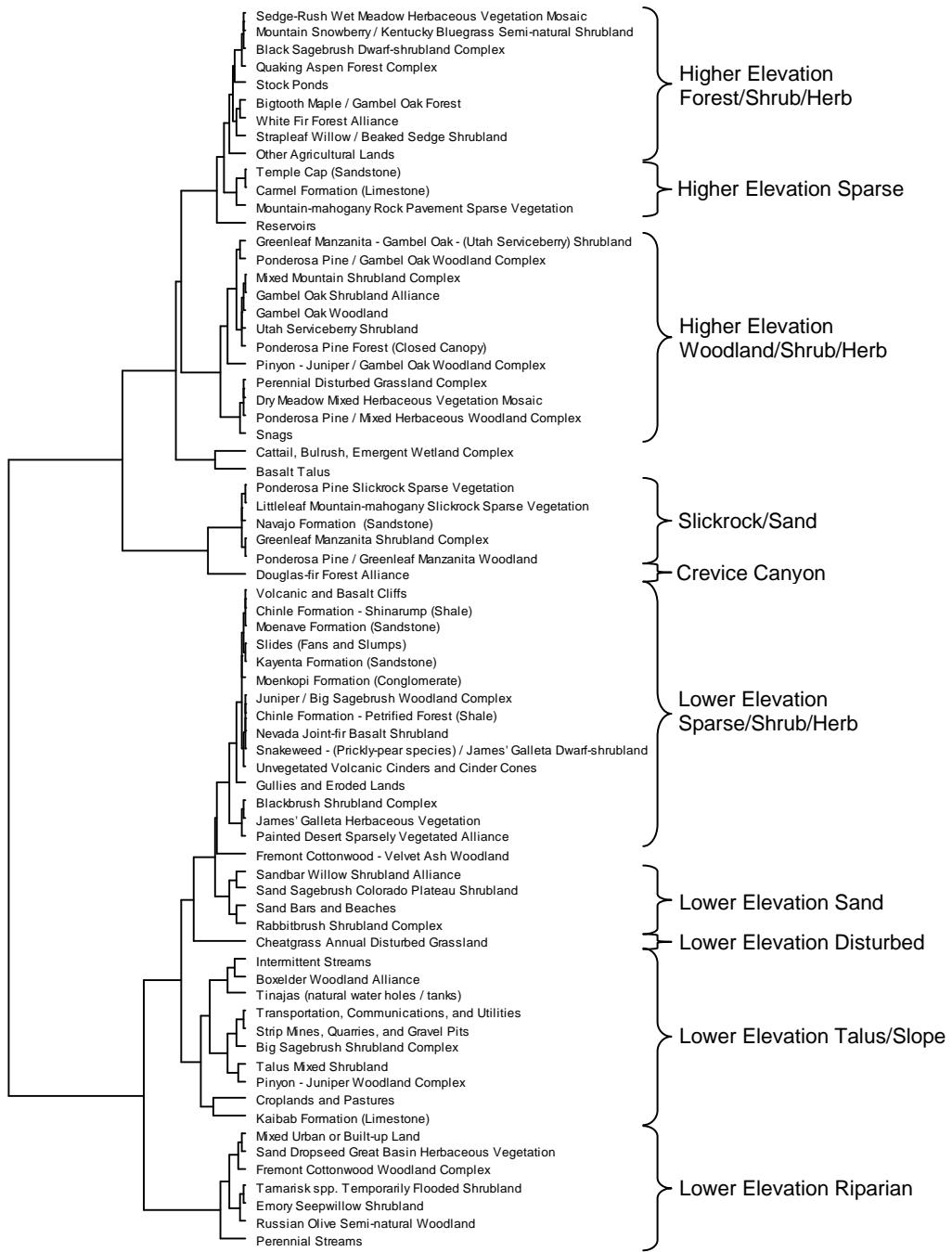


Figure 2.16. Cluster dendrogram of Zion National Park vegetation map units (Cogan et al. 2004) based on pixel-averaged affinities to coalition species groups presented in this paper. Similarities in affinity patterns across species groups were quantified for each map unit pair using Pearson's correlation coefficient, then clustered using average linkage hierarchical clustering (McCune and Grace 2002) in R 2.10.1(R Development Core Team 2010). Names assigned to major clusters (right) describe their elevational/environmental/physiognomic characteristics.

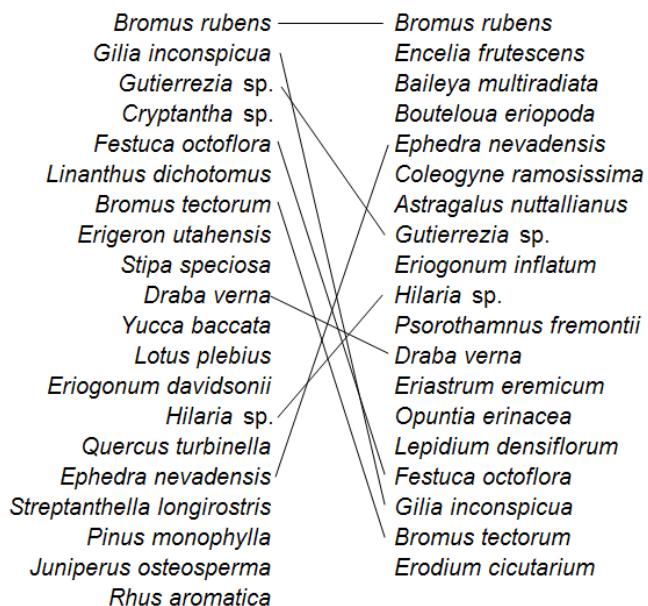


Figure 2.17. Two Arid Lowlands species groups of Zion National Park obtained through coalition clustering in RCLUS at a threshold affinity value of phi=0.2. Species are in descending order of group affinity and lines connect species shared by both groups. The left group has higher elevation/Great Basin affinities and the right group has lower elevation/Mojave Desert affinities.

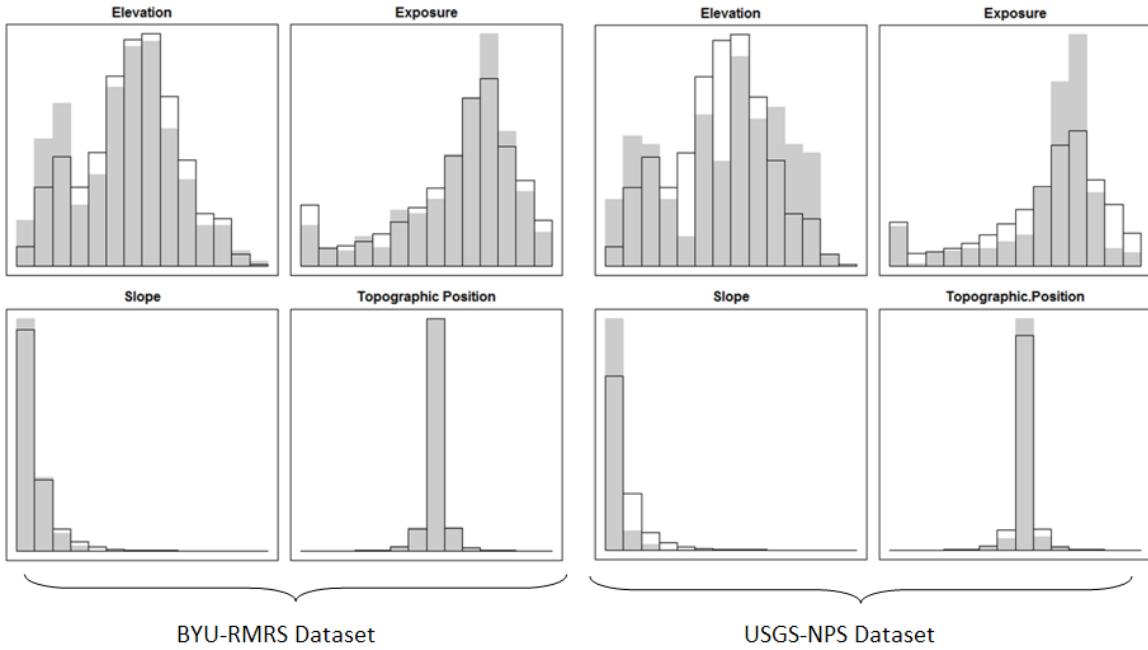


Figure 2.18. Histograms of proportions of background pixels representing four topographic variables at Zion National Park (black line bars) against pixels represented by plots (shaded bars) for the BYU-RMRS dataset (left) and USGS-NPS dataset (right). Based on 50 m pixel resolution generalizations of the original 10 m pixel resolution grids covering the park. Topographic position is TPI at 50 m radius. Created using the adehabitat package (Calenge 2006) in R 2.8.0 (R Development Core Team 2008).

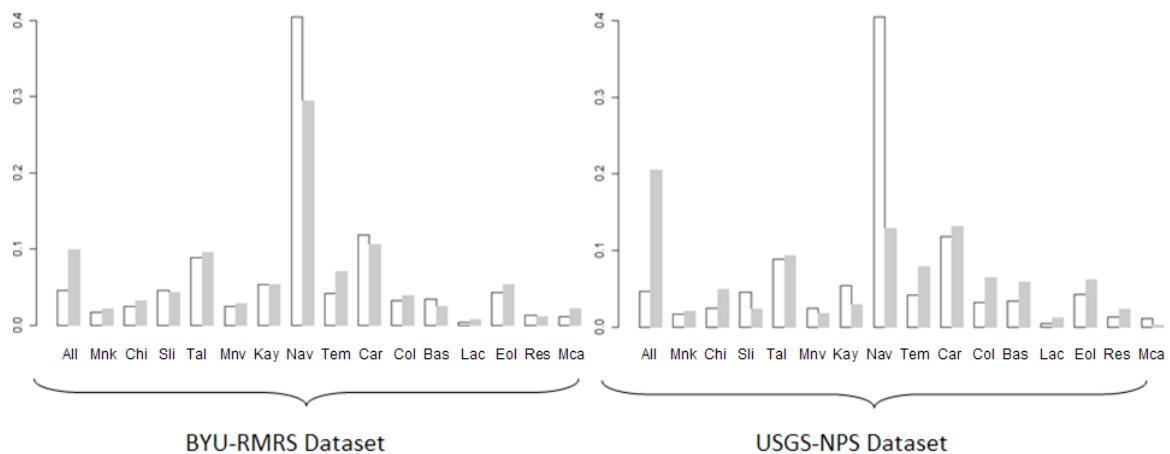


Figure 2.19. Histograms of proportions of background pixels in different geologic substrate classes (see Table 2.1) at Zion National Park (white bars) against pixels represented by plots (shaded bars) for the BYU-RMRS dataset (left) and USGS-NPS dataset (right). Based on 50 m pixel resolution generalizations of the original 10 m pixel resolution grids covering the park. Created using the adehabitat package (Calenge 2006) in R 2.8.0 (R Development Core Team 2008).

Table 2.1. Geologic substrate classes used for environmental modeling of plant species group affinities at Zion National Park. For each substrate class, corresponding map units from a digital geologic map of the Utah Geological Survey (O'Meara 2006) are shown in the rightmost column. Map unit symbols in bold type were represented by one or more plots of the 1987-1989 vegetation survey dataset (Harper 1993).

Code	Substrate Class	Description	Map Units Included in Class
All	Alluvium	river/stream deposits	Qa, Qa1 , Qa2, Qac , Qaco, Qae , Qaec, Qaeo, Qaes, Qaf, Qaf1, Qaf2, Qafc , Qafco, Qafm, Qafo, Qafy, Qafy , Qage , Qal1 , Qaly, Qam , Qao, Qao, Qap2, Qas, Qaso, Qat2 , Qat2, Qat3 , Qat4, Qat5 , Qat6, Qath , Qatm, Qato, Qats , Qav, Qay, Qf
Bas	Basalt	volcanics (flows, cinders, ash)	Qbc , Qbca , Qbcc , Qbg , Qbgc, Qbhr, Qblc, Qblp , Qbp , Qms(b)
Car	Carmel Formation (includes Cedar Mountain Formation)	limestone, shale, sandstone, siltstone, gypsum	Jccl , Jccu , Jcp , Jcw, Jcx , Kcmc
Chi	Chinle Formation	mudstone, claystone, siltstone, sandstone, conglomerate	TRcp , TRcp(s) , TRcs
Col	Colluvium	unconsolidated fine gravity deposits	Qc , Qce, Qces , Qco
Eol	Eolian Deposits	unconsolidated wind deposits	Qea , Qed , Qer , Qes
Kay	Kayenta Formation	siltstone, sandstone, mudstone	Jk , Jkt
Lac	Lacustrine Deposits	lake deposits	Qla , Qlbc , Qls
Mca	Mass Movement/ Colluvium/Alluvial Pediment-mantle Deposits	massive gravity deposits mixed with fine gravity deposits and river/stream deposits	Qmcp1 , Qmcp2 , Qmcp3
Mnk	Moenkopi Formation (includes Kaibab Formation)	siltstone, mudstone, limestone, sandstone, conglomerate, gypsum, dolomite	TRm , TRml, TRmm, TRmr, TRms, TRmt, TRmu, TRmv , Pkf, Pkh
Mnv	Moenave Formation	mudstone, sandstone, siltstone	Jm , Jm(s), Jmd , Jmk(s), Jms , Jmw
Nav	Navajo Sandstone	sandstone	Jn , Jnb , Jnl , Jnp , Jnw
Res	Residuum	deposits from <i>in situ</i> weathering	Qre , Qrlc
Sli	Mass Movement (Slide/Slump/Flow) Deposits	massive gravity deposits (slides, slumps and flows)	Qmfy, Qmsc , Qmsh, Qmso , Qmsy
Tal	Talus	coarse gravity deposits	Qmt , Qmts , QTng
Tem	Temple Cap Formation	sandstone, siltstone, mudstone	Jt, Jts , Jtw

Table 2.2. Coalition species groups and affiliate species of Zion National Park, based on coalition clustering of 1987-89 vegetation survey data. #Occ.=Number of plot occurrences out of a total of 288 plots. Affin.=Affinity of a coalition species to its coalition group, calculated as the mean value of the phi coefficient of association between the species and others in the group. Affil.=Affiliation of an affiliate species with coalition species, in terms of percent co-occurrence. Cvr.=Mean cover of affiliate species weighted by co-occurrences with coalition species (see text for details). Species in boldface are both coalition species and affiliate species for a given group. Underlined species are dominant affiliates (weighted mean cover \geq 2%)

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
-------------------	-------	--------	-------------------	-------	--------	------

Table 2.2A. Streambank

<i>Baccharis salicina</i>	3	0.4594	<u><i>Acer negundo</i></u>	18	0.5345	2.34
<i>Salix exigua</i>	3	0.4594	<u><i>Bromus tectorum</i></u>	130	0.5086	5.10
<i>Tamarix chinensis</i>	5	0.4269	<u><i>Bromus diandrus</i></u>	10	0.4655	8.28
<i>Populus fremontii</i>	6	0.3936	<u><i>Equisetum hyemale</i></u>	8	0.4397	0.79
<i>Equisetum hyemale</i>	8	0.3744	<i>Artemisia ludoviciana</i>	51	0.4224	0.36
<i>Verbascum thapsus</i>	5	0.3743	<u><i>Poa pratensis</i></u>	14	0.4224	4.28
<i>Sonchus</i> sp.	3	0.3652	<i>Poa fendleriana</i>	190	0.4052	1.78
<i>Agrostis exarata</i>	3	0.3453	<i>Phacelia heterophylla</i>	42	0.4052	0.27
<i>Bromus diandrus</i>	10	0.3353	<u><i>Elymus trachycaulus</i></u>	10	0.3966	0.76
<i>Juncus arcticus</i>	3	0.3103	<u><i>Populus fremontii</i></u>	6	0.3966	4.63
<i>Elymus canadensis</i>	4	0.2742	<i>Chrysopsis villosa</i>	74	0.3879	1.65
<i>Acer negundo</i>	18	0.2513	<i>Tamarix chinensis</i>	5	0.3707	0.44
<i>Bromus carinatus</i>	9	0.2511	<i>Bromus carinatus</i>	9	0.3621	0.52
<i>Poa compressa</i>	3	0.2434	<i>Verbascum thapsus</i>	5	0.3534	0.18
<i>Elymus trachycaulus</i>	10	0.2409	<u><i>Quercus gambelii</i></u>	135	0.3362	5.75
<i>Poa pratensis</i>	14	0.2027	<u><i>Baccharis salicina</i></u>	3	0.3017	2.28
<i>Datura wrightii</i>	3	0.2023	<u><i>Salix exigua</i></u>	3	0.3017	4.98
<i>Fraxinus velutina</i>	6	0.1862	<u><i>Elymus canadensis</i></u>	4	0.2931	0.15
			<i>Gutierrezia</i> sp.	94	0.2759	0.33
			<i>Sonchus</i> sp.	3	0.2500	0.13
			<i>Agrostis exarata</i>	3	0.2414	0.12
			<u><i>Fraxinus velutina</i></u>	6	0.2328	4.19
			<i>Artemisia</i> sp.	2	0.2328	0.44
			<i>Melilotus</i> sp.	2	0.2328	0.12
			<i>Oenothera longissima</i>	2	0.2328	0.12

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2B. High Plateau						
<i>Stellaria jamesiana</i>	18	0.3314	<i>Quercus gambelii</i>	135	0.8069	22.85
<i>Senecio eremophilus</i>	3	0.3109	<i>Symporicarpos oreophilus</i>	58	0.6455	10.96
<i>Lupinus sericeus</i>	9	0.2937	<i>Vicia americana</i>	43	0.6138	5.11
<i>Prunus virginiana</i>	10	0.2923	<i>Poa fendleriana</i>	190	0.6081	2.92
<i>Mertensia arizonica</i>	7	0.2907	<i>Polygonum douglasii</i>	39	0.4524	0.35
<i>Bromus carinatus</i>	9	0.2714	<i>Amelanchier utahensis</i>	141	0.4409	3.08
<i>Amelanchier alnifolia</i>	11	0.2711	<i>Carex rossii</i>	67	0.4265	0.44
<i>Stipa nelsonii</i>	5	0.2604	<i>Stellaria jamesiana</i>	18	0.4207	2.42
<i>Stipa lettermanii</i>	12	0.2524	<i>Arabis perennans</i>	124	0.3487	0.17
<i>Vicia americana</i>	43	0.2497	<i>Senecio multilobatus</i>	99	0.3199	0.17
<i>Poa pratensis</i>	14	0.2466	<i>Juniperus scopulorum</i>	21	0.3112	0.91
<i>Rosa woodsii</i>	12	0.2403	<i>Abies concolor</i>	19	0.3055	3.36
<i>Achillea millefolium</i>	6	0.2048	<i>Phacelia heterophylla</i>	42	0.2939	0.19
<i>Taraxacum officinale</i>	11	0.2013	<i>Poa pratensis</i>	14	0.2853	4.39
<i>Symporicarpos oreophilus</i>	58	0.2005	<i>Eriogonum racemosum</i>	59	0.2824	0.14
<i>Thalictrum fendleri</i>	12	0.1990	<i>Solidago velutina</i>	38	0.2738	1.20
<i>Chenopodium fremontii</i>	3	0.1983	<i>Prunus virginiana</i>	10	0.2709	0.72
<i>Abies concolor</i>	19	0.1859	<i>Stipa lettermanii</i>	12	0.2651	0.28
<i>Elymus trachycaulus</i>	10	0.1829	<i>Amelanchier alnifolia</i>	11	0.2651	0.68
<i>Juniperus scopulorum</i>	21	0.1754	<i>Acer grandidentatum</i>	29	0.2622	2.69
<i>Polygonum douglasii</i>	39	0.1711	<i>Rosa woodsii</i>	12	0.2594	0.74
<i>Osmorhiza occidentalis</i>	4	0.1617	<i>Opuntia macrorhiza</i>	120	0.2565	0.36
<i>Bromus anomalus</i>	5	0.1586	<i>Pachystima myrsinites</i>	54	0.2565	1.61
<i>Erigeron speciosus</i>	6	0.1538				

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2C. Crevice Canyon						
<i>Eupatorium herbaceum</i>	12	0.3612	<i>Quercus gambelii</i>	135	0.6871	7.06
<i>Viola canadensis</i>	5	0.3300	<i>Poa fendleriana</i>	190	0.6742	4.52
<i>Cystopteris fragilis</i>	6	0.3184	<i>Acer grandidentatum</i>	29	0.6032	6.17
<i>Acer negundo</i>	18	0.3163	<i>Acer negundo</i>	18	0.5161	5.54
<i>Polypodium hesperium</i>	6	0.2726	<i>Eupatorium herbaceum</i>	12	0.4806	0.24
<i>Galium triflorum</i>	4	0.2616	<i>Pachystima myrsinites</i>	54	0.4581	3.26
<i>Acer grandidentatum</i>	29	0.2614	<i>Pseudotsuga menziesii</i>	18	0.4548	4.13
<i>Rubus leucodermis</i>	4	0.2611	<i>Holodiscus dumosus</i>	21	0.4419	0.87
<i>Bromus ciliatus</i>	12	0.2602	<i>Solidago velutina</i>	38	0.4387	0.35
<i>Pseudotsuga menziesii</i>	18	0.2559	<i>Abies concolor</i>	19	0.4355	5.46
<i>Heuchera rubescens</i>	12	0.2533	<i>Draba asprella</i>	33	0.3903	0.20
<i>Smilacina racemosa</i>	6	0.2531	<i>Artemisia ludoviciana</i>	51	0.3871	0.41
<i>Mimulus guttatus</i>	3	0.2451	<i>Arabis perennans</i>	124	0.3806	0.19
<i>Taraxacum officinale</i>	11	0.2364	<i>Heuchera rubescens</i>	12	0.3806	0.19
<i>Philadelphus microphyllus</i>	10	0.2350	<i>Bromus ciliatus</i>	12	0.3774	0.25
<i>Disporum trachycarpum</i>	4	0.2342	<i>Stephanomeria tenuifolia</i>	51	0.3742	0.24
<i>Thalictrum fendleri</i>	12	0.2305	<i>Zauschneria latifolia</i>	15	0.3710	0.19
<i>Elymus canadensis</i>	4	0.2267	<i>Selaginella underwoodii</i>	17	0.3645	0.67
<i>Abies concolor</i>	19	0.2236	<i>Eriogonum racemosum</i>	59	0.3613	0.19
<i>Epilobium glandulosum</i>	3	0.2225	<i>Chrysopsis villosa</i>	74	0.3452	0.73
<i>Goodyera oblongifolia</i>	5	0.2131	<i>Senecio multilobatus</i>	99	0.3323	0.17
<i>Muhlenbergia racemosa</i>	3	0.2055	<i>Penstemon rostriflorus</i>	30	0.3323	0.17
<i>Aralia racemosa</i>	3	0.2012	<i>Mahonia repens</i>	29	0.3258	0.51
<i>Sphaeromeria ruthiae</i>	3	0.1957	<i>Thalictrum fendleri</i>	12	0.3258	0.44
<i>Zauschneria latifolia</i>	15	0.1956	<i>Philadelphus microphyllus</i>	10	0.3258	0.69
<i>Holodiscus dumosus</i>	21	0.1947	<i>Taraxacum officinale</i>	11	0.2903	0.15
<i>Chimaphila menziesii</i>	5	0.1943	<i>Phacelia heterophylla</i>	42	0.2871	0.29
<i>Epilobium ciliatum</i>	3	0.1889				
<i>Epilobium brachycarpum</i>	3	0.1830				
<i>Selaginella underwoodii</i>	17	0.1728				
<i>Brickellia grandiflora</i>	10	0.1723				
<i>Erigeron sionis</i>	4	0.1662				
<i>Aster welshii</i>	6	0.1633				
<i>Pteridium aquilinum</i>	3	0.1626				
<i>Equisetum hyemale</i>	8	0.1603				
<i>Agrostis exarata</i>	3	0.1570				

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2D. Slickrock						
<i>Petrophytum caespitosum</i>	12	0.3143	<i>Poa fendleriana</i>	190	0.8627	4.15
<i>Holodiscus dumosus</i>	21	0.3078	<i>Chrysopsis villosa</i>	74	0.6139	1.07
<i>Cercocarpus intricatus</i>	40	0.3022	<i>Quercus gambelii</i>	135	0.5574	5.61
<i>Castilleja scabrida</i>	40	0.2903	<i>Senecio multilobatus</i>	99	0.5574	0.31
<i>Muhlenbergia thurberi</i>	12	0.2838	<i>Opuntia macrorhiza</i>	120	0.5347	0.49
<i>Selaginella underwoodii</i>	17	0.2741	<i>Pinus ponderosa</i>	60	0.5347	5.31
<i>Ivesia sabulosa</i>	7	0.2693	<i>Cercocarpus intricatus</i>	40	0.5073	1.55
<i>Zauschneria latifolia</i>	15	0.2515	<i>Stephanomeria tenuifolia</i>	51	0.5040	0.36
<i>Arenaria fendleri</i>	23	0.2407	<i>Castilleja scabrida</i>	40	0.4927	0.25
<i>Erigeron canaani</i>	16	0.2338	<i>Amelanchier utahensis</i>	141	0.4863	1.71
<i>Eriogonum jamesii</i>	10	0.2322	<i>Arabis perennans</i>	124	0.4766	0.24
<i>Erigeron sionis</i>	4	0.2217	<i>Arctostaphylos patula</i>	94	0.4733	5.50
<i>Leptodactylon pungens</i>	8	0.2160	<i>Arenaria macradenia</i>	46	0.4087	0.55
<i>Stephanomeria tenuifolia</i>	51	0.2097	<i>Carex rossii</i>	67	0.3958	0.28
<i>Draba asprella</i>	33	0.2068	<i>Solidago velutina</i>	38	0.3700	0.29
<i>Trifolium longipes</i>	10	0.2031	<i>Draba asprella</i>	33	0.3619	0.18
<i>Penstemon humilis</i>	27	0.2021	<i>Holodiscus dumosus</i>	21	0.3393	0.58
<i>Pinus ponderosa</i>	60	0.1993	<i>Eriogonum racemosum</i>	59	0.3215	0.17
<i>Brickellia grandiflora</i>	10	0.1977	<i>Penstemon humilis</i>	27	0.3150	0.23
<i>Chrysopsis villosa</i>	74	0.1966	<i>Arenaria fendleri</i>	23	0.3086	0.20
<i>Monardella odoratissima</i>	6	0.1961	<i>Chaenactis douglasii</i>	44	0.3053	0.15
<i>Solidago velutina</i>	38	0.1890	<i>Phlox austromontana</i>	54	0.2956	0.42
<i>Linanthastrum nuttallii</i>	19	0.1762	<i>Pachystima myrsinites</i>	54	0.2827	1.43
<i>Sedum lanceolatum</i>	3	0.1751	<i>Selaginella underwoodii</i>	17	0.2811	0.92
<i>Heuchera rubescens</i>	12	0.1744	<i>Bromus tectorum</i>	130	0.2682	0.58
<i>Wyethia arizonica</i>	5	0.1696				
<i>Arenaria macradenia</i>	46	0.1502				

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2E. Upland Sands						
<i>Cryptantha cinerea</i>	17	0.2622	<i>Poa fendleriana</i>	190	0.8255	5.08
<i>Chaenactis douglasii</i>	44	0.2527	<i>Opuntia macrorhiza</i>	120	0.7224	1.15
<i>Abronia fragrans</i>	20	0.2521	<i>Senecio multilobatus</i>	99	0.6388	0.37
<i>Hymenopappus filifolius</i>	11	0.2352	<i>Arctostaphylos patula</i>	94	0.6230	10.05
<i>Artemisia campestris</i>	14	0.2341	<i>Amelanchier utahensis</i>	141	0.5515	2.35
<i>Penstemon laevis</i>	26	0.2316	<i>Quercus gambelii</i>	135	0.5188	6.57
<i>Tradescantia occidentalis</i>	22	0.2178	<i>Chrysopsis villosa</i>	74	0.5006	1.48
<i>Arctostaphylos patula</i>	94	0.2147	<i>Arabis perennans</i>	124	0.4800	0.25
<i>Opuntia macrorhiza</i>	120	0.2100	<i>Erysimum asperum</i>	64	0.4291	0.21
<i>Eriogonum alatum</i>	6	0.2047	<i>Pinus ponderosa</i>	60	0.4085	5.62
<i>Chrysopsis villosa</i>	74	0.1933	<i>Phlox austromontana</i>	54	0.4061	0.60
<i>Senecio multilobatus</i>	99	0.1919	<i>Bromus tectorum</i>	130	0.3903	1.07
<i>Bouteloua gracilis</i>	19	0.1887	<i>Chaenactis douglasii</i>	44	0.3855	0.19
<i>Phlox austromontana</i>	54	0.1886	<i>Juniperus osteosperma</i>	115	0.3564	2.21
<i>Arenaria fendleri</i>	23	0.1880	<i>Gilia inconspicua</i>	84	0.3406	0.22
<i>Linanthastrum nuttallii</i>	19	0.1760	<i>Festuca octoflora</i>	74	0.3345	0.27
<i>Pinus ponderosa</i>	60	0.1735	<i>Eriogonum racemosum</i>	59	0.3091	0.16
<i>Erysimum asperum</i>	64	0.1601	<i>Stephanomeria tenuifolia</i>	51	0.2970	0.22
<i>Polygonum douglasii</i>	39	0.1548	<i>Carex rossii</i>	67	0.2885	0.22
			<i>Polygonum douglasii</i>	39	0.2776	0.14
			<i>Sporobolus cryptandrus</i>	50	0.2703	0.17
			<i>Arenaria macradenia</i>	46	0.2655	0.32
			<i>Yucca angustissima</i>	41	0.2618	0.29
			<i>Stipa hymenoides</i>	65	0.2545	0.17
			<i>Machaeranthera canescens</i>	40	0.2545	0.14
			<i>Penstemon laevis</i>	26	0.2545	0.25

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2F. Rocky Slopes						
<i>Fraxinus anomala</i>	45	0.2408	<i>Poa fendleriana</i>	190	0.8536	5.78
<i>Arabis perennans</i>	124	0.2379	<i>Amelanchier utahensis</i>	141	0.6651	3.47
<i>Erigeron utahensis</i>	41	0.2246	<i>Arabis perennans</i>	124	0.6500	0.35
<i>Poa fendleriana</i>	190	0.2132	<i>Opuntia macrorhiza</i>	120	0.6126	0.99
<i>Quercus turbinella</i>	50	0.2131	<i>Juniperus osteosperma</i>	115	0.5784	4.05
<i>Erysimum asperum</i>	64	0.2064	<i>Senecio multilobatus</i>	99	0.5084	0.28
<i>Opuntia macrorhiza</i>	120	0.1969	<i>Bromus tectorum</i>	130	0.5004	1.31
<i>Pinus monophylla</i>	64	0.1964	<i>Quercus gambelii</i>	135	0.4582	6.98
<i>Juniperus osteosperma</i>	115	0.1868	<i>Gilia inconspicua</i>	84	0.4320	0.24
<i>Haplopappus scopolorum</i>	42	0.1804	<i>Arctostaphylos patula</i>	94	0.4161	6.24
<i>Shepherdia rotundifolia</i>	17	0.1785	<i>Erysimum asperum</i>	64	0.3835	0.19
<i>Penstemon eatonii</i>	23	0.1760	<i>Pinus monophylla</i>	64	0.3715	2.94
<i>Amelanchier utahensis</i>	141	0.1661	<i>Gutierrezia sp.</i>	94	0.3691	0.52
<i>Senecio multilobatus</i>	99	0.1615	<i>Quercus turbinella</i>	50	0.3190	3.64
<i>Gilia inconspicua</i>	84	0.1513	<i>Fraxinus anomala</i>	45	0.3047	0.91
<i>Streptanthus cordatus</i>	38	0.1502	<i>Festuca octoflora</i>	74	0.2999	0.24
			<i>Cryptantha</i> sp. (annual)	60	0.2872	0.17
			<i>Erigeron utahensis</i>	41	0.2808	0.18
			<i>Stipa hymenoides</i>	65	0.2792	0.22
			<i>Chrysopsis villosa</i>	74	0.2745	0.82
			<i>Eriogonum racemosum</i>	59	0.2673	0.15
			<i>Haplopappus scopolorum</i>	42	0.2673	0.54
			<i>Phlox austromontana</i>	54	0.2649	0.50
			<i>Pinus edulis</i>	53	0.2474	1.84

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2G. Arid Lowlands						
<i>Bromus rubens</i>	43	0.3680	<i>Bromus tectorum</i>	130	0.7848	3.42
<i>Gutierrezia sp.</i>	94	0.2817	<i>Gutierrezia sp.</i>	94	0.7039	1.26
<i>Baileya multiradiata</i>	8	0.2654	<i>Juniperus osteosperma</i>	115	0.6865	4.06
<i>Ephedra nevadensis</i>	18	0.2637	<i>Gilia inconspicua</i>	84	0.6424	0.34
<i>Draba verna</i>	40	0.2595	<i>Poa fendleriana</i>	190	0.6107	3.16
<i>Gilia inconspicua</i>	84	0.2558	<i>Festuca octoflora</i>	74	0.5820	0.43
<i>Hilaria sp.</i>	41	0.2507	<i>Arabis perennans</i>	124	0.4908	0.27
<i>Festuca octoflora</i>	74	0.2480	<i>Cryptantha sp. (annual)</i>	60	0.4836	0.27
<i>Coleogyne ramosissima</i>	13	0.2448	<i>Bromus rubens</i>	43	0.4826	0.55
<i>Opuntia erinacea</i>	17	0.2280	<i>Pinus monophylla</i>	64	0.4518	2.56
<i>Bouteloua eriopoda</i>	5	0.2274	<i>Opuntia macrorhiza</i>	120	0.4498	0.73
<i>Eriogonum palmerianum</i>	15	0.2175	<i>Draba verna</i>	40	0.3955	0.21
<i>Astragalus nuttallianus</i>	9	0.2167	<i>Amelanchier utahensis</i>	141	0.3924	1.97
<i>Bromus tectorum</i>	130	0.2135	<i>Hilaria sp.</i>	41	0.3822	1.10
<i>Cryptantha sp. (annual)</i>	60	0.2085	<i>Stipa hymenoides</i>	65	0.3268	0.34
<i>Eriastrum eremicum</i>	7	0.2058	<i>Quercus turbinella</i>	50	0.3064	3.53
<i>Psorothamnus fremontii</i>	11	0.2031	<i>Elymus elymoides</i>	58	0.3043	0.19
<i>Encelia frutescens</i>	4	0.1980	<i>Erigeron utahensis</i>	41	0.2777	0.16
<i>Yucca baccata</i>	17	0.1913	<i>Arctostaphylos patula</i>	94	0.2756	4.00
<i>Stipa speciosa</i>	15	0.1855	<i>Descurainia pinnata</i>	28	0.2541	0.17
<i>Eriastrum sparsiflorum</i>	13	0.1752	<i>Artemisia tridentata</i>	55	0.2377	1.70
<i>Swertia albomarginata</i>	11	0.1751	<i>Stipa comata</i>	39	0.2377	1.09
<i>Descurainia pinnata</i>	28	0.1716	<i>Eriogonum davidsonii</i>	32	0.2357	0.14
<i>Rhus aromatica</i>	23	0.1709	<i>Senecio multilobatus</i>	99	0.2285	0.13
<i>Juniperus osteosperma</i>	115	0.1706				
<i>Eriogonum inflatum</i>	3	0.1657				
<i>Linanthus dichotomus</i>	14	0.1643				
<i>Pinus monophylla</i>	64	0.1617				

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2H. Lowland Flats						
<i>Lycium pallidum</i>	3	0.3355	<i>Bromus tectorum</i>	130	0.7609	18.04
<i>Erodium cicutarium</i>	10	0.2865	<i>Gutierrezia</i> sp.	94	0.5761	1.98
<i>Atriplex canescens</i>	10	0.2722	<i>Chamaesyce albomarginata</i>	23	0.5435	1.20
<i>Tragopogon dubius</i>	8	0.2444	<i>Juniperus osteosperma</i>	115	0.4783	1.62
<i>Lactuca serriola</i>	13	0.2237	<i>Sphaeralcea grossulariifolia</i>	14	0.3913	0.22
<i>Cirsium wheeleri</i>	8	0.2224	<i>Lactuca serriola</i>	13	0.3804	0.19
<i>Chamaesyce albomarginata</i>	23	0.2085	<i>Erodium cicutarium</i>	10	0.3804	0.82
<i>Sphaeralcea grossulariifolia</i>	14	0.2053	<i>Hilaria</i> sp.	41	0.3587	1.30
<i>Elymus smithii</i>	3	0.1929	<i>Atriplex canescens</i>	10	0.3478	0.91
			<i>Poa fendleriana</i>	190	0.3152	1.34
			<i>Sporobolus cryptandrus</i>	50	0.3043	0.48
			<i>Festuca octoflora</i>	74	0.2826	0.17
			<i>Stipa comata</i>	39	0.2826	1.41
			<i>Tragopogon dubius</i>	8	0.2826	0.14
			<i>Gilia inconspicua</i>	84	0.2717	0.14
			<i>Amelanchier utahensis</i>	141	0.2609	1.51
			<i>Cirsium wheeleri</i>	8	0.2609	0.18
			<i>Quercus gambelii</i>	135	0.2391	2.61
			<i>Elymus elymoides</i>	58	0.2391	0.15
			<i>Bromus rubens</i>	43	0.2283	0.35
			<i>Cryptantha</i> sp. (annual)	60	0.2174	0.11
			<i>Artemisia tridentata</i>	55	0.2174	1.77

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2I. Mesic Mesa Top						
<i>Lathyrus lanszwertii</i>	20	0.2432	<i>Amelanchier utahensis</i>	141	0.7787	4.26
<i>Peraphyllum ramosissimum</i>	10	0.2389	<i>Quercus gambelii</i>	135	0.7787	15.05
<i>Balsamorhiza sagittata</i>	8	0.2200	<i>Poa fendleriana</i>	190	0.7753	4.27
<i>Phlox longifolia</i>	11	0.2187	<i>Arabis perennans</i>	124	0.4854	0.25
<i>Penstemon linarioides</i>	13	0.2105	<i>Opuntia macrorhiza</i>	120	0.4545	0.78
<i>Swertia radiata</i>	11	0.1982	<i>Carex rossii</i>	67	0.4477	0.46
<i>Zigadenus paniculatus</i>	12	0.1945	<i>Senecio multilobatus</i>	99	0.4322	0.24
<i>Purshia tridentata</i>	36	0.1832	<i>Juniperus osteosperma</i>	115	0.4254	2.51
<i>Quercus gambelii</i>	135	0.1804	<i>Symporicarpos oreophilus</i>	58	0.4099	3.30
<i>Symporicarpos oreophilus</i>	58	0.1700	<i>Pinus edulis</i>	53	0.3911	3.59
<i>Mertensia fusiformis</i>	3	0.1672	<i>Arctostaphylos patula</i>	94	0.3774	5.28
<i>Chrysothamnus depressus</i>	5	0.1618	<i>Pachystima myrsinites</i>	54	0.3293	1.46
<i>Amelanchier utahensis</i>	141	0.1598	<i>Purshia tridentata</i>	36	0.2916	0.71
<i>Carex rossii</i>	67	0.1553	<i>Pinus ponderosa</i>	60	0.2813	4.00
<i>Pinus edulis</i>	53	0.1513	<i>Erysimum asperum</i>	64	0.2693	0.13
			<i>Bromus tectorum</i>	130	0.2676	0.78
			<i>Vicia americana</i>	43	0.2642	1.55
			<i>Phlox austromontana</i>	54	0.2453	0.69
			<i>Eriogonum racemosum</i>	59	0.2384	0.13
			<i>Cercocarpus montanus</i>	33	0.2316	2.02
			<i>Lathyrus lanszwertii</i>	20	0.2161	0.87
			<i>Solidago velutina</i>	38	0.2041	0.40

Coalition Species	#Occ.	Affin.	Affiliate Species	#Occ.	Affil.	Cvr.
Table 2.2J. Xeric Mesa Top						
<i>Viola purpurea</i>	10	0.2702	<i>Amelanchier utahensis</i>	141	0.7964	4.21
<i>Cercocarpus montanus</i>	33	0.2588	<i>Quercus gambelii</i>	135	0.7696	13.94
<i>Pinus edulis</i>	53	0.2160	<i>Poa fendleriana</i>	190	0.7464	4.12
<i>Carex rossii</i>	67	0.2114	<i>Arabis perennans</i>	124	0.5000	0.25
<i>Pachystima myrsinites</i>	54	0.2096	<i>Carex rossii</i>	67	0.4768	0.57
<i>Amelanchier utahensis</i>	141	0.1990	<i>Opuntia macrorhiza</i>	120	0.4393	0.79
<i>Swertia radiata</i>	11	0.1911	<i>Senecio multilobatus</i>	99	0.4357	0.24
<i>Pedicularis centranthera</i>	10	0.1849	<i>Pachystima myrsinites</i>	54	0.4286	1.85
<i>Peraphyllum ramosissimum</i>	10	0.1847	<i>Arctostaphylos patula</i>	94	0.4107	5.48
<i>Quercus gambelii</i>	135	0.1792	<i>Juniperus osteosperma</i>	115	0.4089	2.65
<i>Physaria newberryi</i>	7	0.1630	<i>Pinus edulis</i>	53	0.4089	3.96
<i>Mahonia repens</i>	29	0.1614	<i>Symphoricarpos oreophilus</i>	58	0.3250	2.40
			<i>Cercocarpus montanus</i>	33	0.3125	2.91
			<i>Pinus ponderosa</i>	60	0.2732	3.61
			<i>Bromus tectorum</i>	130	0.2500	0.76
			<i>Erysimum asperum</i>	64	0.2464	0.12
			<i>Phlox austromontana</i>	54	0.2375	0.59
			<i>Vicia americana</i>	43	0.2375	1.24
			<i>Mahonia repens</i>	29	0.2357	0.27
			<i>Eriogonum racemosum</i>	59	0.2196	0.12
			<i>Fraxinus anomala</i>	45	0.2196	0.64

Table 2.3. Affinity correspondence between vegetation units of Harper et al. (2001) (rows) and coalition species groups of the current study (columns) at Zion National Park. Low mean phi values (green shading) indicate low correspondence between vegetation types and species groups; high values (red shading) indicate high correspondence.

	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slick- rock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream- bank
Abandoned fields	0.1726	0.0687	-0.0333	-0.0366	-0.0442	-0.0663	-0.0533	-0.0325	-0.0317	-0.0069
Blackbrush	0.0229	0.2062	0.0450	-0.0295	-0.0590	-0.0892	-0.0784	-0.0590	-0.0532	-0.0242
Sagebrush	0.0375	0.0867	0.0134	-0.0054	-0.0468	-0.0721	-0.0455	-0.0301	-0.0447	-0.0120
Juniper-Pinyon	-0.0455	0.0264	0.1427	-0.0056	-0.0223	0.0657	0.0424	-0.0569	-0.0375	-0.0657
Mountain Brush	-0.0400	-0.0401	0.0991	0.1252	0.0159	-0.0146	0.0051	0.0260	0.0093	0.0023
Ponderosa Pine	-0.0711	-0.0720	0.0858	0.1516	0.0809	0.0447	0.0603	-0.0199	-0.0300	-0.0549
Rock Crevice	-0.0530	-0.0255	0.0674	0.1320	0.1637	-0.0338	-0.0361	-0.0477	0.0052	-0.0412
Douglas Fir-White Fir	-0.0375	-0.0928	-0.0117	-0.0095	0.0665	0.0894	0.0605	0.0974	0.1080	-0.0084
Hanging Gardens	-0.0148	-0.0308	-0.0264	-0.0098	0.0574	-0.0105	-0.0148	0.0125	0.0962	0.0468
Riparian	-0.0054	-0.0420	-0.0459	-0.0284	0.0239	-0.0411	-0.0425	0.0356	0.1286	0.1821

Table 2.4. Affinity correspondence between National Vegetation Classification System (NVCS) formation classes of Cogan et al. (2004) (rows) and coalition species groups of the current study (columns) at Zion National Park. Low mean phi values (green shading) indicate low correspondence between formation classes and species groups; high values (red shading) indicate high correspondence.

	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slick- rock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream- bank
I. FOREST	-0.0221	-0.0631	-0.0204	-0.0039	0.0225	0.0458	0.0420	0.0940	0.0731	0.0296
II. WOODLAND	-0.0214	0.0078	0.0556	0.0232	-0.0078	0.0141	0.0177	-0.0100	-0.0156	-0.0121
III. SHRUBLAND	0.0038	0.0165	0.0327	0.0261	-0.0134	-0.0091	0.0001	-0.0021	-0.0167	0.0016
IV. DWARF-SHRUBLAND	0.0131	0.0374	0.0472	0.0251	-0.0051	-0.0149	0.0081	-0.0214	-0.0372	-0.0307
V. HERBACEOUS VEGETATION	0.0225	0.0002	-0.0047	0.0153	-0.0067	-0.0270	-0.0080	0.0352	0.0040	0.0606
VII. SPARSE VEGETATION	0.0063	0.0454	0.0554	0.0237	0.0062	-0.0128	-0.0099	-0.0432	-0.0321	-0.0351

Table 2.5. Affinity correspondence between lumped vegetation map units of Cogan et al. (2004) (rows) and coalition species groups of the current study (columns) at Zion National Park. Low mean phi values (green shading) indicate low correspondence between map units and species groups; high values (red shading) indicate high correspondence.

	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slick- rock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream- bank
Bare - Soil/Stone Formations	-0.0115	0.0579	0.0575	0.0102	-0.0094	-0.0083	-0.0074	-0.0054	-0.0118	-0.0238
Grasslands	0.0100	0.0124	0.0246	0.0151	-0.0160	-0.0038	0.0061	0.0100	-0.0084	0.0005
Higher Elevation Forest	-0.0277	-0.0325	0.0329	0.0370	0.0183	0.0370	0.0385	0.0356	0.0154	-0.0176
Higher Elevation Shrub	-0.0286	-0.0312	0.0362	0.0326	0.0093	0.0475	0.0494	0.0356	0.0056	-0.0221
Lower Elevation Forest (P/J)	-0.0105	0.0493	0.0623	0.0151	-0.0076	-0.0091	-0.0034	-0.0257	-0.0205	-0.0284
Lower Elevation Shrub	0.0116	0.0517	0.0405	0.0106	-0.0151	-0.0235	-0.0151	-0.0189	-0.0218	-0.0085
Riparian/Flooded Woodland	-0.0002	0.0208	0.0243	0.0145	-0.0066	-0.0143	-0.0067	0.0025	0.0009	0.0310
Slick Rock	-0.0364	-0.0065	0.0669	0.0812	0.0678	0.0064	0.0033	-0.0203	-0.0123	-0.0317

CHAPTER 3

PHYLOGENETIC NICHE OVERLAP IN PLANT COMMUNITIES OF ZION NATIONAL PARK

ABSTRACT

Question: Because species sharing common ancestry are likely to have similar niches (phylogenetic niche conservatism), species distributions within and among communities may be non-random with respect to phylogeny. Related species are likely to be segregated at small spatial scales because of alpha niche conservatism, yet aggregated within habitats because of beta niche conservatism. To what extent are these patterns evident across lineages and phylogenetic scales in plant communities?

Location: Zion National Park, Utah, U.S.A.

Methods: Alpha- and beta-niche overlap were calculated for all sister groups of a genus-level phylogeny and all congeneric species pairs from a vegetation survey dataset. Alpha-niche was defined at the plot scale, while beta-niche overlap was defined in relation to vegetation classes. A randomization procedure that re-allocated species to plots according to spatial and environmental proximity criteria was used to test statistical significance of each overlap.

Results: Of the 341 congeneric species pairs tested, 42 and 34 showed significant beta-niche aggregation in tests using presence and cover data, respectively. Fewer congeneric species pairs (8 for presence, 7 for cover) showed significant alpha-niche segregation patterns. For sister groups above genus (231 total), an even smaller proportion of tests

were significant (8 and 10 for beta niche presence and cover; 1 and 6 for alpha niche presence and cover). A few examples of beta-niche aggregation coupled with alpha-niche aggregation were noted but none were simultaneously significant for both. On the other hand, other patterns suggesting various types of niche divergence were common. Many related taxa were segregated in their beta as well as alpha-niche, suggesting divergence of habitat affinities driven by competitive interactions. Others were aggregated in both niche types, suggesting evolution of coexistence mechanisms.

Conclusions: These results draw attention to the complex mixture of niche conservatism and divergence in plant lineages and demonstrate that niche conservatism cannot be uncritically invoked in community assembly studies.

INTRODUCTION

Studies addressing phylogenetic patterns in ecological communities have the potential to provide greater insight than those based simply on species patterns (Webb et al. 2002, Webb et al. 2006, Emerson and Gillespie 2008, Cavender-Bares et al. 2009, Vamosi et al. 2009). Because ecological traits have an evolutionary origin, and may be preserved over evolutionary time through phylogenetic niche conservatism (Harvey and Pagel 1991), species sharing common ancestry may assemble themselves into communities in predictable ways. In an influential review paper, Webb et al. (2002) described two potential consequences of phylogenetic niche conservatism on community structure: (1) phylogenetically-related species are likely to be segregated (spatially overdispersed) at small spatial scales corresponding to local neighborhoods of interacting individuals because conserved niche similarity should result in competitive exclusion;

although (2) at larger spatial scales encompassing habitat variation, related species are likely to be aggregated (spatially clustered) due to conserved similarities in their habitat affinities (Webb et al. 2002; see also Cavender-Bares et al. 2009, Vamosi et al. 2009). By consolidating these predictions into an explicitly phylogenetic model and offering analytical tools for measuring phylogenetic relatedness of entire communities, Webb et al. (2002) set in motion a subdiscipline of ‘community phylogenetics’ (Swenson et al. 2006) intent on testing for phylogenetic segregation and aggregation through empirical analysis of field data (Webb et al 2006, Cavender-Bares et al 2009, Vamosi et al. 2009).

Terrestrial plant communities have been particularly well represented in phylogenetic community studies, apparently in part because of their stationary spatial structure and ease of sampling (Vamosi et al. 2009). The sample units of plant community studies (vegetation plots) are typically spatially defined areas within which plant species composition and abundance can be exhaustively inventoried at a given point in time. Appropriately-sized vegetation plots can be construed as local neighborhoods of interacting plant species. Whittaker (1975) used the term *alpha diversity* to refer to species diversity at local scales exemplified by plots, and by extension the term *alpha niche* has been used to describe niche characteristics permitting (or not) local species coexistence (Pickett and Bazazz 1978, Silvertown et al. 2006). The *beta niche*, like *beta diversity*, refers to a broader scale of environmental differentiation, i.e. the scale of multiple plots arranged along an environmental gradient. Moisture requirements and edaphic preferences are examples of beta niche attributes, while resource acquisition strategy fits the alpha niche category (Silvertown 2004, Silvertown et al. 2006). The alpha niche translates to those aspects of the niche that Webb et al. (2002) predicted

would lead to local segregation of phylogenetically-related species, and the beta niche translates to habitat affinities that should lead to phylogenetic aggregation by habitat.

Empirical analyses of phylogenetic structure in plant communities have revealed instances of phylogenetic segregation and aggregation consistent with the alpha/beta-niche conservatism model (reviewed in Cavender-Bares et al. 2009, Vamosi et al. 2009). However, different phylogenetic structure patterns have been noted for different lineages, locations, and scales. Several authors have pointed out that taxonomic or phylogenetic scale (the number of lineages under consideration) affects the outcome of phylogenetic community analyses (Slingsby and Verboom 2006, Cavender-Bares et al. 2006, Swenson et al. 2006, 2007). Cavender-Bares et al. (2006), for example, observed alpha-scale phylogenetic segregation within certain narrowly-defined lineages (e.g. the genus *Quercus*, see also Cavender-Bares et al. 2004), but phylogenetic aggregation when all seed plants were considered simultaneously. These results suggest that the strength and/or effects of niche conservatism vary across lineages having different degrees of shared evolutionary history. One might expect the imprint of niche conservatism to be more pronounced within a narrow, recently-evolved lineage than a broad, ancient lineage that has had more time to diverge ecologically (Vamosi et al. 2009). But because different lineages have the potential to evolve at different rates and in different ways, the balance between niche conservatism and divergence may not be consistently related to lineage age (or other measures of phylogenetic relatedness). Given these considerations, phylogenetic community structure may not be as easily predicted as the alpha-beta niche conservatism model implies. Rather than asking whether phylogenetic aggregation or segregation is detectable in a community as a whole, one may need to dissect the

community to discover the specific lineages, scales and circumstances under which such patterns are evident.

This paper presents an analysis of phylogenetic community structure using data from Zion National Park, Utah, U.S.A. (Cogan et al. 2004). One of my objectives is simply to add an evolutionary dimension to previous species-based plant community characterizations of the park. I also wish to add to the body of studies that have examined phylogenetic community structure in other settings. However, unlike many previous studies my objective is not to quantify general patterns of phylogenetic structure using lineage age or other phylogenetic relatedness measures. Instead, I aim to document the extent to which patterns of aggregation and segregation can be found across different lineages and phylogenetic scales. I adopt a comprehensive exploratory approach examining patterns individually for each lineage and node of a community phylogeny. With this approach, phylogenetic relatedness (branch length) information is not required and community phylogenetic structure is re-cast as niche overlap between sister groups (cf. Warren et al. 2008). I explore niche overlap based on abundance (measured as cover) as well as presence/absence.

METHODS

Study Area and Plant Community Data

Zion National Park (Zion) is world-renowned for its geologic formations, including its massive colored sandstone cliffs, towers and canyons. Zion is also ecologically significant because of its environmental heterogeneity and high biological diversity. Numerous floristic studies and plant community/vegetation studies (e.g.,

Woodbury 1932, Meyer 1974, Welsh 1989, Harper 1993, Cogan et al. 2004, Fertig and Alexander 2009) have documented the geography and ecology of plants within the park. The current study builds on a vegetation survey carried out at Zion in 1999-2000 as part of the U.S. Geological Survey (USGS)-National Park Service (NPS) Vegetation Characterization and Mapping Program (Cogan et al. 2004).

The USGS-NPS vegetation survey at Zion utilized a gradsect strategy (Gillison and Brewer 1985) to stratify the park's environment into zones differentiated by elevation, substrate, hydrology, solar insolation and fire history (Cogan et al. 2004). Gradsect zones provided sampling targets for vegetation survey crews who ultimately placed plots at sites considered homogeneous and representative of major vegetation types. Plot size varied with vegetation type: 100 m² for vegetation dominated by herbs or dwarf shrubs, 400 m² for shrub- and tree-dominated vegetation. In some areas, e.g. narrow riparian zones, plots took a rectangular shape; otherwise plots were square or circular (Cogan et al. 2004). A Daubenmire (1959) cover class (or in some cases, a direct percent cover estimate) was recorded by species and vegetation stratum for vascular plant species occurring within plots. Eight vegetation strata were used: herbs, vines, three shrub size classes, and three tree size classes (Cogan et al. 2004). Additional species occurring near but not within plot boundaries were listed for some plots, but cover was not recorded for these species.

Cogan et al. (2004) used the USGS-NPS plot data to classify and map the vegetation of Zion following the U.S. National Vegetation Classification System (NVCS) (Grossman et al. 1998, Jennings et al. 2009). Multivariate analyses of species cover data in PC-ORD (McCune and Mefford 1997) guided the classification process (Cogan et al.

2004). At the time of data release in 2004, 332 of the 346 plots had been formally classified to the association and alliance levels of the NVCS (Cogan et al. 2004, USGS 2004). I grouped alliances into ten ‘ecological divisions’ (Table 3.1) that reflect major environmental settings at Zion, modified from ecological groupings imbedded in the digital vegetation map accompanying the vegetation classification (Cogan et al. 2004, USGS 2004). I used the combination of ecological divisions, alliances and associations to define habitat or beta-niche variation for testing phylogenetic niche overlap.

Taxonomic/phylogenetic Data Synthesis

Vascular plant species recorded by the USGS-NPS vegetation survey (Cogan et al. 2004), following Kartesz (1999), were standardized to the nomenclature of Welsh et al. (2003) for consistency with accepted usage by the U.S. National Park Service at Zion (Fertig and Alexander 2009) (Appendix 3.1). However, classification at the genus level and above followed the National Center for Biotechnology Information (NCBI 2008), which in some cases differed from Welsh et al. (2003) in name and/or circumscription (e.g. splitting of the genus *Aster*) (Appendix 3.1). The NCBI (2008) taxonomic classification provided a working hypothesis of phylogenetic relationships that was refined and modified where appropriate to match recent findings of plant systematists (Table 3.2). For angiosperms, ordinal phylogenetic relationships were based on APG III (APG 2009) and relationships of families within orders were obtained from Davis et al. 2004 (monocots), Tank et al. 2006 (Scrophulariaceae s.l.), Stevens 2008 (angiosperms in general) and Wang et al. 2009 (Ranunculales). Fern family relationships were obtained from Smith et al. (2006) and Schuettpelz and Pryer (2007). Nearly all polychotomies in a

genus-level phylogenetic supertree (Fig. 3.1) could be resolved using this approach¹.

Although I made no attempt to integrate branch length information into the phylogeny, I use taxonomic ranks as benchmarks of phylogenetic depth relative to the genus level.

Niche Overlap Calculations

Using species occurrence and cover data from the 332 classified USGS-NPS vegetation plots, combined with taxonomic/phylogenetic information for each species, I calculated niche overlap for all ‘sister’ group pairs for each dataset. The number of ‘sister’ groups was expanded to include all species pairs within genera (Appendix 3.2) as well as pairs at phylogenetic nodes above the genus level (Appendix 3.3, Fig. 3.1).

Niche overlap was calculated using the Renkonen index (Krebs 1999) which measures the overlapping area under two discretely-defined niche occupancy curves:

$$P_{jk} = \sum_{i=1}^n (\min p_{ij}, p_{ik})$$

where P_{jk} = Renkonen index between group i and group j (as a proportion)

p_{ij} = Proportion niche state i is of the total niche of group j

p_{ik} = Proportion niche state i is of the total niche of group k

n = Total number of niche states

As used here, ‘groups’ are congeners or sister groups and ‘niche states’ are spatial units representing variation of alpha and beta niches. For alpha niche overlap calculations, each vegetation plot was a single niche state. For beta niche calculations, niche states were defined by vegetation classes. Plots belonging to the same association were

¹ Available literature was insufficient to resolve ordinal relationships within the core lamiids (Gentianales, Lamiales/Solanales and Boraginaceae) and familial relationships within the Malpighiales (Euphorbiaceae, Linaceae, Salicaceae, and Violaceae). For the analysis presented here these groups were arbitrarily nested as (Gentianales, (Lamiales/Solanales, Boraginaceae)) and ((Euphorbiaceae, Linaceae), (Salicaceae, Violaceae))

considered to be identical beta niche units, whereas plots belonging to the same alliance but different associations were considered 2/3 similar, plots belonging to the same ecological division but different alliances were 1/3 similar, and plots belonging to different ecological divisions had a beta niche similarity of zero.

All niche overlap calculations were carried out twice, once for presence/absence (i.e. co-occurrence) and once for cover, using arithmetic midpoints of cover classes as percent cover values. Cover calculations posed an interesting challenge in that cover values of individual species and strata had to be aggregated at multiple levels. Cover values were first aggregated across strata for each species, then across species for each higher phylogenetic grouping. Because of overlapping cover within plots, simple summation of cover values was not satisfactory, leading to aggregated cover values in excess of 100%. This problem was overcome through a sequential, probabilistic algorithm as illustrated in Fig. 3.2. Cover values to be aggregated were sorted from highest to lowest and accreted in sequence. The question of how much additional cover to add with each additional species was resolved by assuming (given the absence of further information) that cover was proportionally distributed between overlapping and non-overlapping portions. Thus a species with 35% cover would contribute an additional 14% (0.35×0.40) to an existing total of 60% since the other 21% (0.35×0.60) is assumed to overlap with the existing total (Fig. 3.2).

Significance tests

The statistic of interest in this analysis was not niche overlap *per se* but the degree to which overlap was greater than expected (aggregation) or less than expected

(segregation) relative to null models. Using the R package for statistical computing (R Development Core Team 2009) I generated null distributions of alpha and beta niche overlap against which observed overlap values were compared. Plot occurrences (and corresponding cover values) were randomized 1000 times for each congeneric pair and sister group comparison. This procedure resembles niche overlap randomization tests in the EcoSim program (Gotelli and Entsminger 2009) except that I added spatial criteria to the randomization process. Re-allocation probabilities were not truly random but rather proportional to distance, such that an occurrence was more likely to be re-allocated to a plot in close proximity (including the plot where actually observed). In this way, dispersal effects were incorporated into null models and randomization tests were conservative with respect to pseudoreplication risks (cf. Wilson 2007).

For alpha niche overlap tests I imposed additional constraints on the randomization process to ensure that I was testing for overlap within rather than between habitats. I multiplied re-allocation probabilities based on spatial proximity by probabilities based on habitat similarity. Following the same scale used in niche overlap calculations (previous section), a similarity value of 3 was given to plots within the same association, 2 for plots in different associations of the same alliance, 1 for plots in different alliances of the same ecological division, and 0 for plots in different ecological divisions. Consequently, occurrences were never re-assigned to a different ecological division when testing alpha niche overlap, and were more likely to be re-assigned within their own alliance and association.

RESULTS AND DISCUSSION

Although the repetitive significance testing approach followed here does not adhere to strict statistical standards, it poses little problem in an exploratory framework (cf. Tukey 1980). As noted in my methods description, I expanded the exploratory realm to include comparisons of all possible congeneric species pairs. This was partially a practical move, since well-resolved phylogenies integrating all species of interest were unavailable for many plant genera at Zion. It also represents an attempt to look beyond the strict confines of bifurcating phylogenetic models. Since hybridization and introgression are known to occur among closely-related species in many plant groups (Stebbins 1969, Linder and Rieseberg 2004, Whitney et al. 2010), reticulate evolutionary models may in many cases be more accurate representations of intra-generic relationships than phylogenetic trees. Diffuse comparisons among all congeneric species pairs leave this possibility open for examination.

Niche overlap test results for congeneric species pairs (Table 3.3A, Appendix 3.2) and phylogenetic sister groups (Table 3.3B, Fig. 3.3-3.6, Appendix 3.3) are given as probabilities ranging from -1 to 1. Probability values above and below zero indicate greater aggregation and segregation, respectively, than expected relative to null models, and values above 0.95 and below -0.95 correspond to $p < 0.05$ in a two-tailed significance test. Only a portion of tests were statistically significant and many did not even deviate from the null expectation (Table 3.3). Alpha niche overlap tests, in particular, had a high number of null results, suggesting that these tests were particularly conservative and/or had low statistical power. The number of plots available for randomization was low for some habitats within which alpha niche overlap randomizations were constrained. In

addition, both alpha and beta niche overlap tests had low power among species or groups with few occurrences. An increase in the proportion of significant test results can be seen when moving from species, some of which occurred only once in the dataset, to higher taxonomic levels where species aggregation resulted in larger sample sizes (Fig. 3.3-3.6, Appendices 3.2-3.3).

In accordance with the alpha-beta niche conservatism model, I expected beta niche overlap tests to yield numerous cases of aggregation by habitat, and segregation by plot within habitats for alpha niche overlap tests. I was especially interested in finding examples of beta niche aggregation coupled with alpha niche segregation. Several species pairs and sister groups fit this coupled pattern, including *Echinocereus engelmannii*/*E. triglochidiatus*, *Opuntia basilaris*/*O. macrorhiza*, *Phlox austromontana*/*P. hoodii*, *Purshia mexicana*/*P. tridentata*, *Gutierrezia microcephala*/*G. sarothrae*, *Arabis*/*Draba*, and Boraginoideae/Hydrophylloideae. However, these cases were limited and none of them were simultaneously statistically significant for both beta niche aggregation and alpha niche segregation, whether measured using presence or cover.

On the other hand, many tests revealed patterns different from the expectations of the alpha-beta niche conservatism model. Some pairs were segregated by plot within habitats, suggesting alpha niche conservatism, but were also more segregated than expected among habitats, suggesting beta niche divergence. These may be examples of taxa that have a history of competition with one another, leading to the evolution of different habitat affinities that reduce competitive encounters—Connell’s (1980) “ghost of competition past”. Many of the taxon pairs that fit this pattern are dominants that may

indeed be in competition with one another, e.g. the trees *Pinus edulis/P. ponderosa*; shrubs *Quercus gambelii/Q. turbinella*, and *Amelanchier alnifolia/A. utahensis*; grasses *Poa fendleriana/P. pratensis*; and even higher taxa such as commelinids/misc. monocots, Ericales/[lamiids + campanulids] and Gymnosperms/Magnoliophyta. In many of these cases, alpha niche segregation was more significant when measured using cover, which would be expected to be a more sensitive measure of competitive effect than presence/absence.

The opposite pattern, aggregation by both habitat and plot, was also fairly common and significant. This pattern suggests beta niche conservatism combined with alpha niche divergence, i.e. related taxa with similar habitat affinities that appear to have evolved coexistence mechanisms. Taxa with riparian/wetland affinities are especially well represented in this group, including Cyperaceae/Juncaceae, *Populus/Salix* and various species pairs within *Juncus*. At the opposite moisture extreme, several members of the cactus genus *Opuntia* also fall in this category.

The most surprising and puzzling result was the widespread coupling of significant beta niche segregation with alpha niche aggregation. Many sister groups overlapped less often than expected in their habitat preferences, but when they did occur in the same habitat they tended to also occur in the same plot. This pattern suggests the ability to coexist due to alpha niche divergence, but why this would lead to increased co-occurrence despite different habitat preferences is difficult to explain. One possibility is that these groups are responding to environmental variation that was not captured by vegetation-based habitat characterizations. Groups exhibiting this pattern spanned a range of lineages and scales, e.g. *Elymus elymoides/E. lanceolatus*, *Acer*

grandidentatum/A. negundo, *Bromus diandrus/B. tectorum*, *Echinocereus/Opuntia*,
Anthemideae/Astereae, Cupressaceae/Pinaceae, and fabids/malvids.

CONCLUSIONS

Despite the limitations of the USGS-NPS plant community dataset, the exploratory null model approach used in this study proved useful for quantifying alpha and beta niche overlap across multiple lineages and phylogenetic scales at Zion. Predicted patterns of beta niche aggregation and alpha niche segregation were detected for some lineages and scales but were countered by numerous instances of other patterns suggesting niche divergence rather than conservatism. These results suggest that phylogenetic niche conservatism has left an imprint on plant community patterns at Zion, but only to a limited extent. Mirroring the criticisms of Giannani (2003) and Losos (2008), who questioned that assumption that phylogenetic relatedness and ecological similarity are consistently correlated, this study raises questions regarding the universality of the alpha-beta niche conservatism community assembly model.

Results of this study are best viewed as preliminary since they could be followed up with additional exploratory analyses as well as other research approaches. A larger vegetation plot sample is needed to fully characterize plant community patterns at Zion, including the many plant species known to occur in the park that were not represented in the USGS-NPS dataset (Cogan et al. 2004, Fertig and Alexander 2009). The discrete, vegetation-based habitat classes used in this study may not be the optimal way to characterize environmental variation at Zion; continuous-scale environmental models (e.g. regression tree models introduced in Chapter 2) offer an alternative. The spatial

scale of the current analysis may also be suboptimal for detecting phylogenetic community patterns. A smaller spatial grain (plot size) than 100-400 m² might be necessary to accurately capture the ‘neighborhood’ zone of alpha niche interactions, especially for species that are small in stature. A larger spatial extent (study area size), covering the entire region surrounding Zion or beyond, might reveal additional cases of beta niche conservatism if it captures allopatric relatives of species occurring at Zion (see Chapter 4).

Empirical analyses of community phylogenetic structure are but a starting point for understanding the influence of evolutionary history on community assembly. Phylogenetic aggregation and segregation patterns suggest but do not confirm phylogenetic niche conservatism. More explicit studies are needed to experimentally demonstrate the effects of competition and habitat filtering, and to identify specific, phylogenetically-conserved traits related to the alpha and beta niche. The study presented here highlights putative cases of phylogenetic niche conservatism that may be worth investigating in greater detail.

REFERENCES

- Albach, D. C., H. M. Meudt, and B. Oxelman. 2005. Piecing together the "new" Plantaginaceae. *American Journal of Botany* 92:297-315.
- APG [Angiosperm Phylogeny Group: B. Bremer, K. Bremer, M. W. Chase, M. F. Fay, J. L. Reveal, D. E. Soltis, P. S. Soltis, P. F. Stevens, A. A. Anderberg, M. J. Moore, R. G. Olmstead, P. J. Rudall, K. J. Sytsma, D. C. Tank, K. Wurdack, J. Q. Y. Xiang, and S. Zmarzty]. 2009. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG III. *Botanical Journal of the Linnean Society* 161:105-121.
- Baldwin, B. G., B. L. Wessa, and J. L. Panero. 2002. Nuclear rDNA evidence for major lineages of helenioid Heliantheae (Compositae). *Systematic Botany* 27:161-198.
- Barkworth, M. E., M. O. Arriaga, J. F. Smith, S. W. L. Jacobs, J. Valdes-Reyna, and B. S. Bushman. 2008. Molecules and morphology in South American Stipeae (Poaceae). *Systematic Botany* 33:719-731.
- Beck, J. B., G. L. Nesom, P. J. Calie, G. I. Baird, R. L. Small, and E. E. Schilling. 2004. Is subtribe Solidagininae (Asteraceae) monophyletic? *Taxon* 53:691-698.
- Beilstein, M. A., I. A. Al-Shehbaz, S. Mathews, and E. A. Kellogg. 2008. Brassicaceae phylogeny inferred from phytochrome A and ndhf sequence data: Tribes and trichomes revisited. *American Journal of Botany* 95:1307-1327.
- Cai, Y. F., S. W. Li, Y. Liu, S. Quan, M. Chen, Y. F. Xie, H. Z. Jiang, E. Z. Wei, N. W. Yin, L. Wang, R. Zhang, C. L. Huang, X. H. He, and M. F. Jiang. 2009. Molecular phylogeny of Ranunculaceae based on internal transcribed spacer sequences. *African Journal of Biotechnology* 8:5215-5224.
- Cavender-Bares, J., D. D. Ackerly, D. A. Baum, and F. A. Bazzaz. 2004. Phylogenetic overdispersion in Floridian oak communities. *American Naturalist* 163:823-843.
- Cavender-Bares, J., A. Keen, and B. Miles. 2006. Phylogenetic structure of Floridian plant communities depends on taxonomic and spatial scale. *Ecology* 87:S109-S122.
- Cavender-Bares, J., K. H. Kozak, P. V. A. Fine, and S. W. Kembel. 2009. The merging of community ecology and phylogenetic biology. *Ecology Letters* 12:693-715.
- Cogan, D., M. Reid, K. Schulz, and M. Pucherelli. 2004. Zion National Park, Utah: 1999-2003 Vegetation Mapping Program, Final Report—March 31, 2004. Report Technical Memorandum 8260-03-01, Remote Sensing and GIS Group, Technical Service Center, Bureau of Reclamation, Denver, CO.
<http://biology.usgs.gov/npsveg/zion/zionrpt.pdf>.

- Connell, J. H. 1980. Diversity and the coevolution of competitors, or the ghost of competition past. *Oikos* 35:131-138.
- Daubenmire, R. 1959. A canopy-coverage method of vegetational analysis. *Northwest Science* 23:69-82.
- Davis, J. I., D. W. Stevenson, G. Petersen, O. Seberg, L. M. Campbell, J. V. Freudenstein, D. H. Goldman, C. R. Hardy, F. A. Michelangeli, M. P. Simmons, C. D. Specht, F. Vergara-Silva, and M. Gandolfo. 2004. A Phylogeny of the monocots, as inferred from rbcL and atpA sequence variation, and a comparison of methods for calculating jackknife and bootstrap values. *Systematic Botany* 29:467-510.
- Douglas, N. A., and P. S. Manos. 2007. Molecular phylogeny of Nyctaginaceae: Taxonomy, biogeography, and characters associated with a radiation of xerophytic genera in North America. *American Journal of Botany* 94:856-872.
- Duvall, M. R., J. D. Noll, and A. H. Minn. 2001. Phylogenetics of Paniceae (Poaceae). *American Journal of Botany* 88:1988-1992.
- Duvall, M. R., P. M. Peterson, and A. H. Christensen. 1994. Alliances of *Muhlenbergia* (Poaceae) within new-world Eragrostideae are identified by phylogenetic analysis of mapped restriction sites from plastid DNAs. *American Journal of Botany* 81:622-629.
- Emerson, B. C., and R. G. Gillespie. 2008. Phylogenetic analysis of community assembly and structure over space and time. *Trends in Ecology and Evolution* 23:619-630.
- Fertig, W., and J. Alexander. 2009. Annotated Checklist of Vascular Flora: Zion National Park. Natural Resource Technical Report NPS/NCPN/NRTR-2009/157. USDI National Park Service, Natural Resource Program Center, Fort Collins, CO.
- Flores, H., and J. I. Davis. 2001. A cladistic analysis of Atripliceae (Chenopodiaceae) based on morphological data. *Journal of the Torrey Botanical Society* 128:297-319.
- Funk, V. A., R. J. Bayer, S. Keeley, R. Chan, L. Watson, B. Gemeinholzer, E. Schilling, J. L. Panero, B. G. Baldwin, N. Garcia-Jacas, A. Susanna, and R. K. Jansen. 2005. Everywhere but Antarctica: Using a supertree to understand the diversity and distribution of the Compositae. Pages 343-374 in I. Friis and H. Balslev, editors. *Plant Diversity and Complexity Patterns—Local, Regional and Global Dimensions*. Biol. Skr. 55.
- German, D. A., N. Friesen, B. Neuffer, I. A. Al-Shehbaz, and H. Hurka. 2009. Contribution to ITS phylogeny of the Brassicaceae, with special reference to some Asian taxa. *Plant Systematics and Evolution* 283:33-56.

- Giannini, N. P. 2003. Canonical phylogenetic ordination. *Systematic Biology* 52:684-695.
- Gillespie, L. J., R. J. Soreng, R. D. Bull, S. W. L. Jacobs, and N. F. Refulio-Rodriguez. 2008. Phylogenetic relationships in subtribe Poinae (Poaceae, Poeae) based on nuclear ITS and plastid trnT-trnL-trnF sequences. *Botany-Botanique* 86:938-967.
- Gillison, A. N., and K. R. W. Brewer. 1985. The use of gradient directed transects or gradsects in natural-resource surveys. *Journal of Environmental Management* 20:103-127.
- Gotelli, N. J., and G. L. Entsminger. 2009. EcoSim: Null Models Software for Ecology. Acquired Intelligence Inc. & Kesey-Bear, Jericho, VT.
<http://www.garyentsminger.com/ecosim/index.htm>.
- GPWG [Grass Phylogeny Working Group: Barker, N. P., L. G. Clark, J. I. Davis, M. R. Duvall, G. F. Guala, C. Hsiao, E. A. Kellogg, H. P. Linder, R. J. Mason-Gamer, S. Y. Mathews, M. P. Simmons, R. J. Soreng, and R. E. Spangler]. 2001. Phylogeny and subfamilial classification of the grasses (Poaceae). *Annals of the Missouri Botanical Garden* 88:373-457.
- Grossman, D. H., D. Faber-Langendoen, A. S. Weakley, M. Anderson, P. Bourgeron, R. Crawford, K. Goodin, S. Landaal, K. Metzler, K. Patterson, M. Pyne, M. Reid, and L. Sneddon. 1998. International Classification of Ecological Communities: Terrestrial Vegetation of the United States, Volume 1: The National Vegetation Classification System: Development, Status, and Applications. The Nature Conservancy, Arlington, VA.
- Harper, K. T. 1993. Zion National Park Vegetation: Summary Report. Zion National Park, Springdale, Utah.
- Harvey, P. H., and M. D. Pagel. 1991. The Comparative Method in Evolutionary Biology. Oxford University Press, Oxford; New York.
- Hilu, K. W., and L. A. Alice. 2001. A phylogeny of Chloridoideae (Poaceae) based on matK sequences. *Systematic Botany* 26:386-405.
- Jennings, M. D., D. Faber-Langendoen, O. L. Loucks, R. K. Peet, and D. Roberts. 2009. Standards for associations and alliances of the US National Vegetation Classification. *Ecological Monographs* 79:173-199.
- Johnson, L. A., L. M. Chan, T. L. Weese, L. D. Busby, and S. McMurry. 2008. Nuclear and cpDNA sequences combined provide strong inference of higher phylogenetic relationships in the phlox family (Polemoniaceae). *Molecular Phylogenetics and Evolution* 48:997-1012.

- Kartesz, J. T. 1999. A Synonymized Checklist and Atlas with Biological Attributes for the Vascular Flora of the United States, Canada, and Greenland, First Edition. In J. T. Kartesz and C. A. Meacham, editors. Snythesis of the North American Flora, Version 1.0. North Carolina Botanical Garden, Chapel Hill, NC.
- Kilian, N., B. Gemeinholzer, and H. W. Lack. 2009. Tribe Cichorieae. in V. A. Funk, A. Susanna, T. Stuessy, and R. Bayer, editors. Systematics, Evolution, and Biogeography of the Compositae. IAPT, Vienna.
- Krebs, C. J. 1999. Ecological Methodology, 2nd edition. Benjamin Cummings, Menlo Park, CA.
- Langstrom, E., and M. W. Chase. 2002. Tribes of Boraginoideae (Boraginaceae) and placement of *Antiphytum*, *Echiochilon*, *Ogastemma* and *Sericostoma*: A phylogenetic analysis based on atpB plastid DNA sequence data. Plant Systematics and Evolution 234:137-153.
- Lee, J., B. G. Baldwin, and L. D. Gottlieb. 2003. Phylogenetic relationships among the primarily North American genera of Cichorieae (Compositae) based on analysis of 18S-26S nuclear rDNA ITS and ETS sequences. Systematic Botany 28:616-626.
- Linder, C. R., and L. H. Rieseberg. 2004. Reconstructing patterns of reticulate evolution in plants. American Journal of Botany 91:1700-1708.
- Losos, J. B. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. Ecology Letters 11:995-1003.
- Martins, T. R., and T. J. Barkman. 2005. Reconstruction of Solanaceae phylogeny using the nuclear gene SAMT. Systematic Botany 30:435-447.
- Mason-Gamer, R. J. 2005. The beta-amylase genes of grasses and a phylogenetic analysis of the Triticeae (Poaceae). American Journal of Botany 92:1045-1058.
- McCune, B., and M. J. Mefford. 1997. PC-ORD. Multivariate Analysis of Ecological Data. MjM Software Design, Gleneden Beach, OR.
- Meyer, S. E. 1974. Floristic investigations at the northeastern boundary of the Mojave Desert. Journal of the Arizona Academy of Science 9 Suppl.
- Moon, H. K., S. P. Hong, E. Smets, and S. Huysmans. 2009. Phylogenetic significance of leaf micromorphology and anatomy in the tribe Mentheae (Nepetoideae: Lamiaceae). Botanical Journal of the Linnean Society 160:211-231.

- Moore, A. J., and L. Bohs. 2007. Phylogeny of *Balsamorhiza* and *Wyethia* (Asteraceae : Heliantheae) using ITS, ETS, and trnK sequence data. Systematic Botany 32:682-691.
- Muasya, A. M., D. A. Simpson, G. A. Verboom, P. Goetghebeur, R. F. C. Naczi, M. W. Chase, and E. Smets. 2009. Phylogeny of Cyperaceae based on DNA sequence data: Current progress and future prospects. Botanical Review 75:2-21.
- NCBI. 2008. The National Center for Biotechnology Information.
<http://www.ncbi.nlm.nih.gov/>.
- Noyes, R. D., and L. H. Rieseberg. 1999. ITS sequence data support a single origin for North American Astereae (Asteraceae) and reflect deep geographic divisions in *Aster* s.l. American Journal of Botany 86:398-412.
- Panero, J. L., and V. A. Funk. 2002. Toward a phylogenetic subfamilial classification for the Compositae (Asteraceae). Proceedings of the Biological Society of Washington 115:909-922.
- Panero, J. L., and V. A. Funk. 2008. The value of sampling anomalous taxa in phylogenetic studies: Major clades of the Asteraceae revealed. Molecular Phylogenetics and Evolution 47:757-782.
- Pickett, S. T. A., and F. A. Bazzaz. 1978. Organization of an assemblage of early successional species on a soil-moisture gradient. Ecology 59:1248-1255.
- Potter, D., T. Eriksson, R. C. Evans, S. Oh, J. E. E. Smedmark, D. R. Morgan, M. Kerr, K. R. Robertson, M. Arsenault, T. A. Dickinson, and C. S. Campbell. 2007. Phylogeny and classification of Rosaceae. Plant Systematics and Evolution 266:5-43.
- Quintanar, A., S. Castroviejo, and P. Catalan. 2007. Phylogeny of the tribe Aveneae (Pooideae, Poaceae) inferred from plastid trnT-F and nuclear ITS sequences. American Journal of Botany 94:1554-1569.
- R Development Core Team. 2009. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria.
<http://www.r-project.org/>.
- Ro, K. E., C. S. Keener, and B. A. McPheron. 1997. Molecular phylogenetic study of the Ranunculaceae: Utility of the nuclear 26S ribosomal DNA in inferring intrafamilial relationships. Molecular Phylogenetics and Evolution 8:117-127.
- Roberts, R. P., and L. E. Urbatsch. 2003. Molecular phylogeny of *Ericameria* (Asteraceae, Astereae) based on nuclear ribosomal 3' ETS and ITS sequence data. Taxon 52:209-228.

- Roberts, R. P., and L. E. Urbatsch. 2004. Molecular phylogeny of *Chrysanthemum* and related genera (Asteraceae, Astereae) based on nuclear ribosomal 3' ETS and ITS sequence data. *Systematic Botany* 29:199-215.
- Sanchez, A., T. M. Schuster, and K. A. Kron. 2009. A large-scale phylogeny of Polygonaceae based on molecular data. *International Journal of Plant Sciences* 170:1044-1055.
- Schilling, E. E., and J. L. Panero. 2002. A revised classification of subtribe Helianthinae (Asteraceae : Hetiantheae). I. Basal lineages. *Botanical Journal of the Linnean Society* 140:65-76.
- Schneider, J., E. Doring, K. W. Hilu, and M. Roser. 2009. Phylogenetic structure of the grass subfamily Pooideae based on comparison of plastid matK gene-3' trnK exon and nuclear ITS sequences. *Taxon* 58:405-424.
- Schuettelpelz, E., and K. M. Pryer. 2007. Fern phylogeny inferred from 400 leptosporangiate species and three plastid genes. *Taxon* 56:1037-1050.
- Selliah, S., and L. Brouillet. 2008. Molecular phylogeny of the North American euryboid asters (Asteraceae, Astereae) based on the nuclear ribosomal internal and external transcribed spacers. *Botany-Botanique* 86:901-915.
- Silvertown, J. 2004. Plant coexistence and the niche. *Trends in Ecology & Evolution* 19:605-611.
- Silvertown, J., M. Dodd, D. Gowing, C. Lawson, and K. Mcconway. 2006. Phylogeny and the hierarchical organization of plant diversity. *Ecology* 87:S39-S49.
- Slingsby, J. A., and G. A. Verboom. 2006. Phylogenetic relatedness limits co-occurrence at fine spatial scales: Evidence from the schoenoid sedges (Cyperaceae: Schoeneae) of the Cape Floristic Region, South Africa. *American Naturalist* 168:14-27.
- Smith, A. R., K. M. Pryer, E. Schuettelpelz, P. Korall, H. Schneider, and P. G. Wolf. 2006. A classification for extant ferns. *Taxon* 55:705-731.
- Spangler, R., B. Zaitchik, E. Russo, and E. Kellogg. 1999. Andropogoneae evolution and generic limits in *Sorghum* (Poaceae) using ndhF sequences. *Systematic Botany* 24:267-281.
- Stebbins, G. L. 1969. The significance of hybridization for plant taxonomy and evolution. *Taxon* 18:26-35.
- Stevens, P. F. 2008. Angiosperm Phylogeny Website, Version 9. <http://www.mobot.org/MOBOT/research/APweb/>.

- Swenson, N. G., B. J. Enquist, J. Pither, J. Thompson, and J. K. Zimmerman. 2006. The problem and promise of scale dependency in community phylogenetics. *Ecology* 87:2418-2424.
- Swenson, N. G., B. J. Enquist, J. Thompson, and J. K. Zimmerman. 2007. The influence of spatial and size scale on phylogenetic relatedness in tropical forest communities. *Ecology* 88:1770-1780.
- Tank, D. C., P. M. Beardsley, S. A. Kelchner, and R. G. Olmstead. 2006. Review of the systematics of Scrophulariaceae s.l. and their current disposition. *Australian Systematic Botany* 19:289-307.
- Tukey, J. W. 1980. We need both exploratory and confirmatory. *American Statistician* 34:23-25.
- Urbatsch, L. E., B. G. Baldwin, and M. J. Donoghue. 2000. Phylogeny of the coneflowers and relatives (Heliantheae: Asteraceae) based on nuclear rDNA internal transcribed spacer (ITS) sequences and chloroplast DNA restriction site data. *Systematic Botany* 25:539-565.
- Urbatsch, L. E., R. P. Roberts, and V. Karaman. 2003. Phylogenetic evaluation of *Xylothamia*, *Gundlachia*, and related genera (Asteraceae, Astereae) based on ETS and ITS nrDNA sequence data. *American Journal of Botany* 90:634-649.
- USGS (U.S. Geologic Survey). 2004. USGS Vegetation Characterization: Zion National Park. <http://biology.usgs.gov/npsveg/zion/index.html>.
- Valles, J., M. Torrell, T. Garnatje, N. Garcia-Jacas, R. Vilatersana, and A. Susanna. 2003. The genus *Artemisia* and its allies: Phylogeny of the subtribe Artemisiinae (Asteraceae, Anthemideae) based on nucleotide sequences of nuclear ribosomal DNA internal transcribed spacers (ITS). *Plant Biology* 5:274-284.
- Vamosi, S. M., S. B. Heard, J. C. Vamosi, and C. O. Webb. 2009. Emerging patterns in the comparative analysis of phylogenetic community structure. *Molecular Ecology* 18:572-592.
- Walker, J. B., and K. J. Sytsma. 2007. Staminal evolution in the genus *Salvia* (Lamiaceae): Molecular phylogenetic evidence for multiple origins of the staminal lever. *Annals of Botany* 100:375-391.
- Wang, W., A. M. Lu, Y. Ren, M. E. Endress, and Z. D. Chen. 2009. Phylogeny and classification of Ranunculales: Evidence from four molecular loci and morphological data. *Perspectives in Plant Ecology Evolution and Systematics* 11:81-110.

- Wang, X. Q., D. C. Tank, and T. Sang. 2000. Phylogeny and divergence times in Pinaceae: Evidence from three genomes. *Molecular Biology and Evolution* 17:773-781.
- Warren, D. L., R. E. Glor, and M. Turelli. 2008. Environmental niche equivalency versus conservatism: Quantitative approaches to niche evolution. *Evolution* 62:2868-2883.
- Webb, C. O., D. D. Ackerly, M. A. Mcpeak, and M. J. Donoghue. 2002. Phylogenies and community ecology. *Annual Review of Ecology and Systematics* 33:475-505.
- Webb, C. O., J. B. Losos, and A. A. Agrawal. 2006. Integrating phylogenies into community ecology. *Ecology* 87:S1-S2.
- Welsh, S. L. 1989. Hanging Gardens of Zion National Park: Final Report. Endangered Plant Studies, Inc., Orem, UT.
- Welsh, S. L., N. D. Atwood, S. Goodrich, and L. C. Higgins. 2003. A Utah Flora, 3rd Edition. Brigham Young University, Provo, UT.
- Whitney, K. D., J. R. Ahern, L. G. Cambell, L. P. Albert, and M. S. King. 2010. Patterns of hybridization in plants. *Perspectives in Plant Ecology, Evolution and Systematics* (article in press, accepted 16 Feb. 2010).
- Whittaker, R. H. 1975. Communities and Ecosystems. Macmillan, New York.
- Wilson, J. B. 2007. Priorities in statistics, the sensitive feet of elephants, and don't transform data. *Folia Geobotanica* 42:161-167.
- Wojciechowski, M. F., M. Lavin, and M. J. Sanderson. 2004. A phylogeny of legumes (Leguminosae) based on analyses of the plastid matK gene resolves many well-supported subclades within the family. *American Journal of Botany* 91:1846-1862.
- Woodbury, A. M. 1933. Biotic relationships of Zion Canyon, Utah with special reference to succession. *Ecological Monographs* 3:151-245.

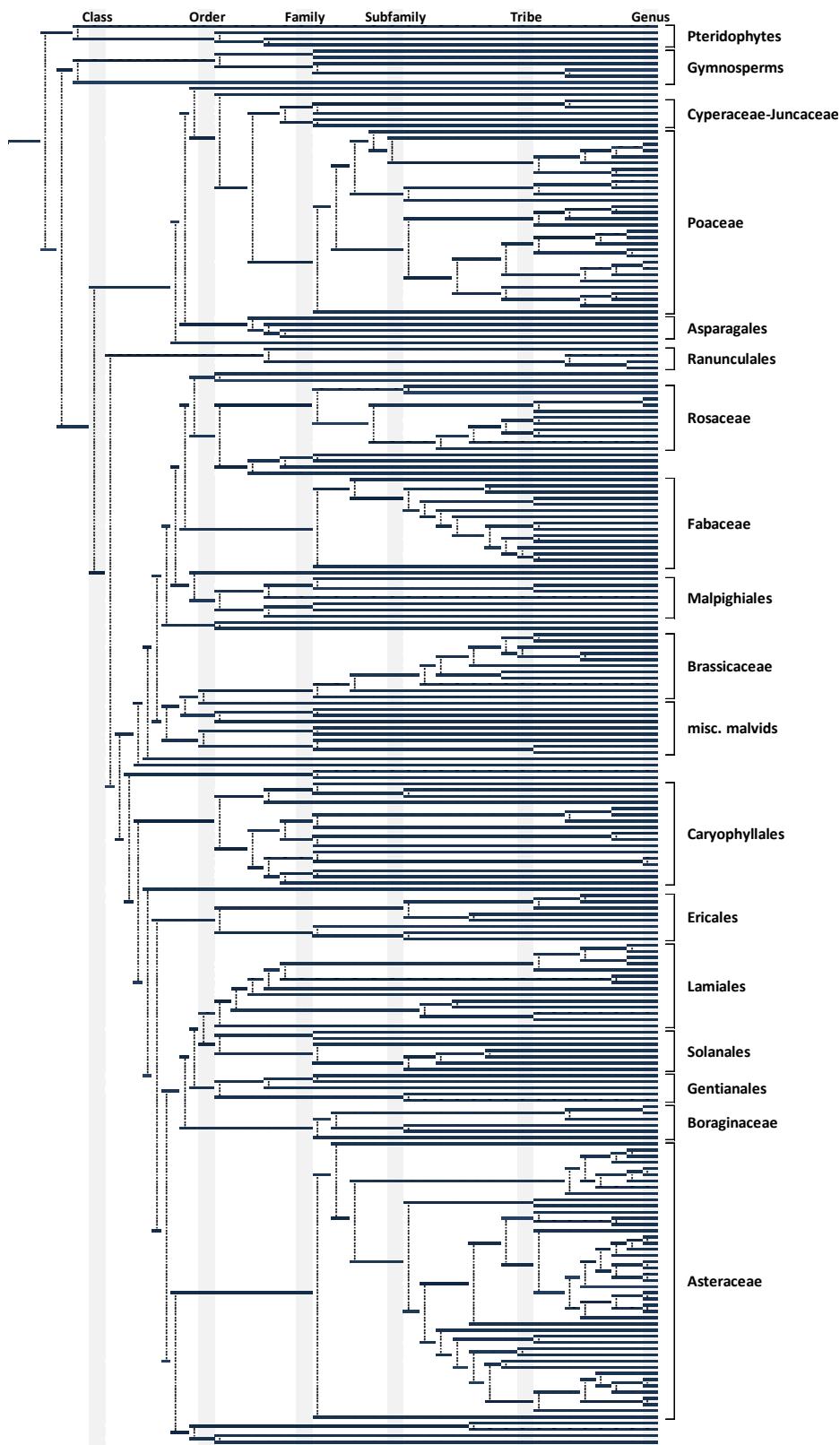


Figure 3.1. Genus-level phylogenetic supertree of vascular plants reported in the USGS-NPS vegetation survey of Zion National Park (Cogan et al. 2004).

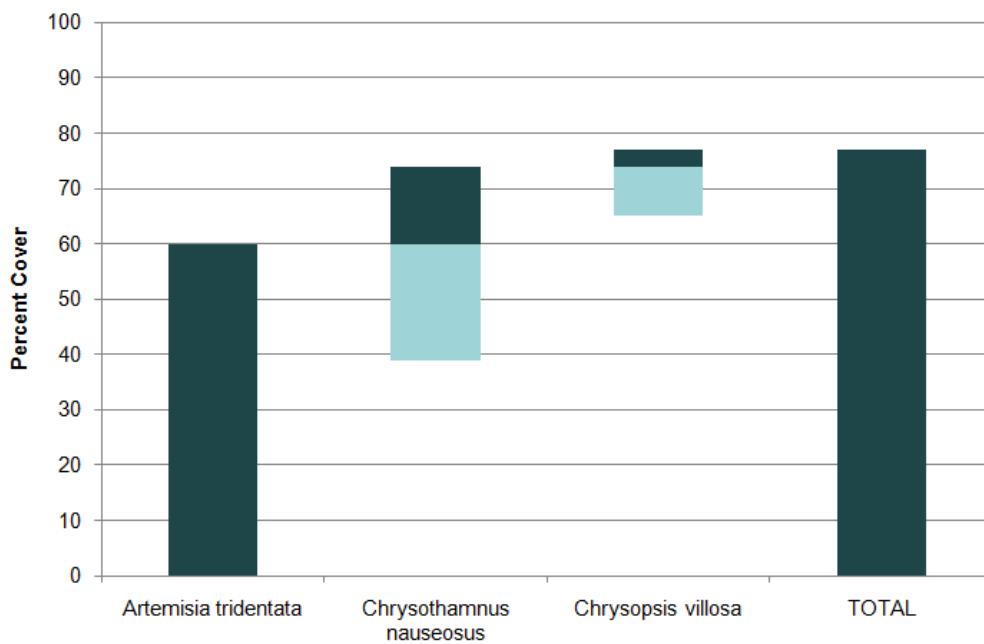


Figure 3.2. Hypothetical scenario illustrating procedure for aggregating cover values of multiple species (or strata) within plots. Three species are arranged in descending order of cover (*Artemisia tridentata*, 60%; *Chrysothamnus nauseosus*, 35%; *Chrysopsis villosa*, 12%). Building on total cover of the first species, cover of secondary species is added as a proportion (dark) of their total cover (dark + light blue) equivalent to the proportion of the plot not yet covered.

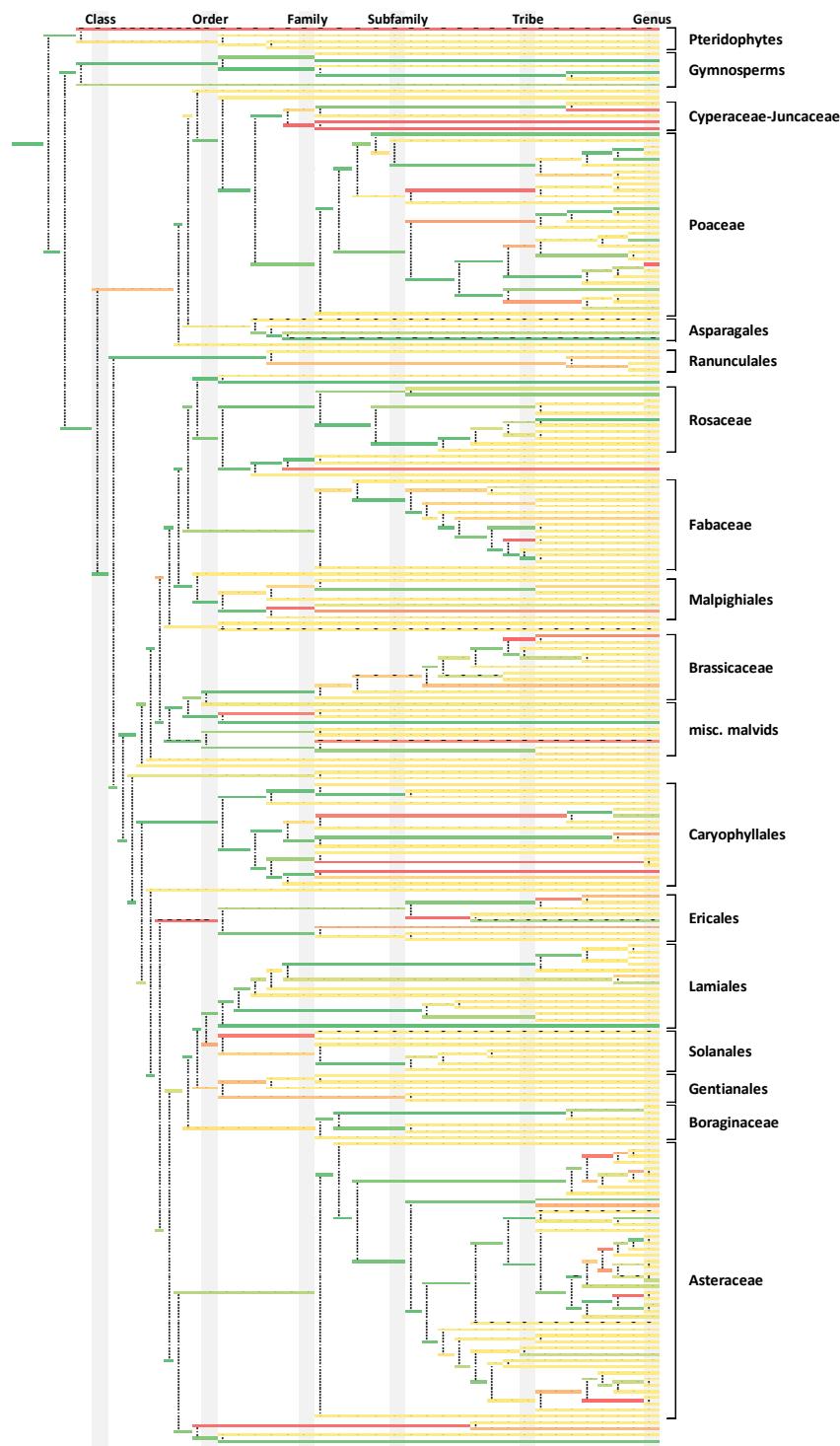


Figure 3.3. Results of **beta niche** overlap tests based on **presence/absence**, mapped onto a genus-level phylogeny of vascular plants of Zion National Park. Sister groups are subtended by colored branches ranging from green (higher segregation than expected) through red (higher aggregation than expected). Terminal branches are null (yellow) if they contain only one species, otherwise they are averages for congeneric species pairs (Appendix 3.2). See also Appendix 3.3.

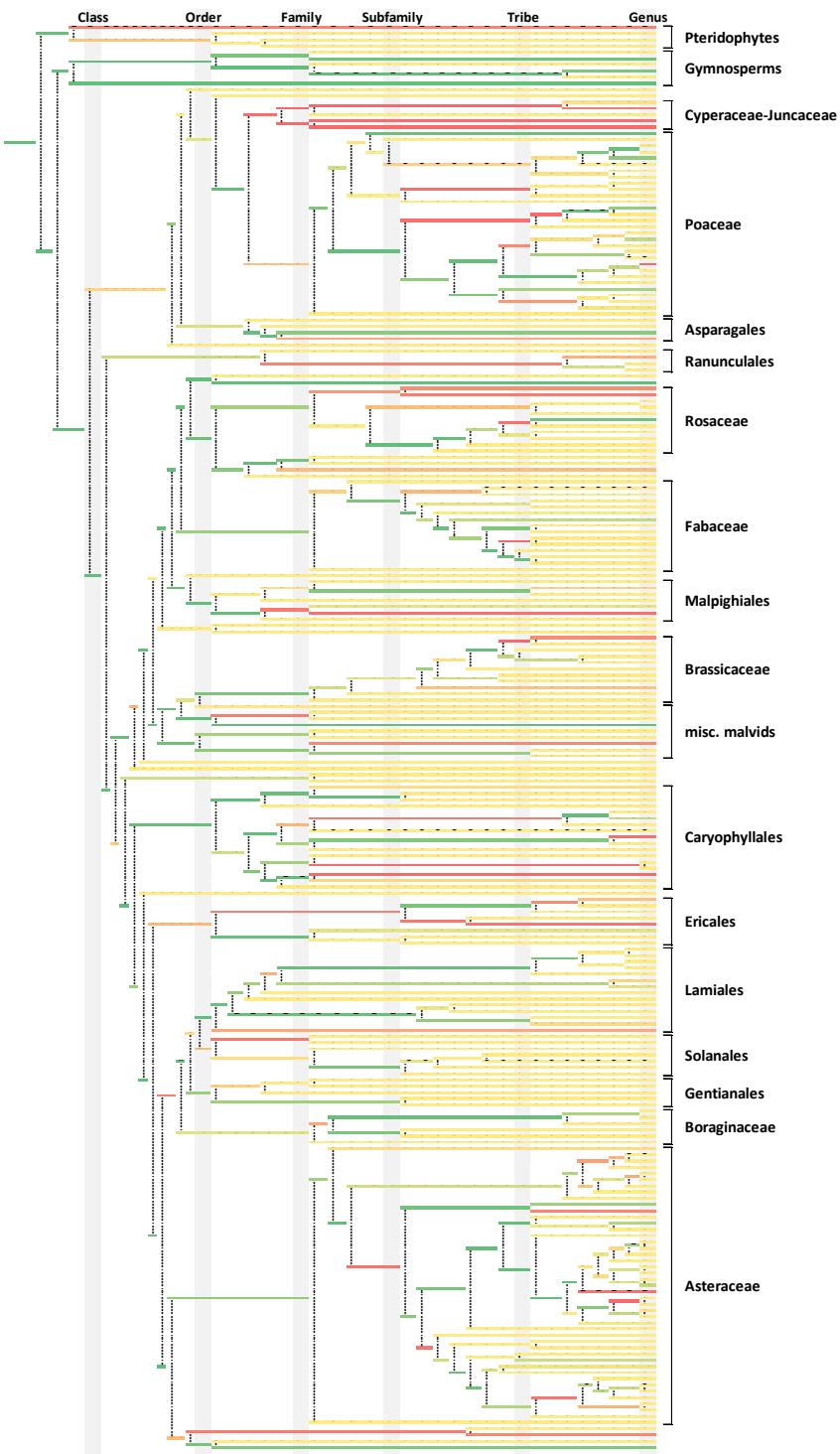


Figure 3.4. Results of **beta niche** overlap tests based on **cover**, mapped onto a genus-level phylogeny of vascular plants of Zion National Park. Sister groups are subtended by colored branches ranging from green (higher segregation than expected) through red (higher aggregation than expected). Terminal branches are null (yellow) if they contain only one species, otherwise they are averages for congeneric species pairs (Appendix 3.2). See also Appendix 3.3.

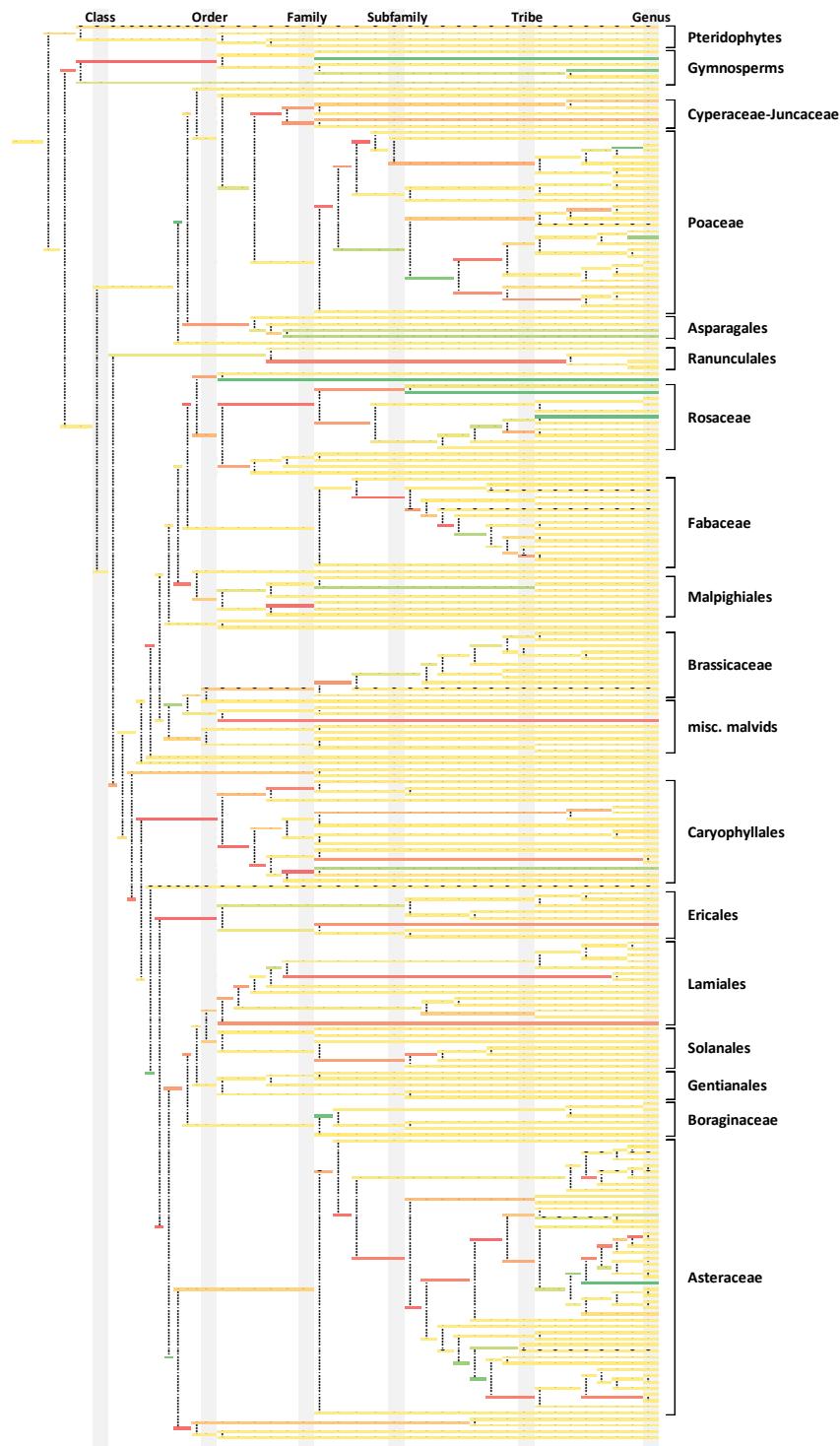


Figure 3.5. Results of **alpha niche** overlap tests based on **presence/absence**, mapped onto a genus-level phylogeny of vascular plants of Zion National Park. Sister groups are subtended by colored branches ranging from green (higher segregation than expected) through red (higher aggregation than expected). Terminal branches are null (yellow) if they contain only one species, otherwise they are averages for congeneric species pairs (Appendix 3.2). See also Appendix 3.3.

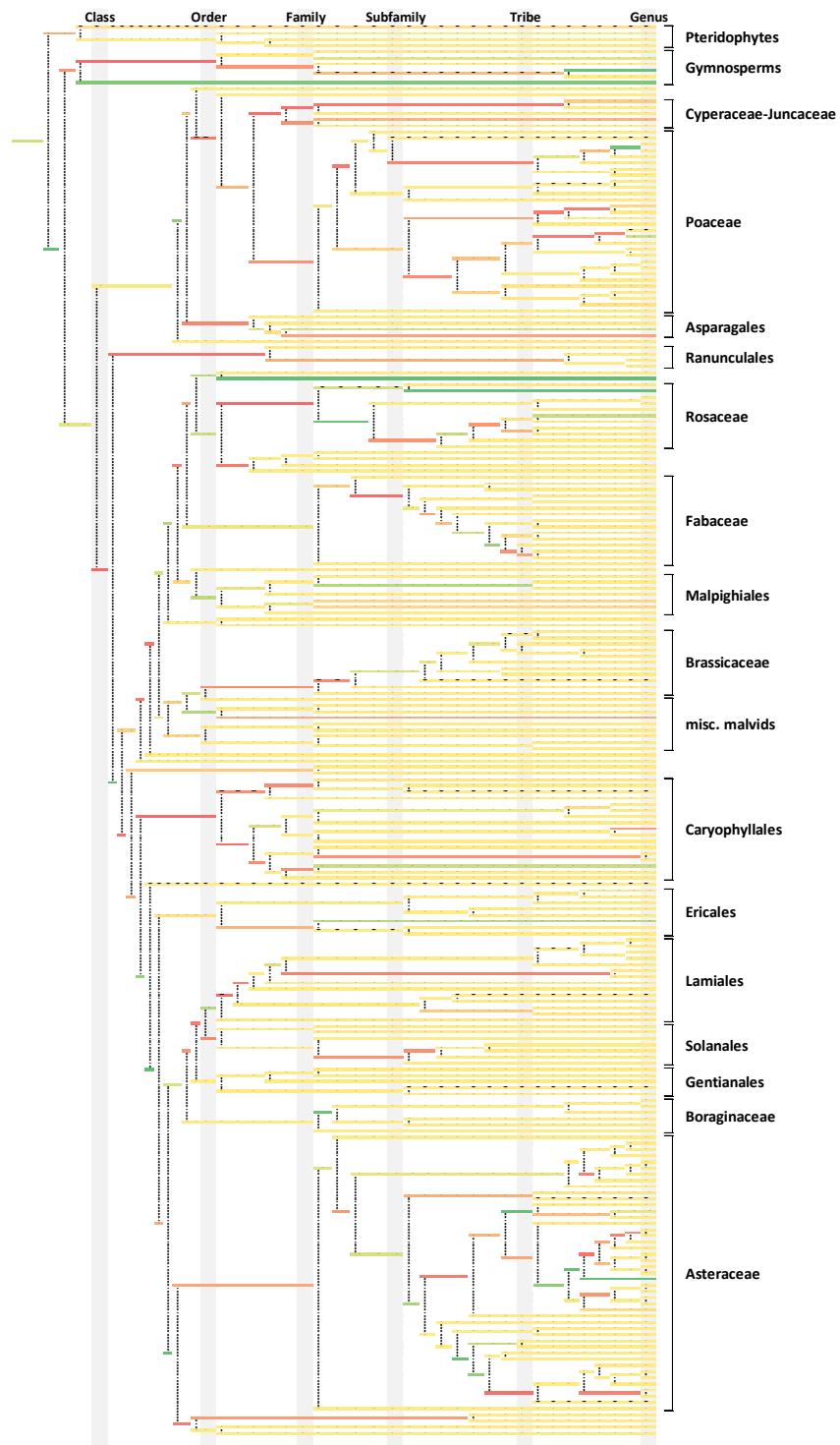


Figure 3.6. Results of **alpha niche** overlap tests based on **cover**, mapped onto a genus-level phylogeny of vascular plants of Zion National Park. Sister groups are subtended by colored branches ranging from green (higher segregation than expected) through red (higher aggregation than expected). Terminal branches are null (yellow) if they contain only one species, otherwise they are averages for congeneric species pairs (Appendix 3.2). See also Appendix 3.3.

Table 3.1. Ecological divisions used to group alliances of the U.S. National Vegetation Classification at Zion National Park.

Ecological Division	Alliances
Coniferous Forests	Douglas-fir Forest, White Fir Forest
Desert Shrublands	Blackbrush Shrubland, Fourwing Saltbush Shrubland, Nevada Joint-fir Shrubland, Painted Desert Sparsely Vegetated, Snakeweed Dwarf-shrubland
High Elevation Deciduous Forests	Bigtooth Maple Montane Forest, Quaking Aspen Forest, Quaking Aspen Temporarily Flooded Forest, White Fir - Quaking Aspen Forest
Higher Elevation Shrublands	(Stansbury Cliff-rose, Mexican Cliff-rose) Shrubland, Black Sagebrush Dwarf-shrubland, Gambel Oak Shrubland, Greenleaf Manzanita Shrubland, Mexican Manzanita Shrubland, Mountain-mahogany Sparsely Vegetated, Utah Serviceberry Shrubland
Lower Elevation Shrublands	Basin Big Sagebrush Shrubland, Big Sagebrush Shrubland, Rubber Rabbitbrush Sand Deposit Sparse Vegetation [provisional], Green Rabbitbrush Shrub Herbaceous, Mormon-tea Shrubland, Mountain Big Sagebrush Shrubland, Rubber Rabbitbrush Shrubland, Sand Sagebrush Shrubland, Turbinella Live Oak Shrubland
Pinyon/Juniper Woodlands	Rocky Mountain Juniper Woodland, Singleleaf Pinyon - (Utah Juniper) Woodland, Two-needle Pinyon - (Juniper species) Woodland, Utah Juniper Woodland
Ponderosa Pine Woodlands	Ponderosa Pine Woodland
Riparian/Wetland Vegetation	(Coyote Willow, Sandbar Willow) Temporarily Flooded Shrubland, (Field Horsetail, Variegated Scouringrush) Semipermanently Flooded Herbaceous, (Swollen-beak Sedge, Beaked Sedge) Seasonally Flooded Herbaceous, Arrow-weed Seasonally Flooded Shrubland, Emory Seepwillow Shrubland [provisional], Baltic Rush Seasonally Flooded Herbaceous, Box-elder Temporarily Flooded Woodland, Fremont Cottonwood Temporarily Flooded Forest, Fremont Cottonwood Temporarily Flooded Woodland, Nebraska Sedge Seasonally Flooded Herbaceous, Single-leaf Ash Temporarily Flooded Woodland, Strapleaf Willow Temporarily Flooded Shrubland, Water Birch Temporarily Flooded Shrubland
Slickrock Sparse Vegetation	Littleleaf Mountain-mahogany Sparsely Vegetated, Wooded Bedrock Sparsely Vegetated
Upland Grasslands	Blue Grama Herbaceous, Intermediate Wheatgrass Semi-natural Herbaceous, James' Galleta Herbaceous, Kentucky Bluegrass Semi-natural Seasonally Flooded Herbaceous, Mountain Muhly Herbaceous, Mountain Snowberry Shrubland, Needle-and-Thread Bunch Herbaceous, Sand Dropseed Herbaceous, Smooth Brome Semi-natural Herbaceous

Table 3.2. References used to resolve phylogenetic relationships within families for the genus-level vascular plant supertree used in the current study.

Family	References
Amaranthaceae	Flores and Davis 2001
Asteraceae	Miao et al. 1995, Noyes and Rieseberg 1999, Urbatsch et al. 2000, Baldwin et al. 2002, Panero and Funk 2002, Schilling and Panero 2002, Lee et al. 2003, Roberts and Urbatsch 2003, Urbatsch et al. 2003, Valles et al. 2003, Beck et al. 2004, Roberts and Urbatsch 2004, Funk et al. 2005, Moore and Bohs 2007, Selliah and Brouillet 2008, Morgan et al. 2009, Panero and Funk 2008, Kilian et al. 2009
Boraginaceae	Langstrom and Chase 2002, Stevens 2008
Brassicaceae	Hauser and Crovello 1982, Beilstein et al. 2008, German et al. 2009
Cyperaceae	Muasya et al. 2009
Ericaceae	Stevens 2008
Fabaceae	Wojciechowski et al. 2004
Lamiaceae	Walker and Sytsma 2007, Moon et al. 2009
Nyctaginaceae	Douglas and Manos 2007
Pinaceae	Wang et al. 2000
Plantaginaceae	Albach et al. 2005
Poaceae	Duvall et al. 1994, Spangler et al. 1999, GPWG 2001, Duvall et al. 2001, Hilu and Alice 2001, Mason-Gamer 2005, Quintanar et al. 2007, Barkworth et al. 2008, Gillespie et al. 2008, Schneider et al. 2009
Polemoniaceae	Johnson et al. 2008
Polygonaceae	Sanchez et al. 2009
Ranunculaceae	Ro et al. 1997, Cai et al. 2009, Wang et al. 2009
Rosaceae	Potter et al. 2007
Solanaceae	Martins and Barkman 2005

Table 3.3. Summary of niche overlap tests for congeneric pairs and phylogenetic sister groups (genus and above) of vascular plant species at Zion National Park. Numbers of cases of aggregation (higher overlap than expected), segregation (lower overlap than expected) and null (neither higher nor lower overlap than null models) are shown, along with numbers of significant cases ($p < 0.05$ for two-tailed test) in parentheses. See Appendices 3.2-3.3 for full detail.

A. Congeneric Pairs		Aggregation	Segregation	Null
Beta Niche Overlap	Presence	123 (42)	151 (30)	67
	Cover	141 (34)	129 (24)	71
Alpha Niche Overlap	Presence	44 (7)	41 (8)	256
	Cover	45 (9)	38 (7)	258

B. Sister Groups		Aggregation	Segregation	Null
Beta Niche Overlap	Presence	52 (7)	164 (67)	15
	Cover	64 (10)	152 (62)	15
Alpha Niche Overlap	Presence	94 (14)	42 (1)	96
	Cover	96 (9)	56 (6)	79

CHAPTER 4

SHARPENING THE FOCUS OF COMMUNITY PATTERNS BY ADJUSTING PHYLOGENETIC SCALE

ABSTRACT

Scale is an important issue in community ecology because community patterns differ according to the scale of observation. Spatial scale determines which taxa are captured and which spatial phenomenon are highlighted in community analysis. If community-environment relationships are the phenomenon of interest, the influence of ecological interactions at a small spatial grain (i.e., small sample unit size) or species dispersal limitation at a large spatial extent (i.e., large sampled area size) could be considered noise. Here I consider the possibility that these types of noise can be reduced by adjusting the phylogenetic scale of analysis. Phylogenetic scale can be defined in a manner similar to spatial scale, defining extent as the phylogenetic scope of a community study (e.g. vascular plants) and grain as the phylogenetic resolution of community units (e.g. species). Focus, another spatial scale concept referring to sample unit averaging, can also be applied to taxa in the form of phylogenetic lumping. Lumping of phylogenetically-related taxa may be appropriate if the taxa exhibit phylogenetic niche conservatism and thus respond to environmental gradients in similar ways, even though they may be spatially segregated due to ecological interactions or historical factors. Under these conditions the effectively coarser phylogenetic grain of lumped taxa could

sharpen community-environment patterns under the constraints of a given spatial grain and extent. This raises the question of how to identify such an “optimal” phylogenetic grain, given that patterns of niche conservatism may differ in different lineages and on different environmental gradients. I suggest a solution that entails exploratory analysis of association patterns among taxa at multiple levels of a phylogenetic hierarchy, leading to identification of clusters of strongly-associated taxa of potentially variable phylogenetic resolution. This approach follows the logic of indirect gradient analysis by using community patterns to infer environmental influences while also reducing potential noise arising from the use of a fixed community taxonomic level. I present an application of this approach using data from the Carolina Vegetation Survey.

INTRODUCTION

For many years, community ecologists have sought to characterize communities based on patterns of selected taxa within defined study areas. Taxonomically-based, spatially-defined community characterization is exemplified by studies employing classification or ordination techniques (McCune and Grace 2002) to elucidate patterns of spatial variation in species composition of a targeted taxonomic group in a study area of interest. For example, vascular plants have been a commonly-targeted taxonomic group for studies in which the presence or abundance of species in sample plots has been used to classify or ordinate plant communities of local to regional study areas (Mueller-Dombois and Ellenberg 1974, Kent and Coker 1992, Jennings et al. 2009).

An implicit assumption of such studies is that spatial variation in species composition (presence and/or abundance) is due, at least in part, to environmental factors

that also vary spatially within the study area. Identifying the environmental factors responsible for community patterns is a common objective that is frequently pursued using correlative approaches. Correlative relationships between community patterns and environmental variables may be difficult to detect because the relevant environmental factors are difficult to measure, or because chance events contribute to noisy patterns (McCune and Grace 2002). Another possible reason for weak community-environment relationships, which forms the topic of this paper, is that community patterns are often described from an inappropriate scale. In particular, I consider the often overlooked issue of phylogenetic scale.

As used here, the term phylogenetic scale refers to the manner in which a set of taxa are indexed relative to an evolutionary hierarchy. The hierarchy could be based on a taxonomic classification system, in which case the term taxonomic scale would also apply, or the more explicit hierarchy of a phylogenetic tree model. Although similar concepts have been discussed under other names for many years, references to phylogenetic or taxonomic scale have appeared relatively recently in the community ecology literature (e.g. Williams et al. 1994, Roberts and Cohan 1995, Saiz-Salinas and Gonzalez-Oreja 1999, Pandolfi 2001, McGill et al. 2005, Oline 2006, Slingsby and Verboom 2006). Different authors have used these terms in different ways, suggesting a need for standardized usage and more specific terminology. A similar problem of ambiguous usage led the more widely-discussed spatial scale concept to be defined more explicitly in terms such as extent, grain and focus (Wiens 1989, Scheiner et al. 2000). One of the objectives of this paper is to clarify the meaning of phylogenetic scale by drawing parallels with these spatial scale concepts.

Both spatial scale and phylogenetic scale are important in community-environment studies because they jointly determine which evolutionary and ecological processes are highlighted by community patterns. If the highlighted processes are not relevant to the environmental variables of interest, they contribute to noise that dilutes the strength of correlative relationships. This paper uses conceptual models to illustrate the effects of interacting ecological and evolutionary processes on community patterns, including processes that may contribute to noise when community-environment relationships are the pattern of interest. I then consider the implications of adjusting an easily-manipulated component of phylogenetic scale—the phylogenetic grain, or resolution—for bringing community-environment relationships into sharper focus.

The idea that community patterns may be in sharper focus at some phylogenetic scales than others led to the development of an exploratory analysis technique capable of examining community patterns at multiple levels of a phylogenetic or taxonomic hierarchy. This technique, as introduced and applied in this paper, is a modification of species clustering expanded to allow taxa (or clades) above the species level to be included in clusters. Although environmental variables are not used directly in the clustering process, the clusters identified through this technique represent taxa with similar distributions that are likely to be correlated with one or more environmental variables at the spatial scale defined for the analysis. This technique is demonstrated here for a set of plant communities of the Southern Appalachian Mountains, U.S.A. using data from the Carolina Vegetation Survey (Peet et al. 1993).

Each section of this paper covers one of the stated objectives, namely to (1) clarify the meaning of phylogenetic scale by drawing parallels with spatial scale concepts

of extent, grain and focus; (2) elucidate ecological and evolutionary processes that can be inferred at various spatial and phylogenetic scales; (3) explain how noise in community-environment relationships can potentially be reduced by refocusing the phylogenetic grain; (4) introduce an exploratory analysis approach for phylogenetic refocusing of community patterns; and (5) demonstrate an application of phylogenetic refocusing using plant community data from the Carolina Vegetation Survey.

1. SCALE CONCEPTS: EXTENT, GRAIN AND FOCUS

Taxa in a community sample can be arranged in a nested hierarchy of evolutionary ancestry just as community sample units can be arranged in a hierarchy of spatial proximity. The similar hierarchical structure allows spatial scale concepts of extent, grain and focus to be logically translated into phylogenetic scale concepts (Fig. 4.1-4.2).

Spatial extent refers to the sampled spatial universe, e.g. the area covered by a set of plots (Fig. 4.1a) (Wiens 1989, Palmer and White 1994). By analogy, phylogenetic extent is that portion of the evolutionary universe covered by a community study (Fig. 4.1b). Major taxonomic groups traditionally used to define communities (vascular plants, mammals, arthropods, etc.) exemplify this concept². By this general definition phylogenetic extent would include all members of a specified group sharing a common ancestor, but in reality only a subset of the group will be represented: extant taxa residing within the spatial extent that are captured by sampling (Ackerly 2000, Webb et al. 2002). This definition of phylogenetic extent corresponds with recent usage of the terms

² Communities can also be defined without reference to phylogeny, e.g. in food web studies. Here I follow the tradition of taxonomically/trophically defined communities.

“phylogenetic scale” and “taxonomic scale” in studies of community phylogenetic structure (Cavender-Bares et al. 2006, Slingsby and Verboom 2006, Swenson et al. 2006).

Phylogenetic grain likewise draws its meaning through analogy with its spatial counterpart. Spatial grain refers to the size of sample units, such as the two-dimensional area of plant community plots (Fig. 4.1a) (Wiens 1989, Palmer and White 1994). The equivalent “sample units” of the evolutionary hierarchy are operational taxonomic units (OTUs) that serve as the currency of biodiversity in community studies (Kelt and Brown 2000, Schloss and Handelsman 2006), and the “size” of the OTUs is related to their position in the hierarchy. Phylogenetic grain can be defined as the level of phylogenetic resolution of OTUs, commonly expressed as a taxonomic rank (Fig. 4.1b). Community ecologists have traditionally used species as the taxonomic rank of choice, reflecting the importance placed on species in taxonomy, natural history and evolutionary biology (Kelt and Brown 2000, Rieseberg and Burke 2001, Sites and Marshall 2003). However, OTUs at levels of phylogenetic resolution above species (genera, families, phyla, etc.) or below (subspecies, varieties, ecotypes, etc.) have also found their place in community studies (e.g. Moral and Denton 1977, Deil 1989, Deil 1994, Kuerschner and Parolly 1999).

In principle, members of any taxonomic rank could be used for community OTUs provided they correspond to ecologically-meaningful units. Furthermore, the OTUs need not all reside at the same taxonomic rank, given that ranks do not necessarily convey a consistent ecological meaning for different groups classified by different taxonomists (Bachmann 1998). In practice, the taxonomic rank or level of phylogenetic resolution may differ among different groups within a community study according to the ease with

which taxonomic identities can be specified. The commonly-discussed tradeoff between taxonomic specificity and time investment in community studies (Beattie and Oliver 1994, Williams and Gaston 1994, Pik et al. 1999, Saiz-Salinas and Gonzalez-Oreja 1999) can be framed as an issue of phylogenetic grain.

Focus is another spatial scale concept referring to a level of sample unit aggregation (Scheiner et al. 2000). Sample units within some level of spatial proximity can be aggregated and averaged, effectively increasing the spatial grain size. When applied to phylogenetic scale, this translates into lumping of taxa; phylogenetic focus is the level at which taxa are lumped. Many studies have applied the phylogenetic focus concept by analyzing communities at taxonomic ranks higher than the level of recorded OTUs (Dale and Clifford 1976, Dale et al. 1989, Ferraro and Cole 1992, Somerfield and Clarke 1995, Bowman and Bailey 1997, Chapman 1998, Olsgard et al. 1998, Doledec et al. 2000, Mistri and Rossi 2000, Metzeling et al. 2002, Narayanaswamy et al. 2003, Anderson et al. 2005a, Anderson et al. 2005b). Phylogenetic focus could also conceivably involve aggregation to a level defined by branch length distance on a phylogenetic tree. Phylogenetic aggregation to either a uniform taxonomic rank or a consistent branch length distance are both examples of what I define as rigid phylogenetic focus, analogous to rigid spatial focus where sample units are aggregated within a specific spatial distance (Fig. 4.2a-b). Alternatively flexible focus, whether spatial or phylogenetic, is not based on external distance constraints but allows other criteria to guide the aggregation process, possibly resulting in units aggregated to different distances in different spatial or phylogenetic zones (Fig. 4.2c-d). The flexible focus concept is central to the analysis approach presented below (Sections 3-5).

2. PROCESSES LINKED TO SPATIAL AND PHYLOGENETIC SCALE

Community patterns at various spatial and phylogenetic scales can be linked to ecological and evolutionary processes operating at various temporal scales. For conventional communities composed of species (as the phylogenetic grain) within a taxonomic group (the phylogenetic extent), ecological processes of interest include species interactions and environmental filtering, which operate on shorter temporal scales than species-level evolutionary processes such as speciation and niche divergence.

At small spatial scales, one would expect to see the imprint of recent ecological processes resulting in combinations of species that are adapted to local environmental conditions and compatible with one another. Species with similar alpha-niches (similar interactive effects in a local community) would be expected to have limited compatibility, and thus rarely co-occur, while species with similar beta-niches (similar environmental tolerances and affinities) would be expected to be found together (Silvertown et al. 2001, Webb et al. 2002, Silvertown 2004, Silvertown et al. 2006).

Niche attributes of species are ultimately the result of evolutionary niche divergence over longer time scales. Within an evolving lineage, divergent niches may arise among related species as they diverge genetically, although ancestral niches might alternatively be retained through niche conservatism (Harvey and Pagel 1991, Webb et al. 2002). Niche conservatism leads to a correspondence between phylogenetic relatedness and niche attributes that should lead to patterns of local aggregation or segregation of related species, depending on whether the alpha or beta niche (or both) is conserved (Webb et al. 2002, Cavender-Bares et al. 2004, Cavender-Bares et al. 2006). More precisely, related species with conserved beta niches are likely to be segregated at small-

grain spatial scales if their alpha niches are conserved, but may be aggregated if their alpha niches are divergent (Fig. 4.3a, left column). If beta niches are not conserved, the alpha niche becomes irrelevant for small-grain patterns because the species are segregated into different environments where they do not interact (Fig. 4.3a, right column). Likewise, if species are not sympatric at the broader scale of their geographic ranges, small-grain aggregation will not be observed (Fig. 4.3b). However, allopatric species with conserved beta niches will likely occupy similar environments in their respective ranges (Fig. 4.3b, left column). Conserved beta niches can thus be said to result in aggregation by habitat, regardless of segregation patterns at small grain sizes or large spatial extents (Fig. 4.3).

The terms allopatric and sympatric are commonly associated with species-level genetic divergence processes (speciation), but are also more broadly applicable to divergence at any phylogenetic level. Allopatric divergence may be evident in spatially-segregated populations both prior to and following speciation, although the pattern may eventually be obscured by range shifts resulting in secondary contact of the divergent species (Barraclough and Vogler 2000, Barraclough and Nee 2001, Losos and Glor 2003). For the purposes of the model presented here, the terms allopatric and sympatric are defined empirically based on current distributions regardless of historical origins. As such, portions of a taxon's range may be sympatric and other portions allopatric with regard to a related taxon. The relevant point is that allopatric ranges may sometimes segregate related taxa that are, because of phylogenetic niche conservatism, ecologically similar. This becomes an issue when the spatial extent of a community study spans the ranges of such taxa.

Phylogenetic niche conservatism can likewise be defined empirically based on contemporary niche similarity of related taxa (cf. Webb et al. 2002), bypassing the species-level historical meaning sometimes associated with the term (Harvey and Pagel 1991). By expanding the niche conservatism concept beyond the species level, patterns of phylogenetic niche similarity can be considered for any set of related taxa or clades, i.e. at alternative phylogenetic grain sizes. Implications of alternative phylogenetic grain sizes on community pattern and process detection are considered further in the following section.

3. REFOCUSING THE PHYLOGENETIC SCALE OF COMMUNITY PATTERNS

As described in the previous section, spatial segregation of related taxa may occur at a small spatial grain because of alpha-niche similarities, or at a large spatial extent because of allopatric divergence. In addition, patterns of taxa in community samples can be attributed to an element of chance in which species end up at which sites (Gilbert and Lechowicz 2004, Bell 2005, Karst et al. 2005) and which sites end up being sampled. While these patterns and processes are of interest for their own sake, they constitute noise when the objective is to discover community patterns related specifically to the process of environmental filtering. Community classification studies, for example, commonly aim to define communities based on sets of species that jointly occupy similar habitats. If certain species are incompatible at the spatial grain of sampling, if certain species are missing from portions of the spatial extent, or if certain species are erratic in their distribution, the consistency of species occupying the habitat(s) of interest will be compromised.

Figure 4.4 illustrates this point through a hypothetical scenario in which community composition has been sampled at multiple locations spanning a large spatial extent. Each sample plot in Figure 4.4 captures the environmental conditions of a specific habitat that is scattered throughout the extent of the study area. However, the species composition of these plots, as illustrated by letters in Figure 4.4a, is not consistent across the area. Species A represents a widespread, frequently-encountered species, the only species present in all plots. Species B-F are geographically-restricted congeners whose ranges do not overlap because of recent allopatric speciation events. Species G-H are sympatric and widespread but are not found in close proximity because of alpha-niche conservatism resulting in resource competition. Lastly, species J-M have partially-overlapping ranges and seemingly random plot occupancy and abundance, suggesting neutral community assembly processes (Bell 2005).

If the phylogenetic grain is refocused, as illustrated in Fig. 4.4b, each of these clades (A, B-F, G-H, and J-M) is found to be present, in roughly equal proportions, in each of the plots. The conclusion to be drawn is that species within each clade have conserved beta-niches (relative to the habitat under study) and are thus suitable units for characterizing the community at the given spatial grain and extent. Noise is eliminated in this scenario as the phylogenetic grain of analysis is refocused from the species level to higher phylogenetic levels (with the exception of species A, which did not require refocusing to reach maximum consistency). Because the refocused levels are not constrained to a single taxonomic rank or uniform branch length distance across all lineages, this is an example of flexible phylogenetic refocusing (Section 1).

4. AN EXPLORATORY APPROACH FOR PHYLOGENETIC REFOCUSING

For real datasets, refocusing phylogenetic grain size by lumping species to an optimal, lineage-specific level may not be as simple as the hypothetical scenario shown in Figure 4.4. The hypothetical scenario considered only a single habitat instead of the multiple habitats likely to be included in a typical community classification study. Furthermore, discrete habitats may not easily be defined, especially if environmental variables are difficult to measure and/or occur as continuous gradients across the landscape. Patterns of beta-niche conservatism inferred from habitat occupancy will obviously depend on how the habitat is defined; different patterns of niche conservatism could conceivably be seen along different environmental gradients. Also, the empirical limits of niche conservatism (as drawn by the refocus line in Fig. 4.4b) may not be clearly marked in a phylogenetic hierarchy, especially in lineages where ecological traits have evolved gradually. There may not be a single ‘optimal’ phylogenetic focus where noise is reduced to the degree illustrated in Fig. 4.4b because of random or individualistic patterns of some taxa in relation to environmental gradients. All these complicating issues need to be taken into account when attempting to apply phylogenetic refocusing to real-world community analysis.

The community analysis technique introduced here addresses these issues by taking an exploratory approach for identifying community patterns at multiple levels of a phylogenetic hierarchy. The need for *a priori* habitat characterization is circumvented by using community patterns themselves to define classes or axes of environmental variation, a principle that has been routinely followed in community clustering and indirect gradient analysis (Kent and Coker 1992, Jongman et al. 1995, McCune and

Grace 2002). By analyzing a species/sample unit matrix on its own, then relating it to environmental variables secondarily, the species ‘inform’ the analyst about the environment as ‘seen’ from their own perspective. In the technique presented here, this type of analysis is expanded so that, in addition to species, other phylogenetic units (higher taxa or clades) can contribute to community patterns. The technique identifies combinations of phylogenetic units yielding the strongest community patterns given the data and the criteria supplied by the analyst (see below). By quantifying the strength of the pattern and rejecting weak patterns as noise, this technique accommodates the reality that not all lineages are likely to exhibit phylogenetic niche conservatism nor are all taxa likely to offer informative information about community-environment relationships.

The new analysis technique is a modified version of Coalition Clustering, a non-hierarchical agglomerative clustering algorithm that was designed to identify clusters of positively-associated species in community datasets (Sanderson et al. 2006). The Coalition Clustering algorithm iteratively joins species meeting a user-specified significance criterion into clusters. The significance criterion is a threshold level of mean pairwise association³. Clusters are built as species in the dataset are drawn in random order, the first species initiating the first cluster and subsequent species either being added to existing clusters or initiating new clusters. Species can be added to multiple existing clusters but cannot initiate new clusters if they can be added elsewhere. The clusters are progressively refined by combining similar clusters, adjusting their

³ Pairwise association is based on a two-by-two contingency table of presence/absence. Association coefficients such chi-square and phi utilize information from all four cells of the table including joint absences. Although described here in terms of association, Coalition Clustering can also be carried out using co-occurrence indices that omit joint absences (e.g. Jaccard Index, Sorensen Index) or correlation coefficients that allow the use of abundance information (e.g. Pearson’s R) (Ludwig and Reynolds 1988).

composition and weeding out single-species clusters until a stable set of clusters is obtained. Changing the significance criterion affects the size and number of clusters; a stringent criterion tends to produce many clusters containing few species, while a relaxed criterion tends to produce few clusters containing many species (Sanderson et al. 2006). Species belonging to clusters can be viewed as characteristic species of abstract (compositionally-defined) community types, which can then be used to characterize concrete (spatially-defined, albeit not necessarily discrete) communities based on the degree to which site species composition resembles cluster composition. Concrete community characterization opens the door for correlative analysis between coalition clusters and environmental variables (Sanderson et al. 2006).

The modified version of Coalition Clustering, referred to as Multilevel Ordered Coalition Clustering (MOCC) differs from the original algorithm described by Sanderson et al. (2006) in two principal ways. First, it incorporates an ordered clustering strategy that builds a cluster for each species in the dataset, using each species to initiate a cluster and then adding other species one at a time in descending order of association strength (S.C. Sanderson, pers. comm.). New species are added to each cluster, or removed as needed, until no more species meet the significance criterion. Some species may fail to accrete clusters and such species are removed. Once a cluster has been built for each eligible species, the set of unique clusters can be examined and, if desired, similar clusters can be merged. This procedure has the advantage of providing a unique clustering solution, in contrast to the random accretion procedure of the original Coalition Clustering algorithm for which the solution can vary under identical settings.

The second way in which MOCC differs from the original Coalition Clustering algorithm is by allowing higher taxa or clades to enter into clusters alongside species (hence the prefix ‘multilevel’). This is not a trivial addition to the algorithm, despite the fact that species occurrences can be easily converted to higher taxon/clade occurrences if an appropriate phylogeny or classification hierarchy is available. The challenge arises in building clusters that do not contain phylogenetically-redundant information, as would be the case if a species, plus the genus, family, etc. to which it belongs, were included in the same cluster. Avoiding this situation requires specifying which taxa/clades (hereafter ‘units’) are phylogenetically independent, i.e. not overlapping members of the same lineage. Figure 4.5 illustrates how a pairwise species association matrix can be expanded to a multilevel matrix with non-independent pairwise association values removed. Such a matrix is suitable for selecting the first pair of units in a cluster, but before subsequent units can be added the matrix must be re-filtered to exclude all units that are not independent from either unit of the initial cluster pair. Each addition or subtraction of a unit from each cluster requires a re-assessment of which pairwise association values are phylogenetically independent. The order in which units are accreted to a cluster has a cascading effect on the accretion eligibility of the remaining units. In consequence of these features of the multilevel algorithm, many different combinations of units are possible for clusters that are actually compositionally similar under the surface. There is no easy way to merge compositionally-similar multilevel clusters if they contain phylogenetically non-independent variants of the same lineages; thus the multilevel algorithm (as currently configured) leaves clusters unmerged and allows the analyst to visually compare and screen them as needed.

5. APPLICATION OF PHYLOGENETIC SCALE REFOCUSING FOR SOUTHERN APPALACHIAN PLANT COMMUNITIES

As a simple demonstration of the phylogenetic refocusing analysis approach described above, I present an application of MOCC to a plant community dataset from the Carolina Vegetation Survey (Peet et al. 1993). A dataset of 219 vegetation survey plots was selected representing two habitats in the Southern Appalachian Mountains (USA): high elevation rock outcrops (Wiser et al. 1996) and mountain bogs (Wichmann 2009) (Figure 4.6). This dataset captures the contrasting habitats of the drier, exposed outcrops and the wetter, more sheltered bogs, which are known to harbor different plant communities. The Southern Appalachian Region is also known for having geographically-variable plant species composition that cannot easily be correlated with environmental variables at the regional extent scale (Newell et al. 1995, Nekola and White 1999). Species composition of outcrops and bogs would be expected to be particularly variable between different sites because of the insular, spatially restricted distribution of these habitats. Restricted geographical ranges, species incompatibilities and random community assembly could all contribute to within-habitat species compositional variation. By applying the phylogenetic refocusing approach to these communities, one can investigate whether such noisy variation within habitats can be offset by expanding the phylogenetic grain size beyond the species level.

The 623 OTUs represented by this dataset (primarily species, but also some higher taxa that were not identified to the species level) were placed in a taxonomic hierarchy based on the classification schemes of Weakley (2006) for the genus level and below, and NCBI (2008) above the genus level. Although not strictly phylogenetic, these classification schemes incorporate findings of recent phylogenetic systematic studies for

large numbers of taxa, and hence are useful sources of evolutionary relationship information at a broad phylogenetic extent.

Pairwise association values for taxa at each level of the taxonomic hierarchy were calculated using the phi coefficient (Ludwig and Reynolds 1988). The pairwise association matrix was then fed into the MOCC algorithm, programmed using the R package for statistical computing (R Development Core Team 2008). A subset of clusters extracted at a significance level of phi=0.2 are shown in Table 4.1. The analysis revealed strong associations among groups at different phylogenetic levels as allowed under the flexible phylogenetic focus principle. Within single clusters were found multiple levels including orders, families, tribes, genera, subgenera, sections, species, and varieties. Clusters of taxa characteristic of rock outcrops were distinguishable from bog clusters based on *a priori* knowledge of bog and rock outcrop communities. In addition, some clusters appear to represent taxa common to both habitats (Table 4.1, General Bog + Rock Outcrop Cluster). The MOCC technique proved useful for highlighting community differences as well as commonalities, with flexible phylogenetic focusing revealing stronger patterns than would be captured in a strictly species-level analysis, given the spatial scale of the dataset.

CONCLUSION

Describing phylogenetic scale in terms of grain, extent and focus provides a useful conceptual framework for understanding issues related to taxonomic resolution in community studies. Community analysis using species or other fine-grained taxonomic units may introduce unnecessary noise that can be dampened by lumping taxa to a higher

phylogenetic grain. This resembles the practice of grouping species into guilds, functional groups or trait-based groups (Wilson 1999, Duckworth et al. 2000, Pillar and Sosinski 2003), but does not require trait information beyond phylogenetic relationships. Trait-based groups may be more amenable to certain questions related to, for example, ecosystem attributes, or when comparing communities at intercontinental scales were niche conservatism is overshadowed by convergence of members of different lineages that have evolved similar adaptations to similar environments.

The flexible phylogenetic grain approach also offers advantages over current phylogenetic community analysis methods requiring branch length information and rigid phylogenetic grain (e.g. Webb et al. 2008). Reliable branch length information is often difficult to obtain for “supertrees” representing all members of a community at their highest phylogenetic resolution (Sanderson and Driskell 2003). In contrast, a supertree lacking branch length information is relatively easily to assemble, with taxonomic classification systems offering a default in the absence of more detailed data. Although the branch length difficulty is likely to diminish as phylogenetic systematics continues to progress, the value of branch length information in the ecological context remains an open question. The degree of ecological divergence is sometimes but not always correlated with the degree of genetic divergence between taxa (cf. Losos 2008). This perspective lends itself to the phylogenetic community analysis approach I have presented, which seeks to identify the position of ecological discontinuities in the phylogenetic hierarchy rather than examine the correlative relationship between phylogenetic distance and ecological variables.

Issues of spatial and phylogenetic scale are of particular importance as the scope of community studies expands to encompass larger areas and greater numbers of taxa. Databases of consolidated community samples drawn from multiple studies are increasingly available for analyses aimed at characterizing and understanding community variation (Mucina et al. 1993, Peet et al. 1993, Bruelheide and Chytry 2000). As the spatial extent of community analysis expands to encompass geographic turnover in species ranges, ecologists should consider whether the species level (or any other standard taxonomic rank) is the optimal phylogenetic grain for their analysis. Even in studies carried out at a more local scale, the implications of α -niche conservatism on segregation of closely-related species should be considered in the selection of spatial and phylogenetic grain. In some cases it may be feasible and appropriate to take the phylogenetic focus concept in the opposite direction, breaking OTUs into even finer units than have been presented here, such as haplotypes or genotypes.

The principles and techniques of phylogenetic grain adjustment presented here could also be extended to spatial grain adjustment, providing an alternative to autocorrelative approaches for analyzing spatial influences on sample units (Lichstein et al. 2002, Perry et al. 2002, Podani and Csontos 2006). Rigid phylogenetic re-focusing is conceptually analogous to merging sample units within a zone of spatial autocorrelation as identified through correlograms or semivariograms (Rossi et al. 1992, Wagner 2003). Flexible phylogenetic refocusing, by similar analogy, corresponds to spatial autocorrelation techniques that recognize non-stationarity, or differences in the strength of spatial autocorrelation that differ by location (Osborne et al. 2007). Identifying zones of “spatial community conservatism” due to high dispersal and low environmental

variation, separated by zones of “spatial community divergence” due to opposing processes, is an avenue where parallels between spatial and phylogenetic scale could potentially complement one another.

REFERENCES

- Ackerly, D. D. 2000. Taxon sampling, correlated evolution, and independent contrasts. *Evolution* 54:1480-1492.
- Anderson, M. J., S. D. Connell, B. M. Gillanders, C. E. Diebel, W. M. Blom, J. E. Saunders, and T. J. Landers. 2005a. Relationships between taxonomic resolution and spatial scales of multivariate variation. *Journal of Animal Ecology* 74:636-646.
- Anderson, M. J., C. E. Diebel, W. M. Blom, and T. J. Landers. 2005b. Consistency and variation in kelp holdfast assemblages: Spatial patterns of biodiversity for the major phyla at different taxonomic resolutions. *Journal of Experimental Marine Biology and Ecology* 320:35-56.
- Bachmann, K. 1998. Species as units of diversity: an outdated concept. *Theory in Biosciences* 117:213-230.
- Barraclough, T. G., and S. Nee. 2001. Phylogenetics and speciation. *Trends in Ecology and Evolution* 16:391-399.
- Barraclough, T. G., and A. P. Vogler. 2000. Detecting the geographical pattern of speciation from species-level phylogenies. *American Naturalist* 155:419-434.
- Beattie, A. J., and I. Oliver. 1994. Taxonomic minimalism. *Trends in Ecology and Evolution* 9:488-490.
- Bell, G. 2005. The co-distribution of species in relation to the neutral theory of community ecology. *Ecology* 86:1757-1770.
- Bowman, M. F., and R. C. Bailey. 1997. Does taxonomic resolution affect the multivariate description of the structure of freshwater benthic macroinvertebrate communities? *Canadian Journal of Fisheries and Aquatic Sciences* 54:1802-1807.
- Bruelheide, H., and M. Chytry. 2000. Towards unification of national vegetation classifications: A comparison of two methods for analysis of large data sets. *Journal of Vegetation Science* 11:295-306.
- Cavender-Bares, J., D. D. Ackerly, D. A. Baum, and F. A. Bazzaz. 2004. Phylogenetic overdispersion in Floridian oak communities. *American Naturalist* 163:823-843.
- Cavender-Bares, J., A. Keen, and B. Miles. 2006. Phylogenetic structure of Floridian plant communities depends on taxonomic and spatial scale. *Ecology* 87:S109-S122.

- Chapman, M. G. 1998. Relationships between spatial patterns of benthic assemblages in a mangrove forest using different levels of taxonomic resolution. *Marine Ecology Progress Series* 162:71-78.
- Dale, M. B., and H. T. Clifford. 1976. On the effectiveness of higher taxonomic ranks for vegetation analysis. *Australian Journal of Ecology* 1:37-62.
- Dale, M. B., E. Feoli, and P. Ganis. 1989. Incorporation of information from the taxonomic hierarchy in comparing vegetation types. *Taxon* 38:216-227.
- Deil, U. 1989. Adianteteae communities on the Arabian Peninsula, coenosyntaxa within this class, and general remarks on the phylogeny of plant communities. *Flora (Jena)* 182:247-264.
- Deil, U. 1994. Classification with supraspecific taxa and symphylogenetic approaches in phytosociology. *Phytocoenologia* 24:677-694.
- Doledec, S., J. M. Olivier, and B. Statzner. 2000. Accurate description of the abundance of taxa and their biological traits in stream invertebrate communities: effects of taxonomic and spatial resolution. *Archiv Fur Hydrobiologie* 148:25-43.
- Duckworth, J. C., M. Kent, and P. M. Ramsay. 2000. Plant functional types: an alternative to taxonomic plant community description in biogeography? *Progress in Physical Geography* 24:515-542.
- Ferraro, S. P., and F. A. Cole. 1992. Taxonomic level sufficient for assessing a moderate impact on macrobenthic communities in Puget-Sound, Washington, USA. *Canadian Journal of Fisheries and Aquatic Sciences* 49:1184-1188.
- Fowler, R. T., and R. G. Death. 2000. Effects of channel morphology on temporal variation in invertebrate community structure in two North Island, New Zealand rivers. *N. Z. J. Mar. Freshw. Res.* 34:231-240.
- Gilbert, B., and M. J. Lechowicz. 2004. Neutrality, niches, and dispersal in a temperate forest understory. *Proceedings of the National Academy of Sciences of the United States of America* 101:7651-7656.
- Harvey, P. H., and M. D. Pagel. 1991. *The Comparative Method in Evolutionary Biology*. Oxford University Press, Oxford; New York.
- Jennings, M. D., D. Faber-Langendoen, O. L. Loucks, R. K. Peet, and D. Roberts. 2009. Standards for associations and alliances of the US National Vegetation Classification. *Ecological Monographs* 79:173-199.
- Jongman, R. H. G., C. J. F. ter Braak, and O. F. R. Van Tongeren. 1995. *Data Analysis in Community and Landscape Ecology*. Cambridge University Press.

- Karst, J., B. Gilbert, and M. J. Lechowicz. 2005. Fern community assembly: The roles of chance and the environment at local and intermediate scales. *Ecology* 86:2473-2486.
- Kelt, D. A., and J. H. Brown. 2000. Species as units of analysis in ecology and biogeography: are the blind leading the blind? *Global Ecology and Biogeography* 9:213-217.
- Kent, M., and P. Coker. 1992. *Vegetation Description and Analysis: A Practical Approach*. CRC Press, Boca Raton.
- Kuerschner, H., and G. Parolly. 1999. Pantropical epiphytic rain forest bryophyte communities—Coeno-syntaxonomy and floristic-historical implications. *Phytocoenologia* 29:1-52.
- Lichstein, J. W., T. R. Simons, S. A. Shriner, and K. E. Franzreb. 2002. Spatial autocorrelation and autoregressive models in ecology. *Ecological Monographs* 72:445-463.
- Losos, J. B. 2008. Phylogenetic niche conservatism, phylogenetic signal and the relationship between phylogenetic relatedness and ecological similarity among species. *Ecology Letters* 11:995-1003.
- Losos, J. B., and R. E. Glor. 2003. Phylogenetic comparative methods and the geography of speciation. *Trends in Ecology and Evolution* 18:220-227.
- Ludwig, J. A., and J. F. Reynolds. 1988. *Statistical Ecology: A Primer on Methods and Computing*. John Wiley & Sons, New York.
- McCune, B., and J. B. Grace. 2002. *Analysis of Ecological Communities*. MjM Software Design, Gleneden Beach, OR.
- McGill, B. J., E. A. Hadly, and B. A. Maurer. 2005. Community inertia of Quaternary small mammal assemblages in North America. *Proceedings of the National Academy of Sciences of the United States of America* 102:16701-16706.
- Metzeling, L., D. Robinson, S. Perriss, and R. Marchant. 2002. Temporal persistence of benthic invertebrate communities in south-eastern Australian streams: taxonomic resolution and implications for the use of predictive models. *Marine and Freshwater Research* 53:1223-1234.
- Mistri, M., and R. Rossi. 2000. Levels of taxonomic resolution and choice of transformation sufficient to detect community gradients: an approach to hard-substrata benthic studies. *Italian Journal of Zoology* 67:163-167.

- Moral, R. D., and M. F. Denton. 1977. Analysis and classification of vegetation based on family composition. *Vegetatio* 34:155-165.
- Mucina, L., J. S. Rodwell, J. H. J. Schaminee, and H. Dierschke. 1993. European vegetation survey: current state of some national programmes. *Journal of Vegetation Science* 4:429-438.
- Mueller-Dombois, D., and H. Ellenberg. 1974. *Aims and Methods of Vegetation Ecology*. Wiley, New York.
- Narayanaswamy, B. E., T. D. Nickell, and J. D. Gage. 2003. Appropriate levels of taxonomic discrimination in deep-sea studies: Species vs family. *Mar. Ecol.-Prog. Ser.* 257:59-68.
- NCBI. 2008. The National Center for Biotechnology Information. <http://www.ncbi.nlm.nih.gov/>.
- Nekola, J. C., and P. S. White. 1999. The distance decay of similarity in biogeography and ecology. *Journal of Biogeography* 26:867-878.
- Newell, C. L., K. D. Patterson, T. R. Wentworth, and R. K. Peet. 1995. Consistency of vegetation patterns across three landscapes in the Southern Appalachian Mountains of North Carolina. *Bulletin of the Ecological Society of America* 76:370-371.
- Oline, D. K. 2006. Phylogenetic comparisons of bacterial communities from serpentine and nonserpentine soils. *Applied and Environmental Microbiology* 72:6965-6971.
- Olsgard, F., P. J. Somerfield, and M. R. Carr. 1998. Relationships between taxonomic resolution, macrobenthic community patterns and disturbance. *Marine Ecology Progress Series* 172:25-36.
- Osborne, P. E., G. M. Foody, and S. Suarez-Seoane. 2007. Non-stationarity and local approaches to modelling the distributions of wildlife. *Diversity and Distributions* 13:313-323.
- Palmer, M. W., and P. S. White. 1994. Scale dependence and the species-area relationship. *American Naturalist* 144:717-740.
- Pandolfi, J. M. 2001. Numerical and taxonomic scale of analysis in paleoecological data sets: Examples from neo-tropical Pleistocene reef coral communities. *Journal of Paleontology* 75:546-563.
- Peet, R. K., T. R. Wentworth, P. S. White, M. P. Schafale, and A. S. Weakley. 1993. The North Carolina Vegetation Survey: A university-agency collaboration. *Bulletin of the Ecological Society of America* 74.

- Perry, J. N., A. M. Liebhold, M. S. Rosenberg, J. Dungan, M. Miriti, A. Jakomulska, and S. Citron-Pousty. 2002. Illustrations and guidelines for selecting statistical methods for quantifying spatial pattern in ecological data. *Ecography* 25:578-600.
- Pik, A. J., I. Oliver, and A. J. Beattie. 1999. Taxonomic sufficiency in ecological studies of terrestrial invertebrates. *Australian Journal of Ecology* 24:555-562.
- Pillar, V. D., and E. E. Sosinski, Jr. 2003. An improved method for searching plant functional types by numerical analysis. *Journal of Vegetation Science* 14:323-332.
- Podani, J., and P. Csontos. 2006. Quadrat size dependence, spatial autocorrelation and the classification of community data. *Community Ecology* 7:117-127.
- R_Development_Core_Team. 2009. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria.
<http://www.r-project.org/>
- Rieseberg, L. H., and J. M. Burke. 2001. The biological reality of species: gene flow, selection, and collective evolution. *Taxon* 50:47-67.
- Roberts, M. S., and F. M. Cohan. 1995. Recombination and migration rates in natural populations of *Bacillus subtilis* and *Bacillus mojavensis*. *Evolution* 49:1081-1094.
- Rossi, R. E., D. J. Mulla, A. G. Journel, and E. H. Franz. 1992. Geostatistical tools for modeling and interpreting ecological spatial dependence. *Ecological Monographs* 62:277-314.
- Saiz-Salinas, J. I., and J. A. Gonzalez-Oreja. 1999. Measuring redundancy in data from pollution impact studies. *Environmental Management* 24:415-420.
- Sanderson, M. J., and A. C. Driskell. 2003. The challenge of constructing large phylogenetic trees. *Trends in Plant Science* 8:374-379.
- Sanderson, S. C., J. E. Ott, E. D. McArthur, and K. T. Harper. 2006. RCLUS, a new program for clustering associated species: A demonstration using a Mojave Desert plant community dataset. *Westerns North American Naturalist* 66:285-297.
- Scheiner, S. M., S. B. Cox, M. Willig, G. G. Mittelbach, C. Osenberg, and M. Kaspari. 2000. Species richness, species-area curves and Simpson's paradox. *Evolutionary Ecology Research* 2:791-802.
- Schloss, P. D., and J. Handelsman. 2006. Introducing SONS, a tool for operational taxonomic unit-based comparisons of microbial community memberships and structures. *Applied and Environmental Microbiology* 72:6773-6779.

- Silvertown, J. 2004. Plant coexistence and the niche. *Trends in Ecology and Evolution* 19:605-611.
- Silvertown, J., M. Dodd, and D. Gowing. 2001. Phylogeny and the niche structure of meadow plant communities. *Journal of Ecology* 89:428-435.
- Silvertown, J., M. Dodd, D. Gowing, C. Lawson, and K. Mcconway. 2006. Phylogeny and the hierarchical organization of plant diversity. *Ecology* 87:S39-S49.
- Sites, J. W., Jr., and J. C. Marshall. 2003. Delimiting species: A renaissance issue in systematic biology. *Trends in Ecology and Evolution* 18:462-470.
- Slingsby, J. A., and G. A. Verboom. 2006. Phylogenetic relatedness limits co-occurrence at fine spatial scales: Evidence from the schoenoid sedges (Cyperaceae: Schoeneae) of the Cape Floristic Region, South Africa. *American Naturalist* 168:14-27.
- Sluys, R. 1991. Species concepts, process analysis, and the hierarchy of nature. *Experientia* 47:1162-1170.
- Somerfield, P. J., and K. R. Clarke. 1995. Taxonomic levels, in marine community studies, revisited. *Marine Ecology Progress Series* 127:113-119.
- Swenson, N. G., B. J. Enquist, J. Pither, J. Thompson, and J. K. Zimmerman. 2006. The problem and promise of scale dependency in community phylogenetics. *Ecology* 87:2418-2424.
- Wagner, H. H. 2003. Spatial covariance in plant communities: Integrating ordination, geostatistics, and variance testing. *Ecology* 84:1045-1057.
- Weakley, A. S. 2006. Flora of the Carolinas, Virginia, and Georgia, and Surrounding Areas. http://www.herbarium.unc.edu/FloraArchives/WeakleyFlora_2006-Jan.pdf
- Webb, C. O., D. D. Ackerly, and S. W. Kembel. 2008. Phylocom: software for the analysis of phylogenetic community structure and trait evolution. *Bioinformatics* 24:2098-2100.
- Webb, C. O., D. D. Ackerly, M. A. Mcpeak, and M. J. Donoghue. 2002. Phylogenies and community ecology. *Annual Review of Ecology and Systematics* 33:475-505.
- Wichmann, B. L. 2009. Vegetation of geographically isolated montane non-alluvial wetlands of the southern Blue Ridge of North Carolina. M.S. Thesis, North Carolina State University.
- Wiens, J. A. 1989. Spatial scaling in ecology. *Functional Ecology* 3:385-397.

- Williams, P. H., and K. J. Gaston. 1994. Measuring more of biodiversity—can higher-taxon richness predict wholesale species richness? *Biological Conservation* 67:211-217.
- Williams, P. H., C. J. Humphries, and K. J. Gaston. 1994. Centers of seed-plant diversity—the family way. *Proceedings of the Royal Society of London Series B-Biological Sciences* 256:67-70.
- Wilson, J. B. 1999. Guilds, functional types and ecological groups. *Oikos* 86:507-522.
- Wiser, S. K., R. K. Peet, and P. S. White. 1996. High-elevation rock outcrop vegetation of the Southern Appalachian Mountains. *Journal of Vegetation Science* 7:703-722.

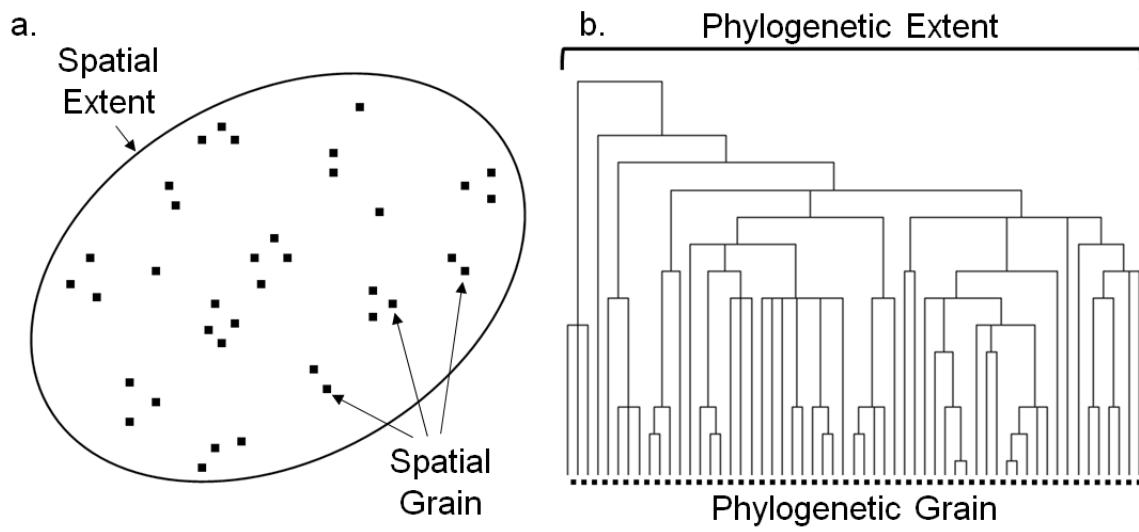


Figure 4.1. Conceptual parallels between spatial scale and phylogenetic scale of community studies: extent and grain. Spatial extent is the entire area covered by sampling; spatial grain the size of sample units (a). By analogy, phylogenetic extent is the entire lineage sampled; phylogenetic grain the resolution of operational taxonomic units (b).

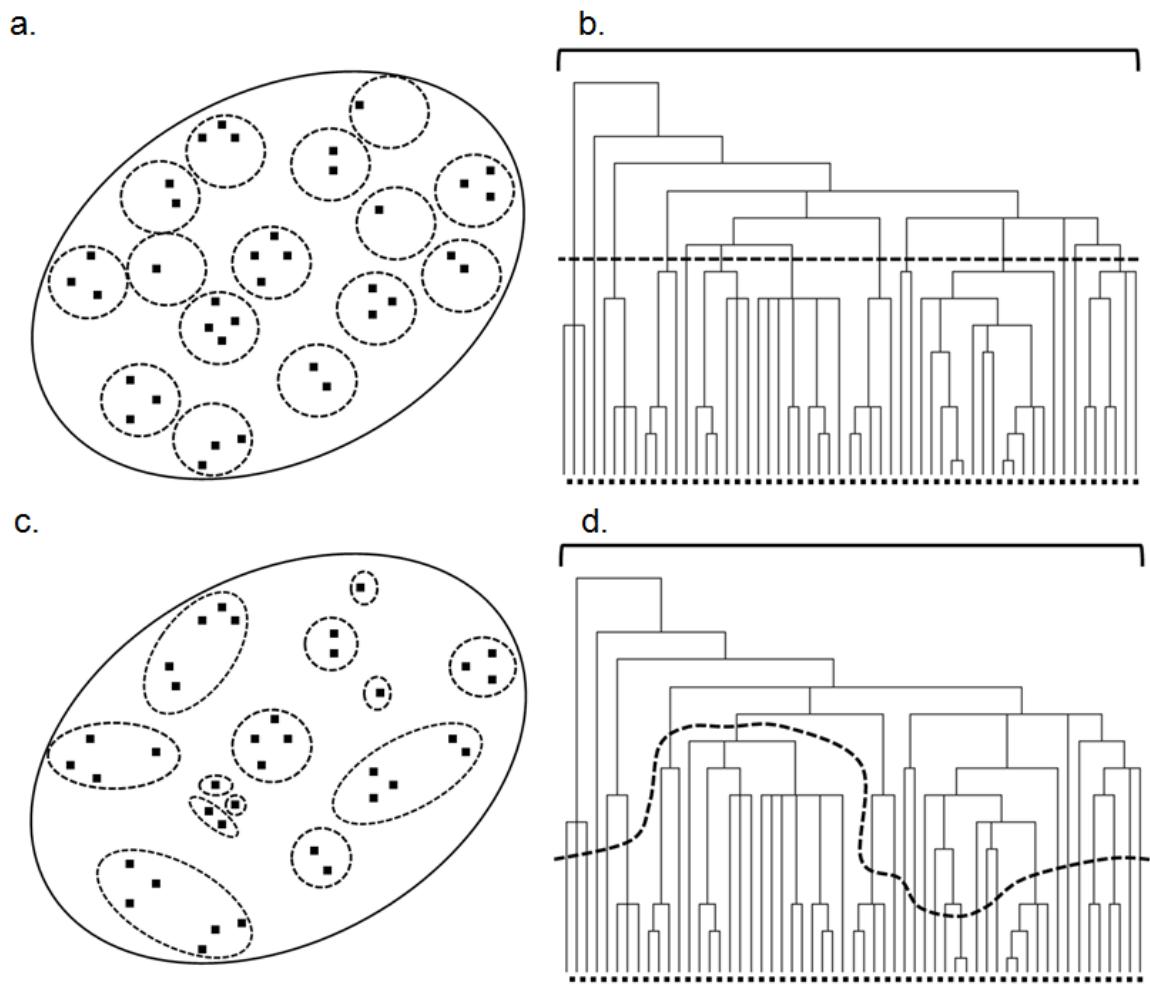


Figure 4.2. Conceptual parallels between spatial scale and phylogenetic scale of community studies: focus. Dashed lines show limits of refocused (aggregated) spatial sample units (left) or operational taxonomic units (right). Rigid focus is based on a uniform spatial distance (a) or phylogenetic distance (b). Flexible focus uses criteria other than distance to aggregate adjacent sample units (c) or operational taxonomic units (d).

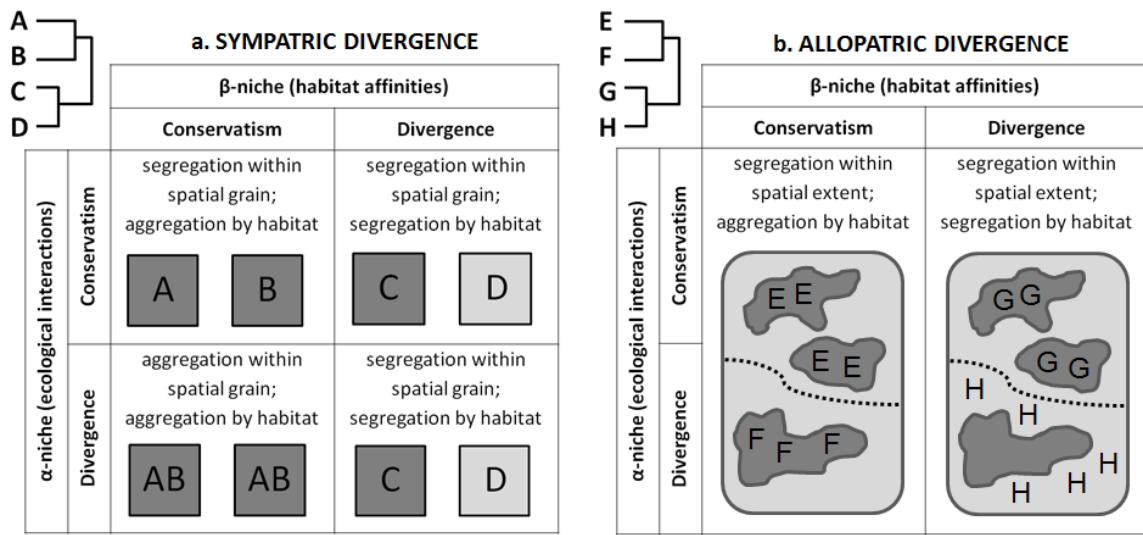


Figure 4.3. Conceptual model of expected aggregation and segregation patterns of closely related species under different scenarios of spatial and ecological divergence, building on Webb et al. (2002). Phylogenetic trees in upper left of a and b show relationships among hypothetical taxa whose expected spatial patterns are diagramed. a) Expected patterns of sympatric species within fine-grained sample units (squares) in different habitats (grayscale shades). b) Expected patterns of allopatric species within large spatial extents containing different habitats (grayscale shaded shapes). The dashed line represents a zone separating allopatric species ranges.

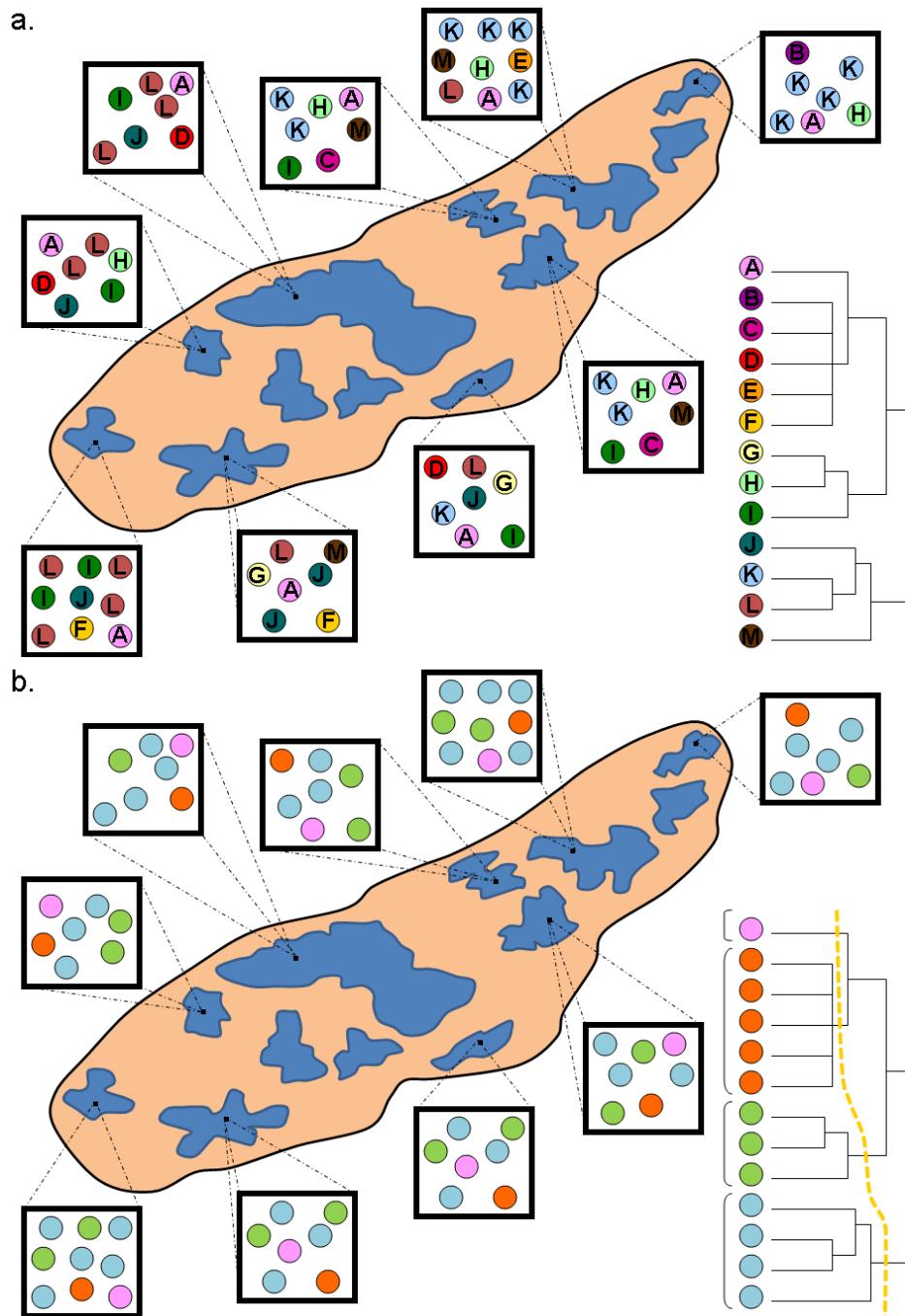


Figure 4.4. Hypothetical scenario illustrating phylogenetic refocusing of community patterns. a and b depict the same study area with different habitats shown in different colors (mimicking uplands and lowlands of the Southern Appalachian Region, USA). Squares are sample plots from different locations within the ‘upland’ habitat, containing taxa represented by letters/symbols, related according to phylogenetic models to the right. a) Species-level patterns with inconsistent composition in the ‘upland’ habitat. b) Phylogenetically-refocused pattern with consistent composition in the ‘upland’ habitat.

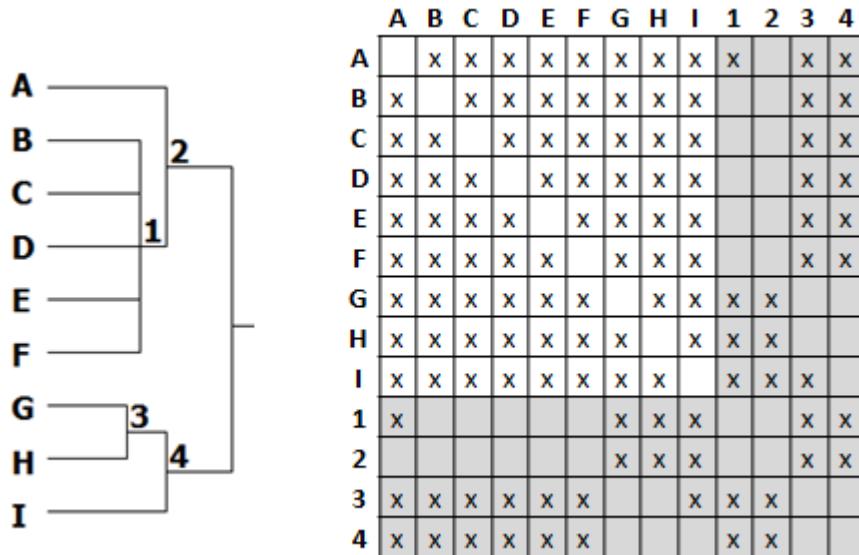


Figure 4.5. Expansion of a species by species pairwise association matrix to include higher taxa. Letters are species, numbers are higher taxa, related as shown in the phylogenetic tree (left). Shaded area of the full matrix (right) shows expansion due to inclusion of higher taxa. An "x" indicates taxon pairs that are phylogenetically independent (not overlapping members of the same lineage).

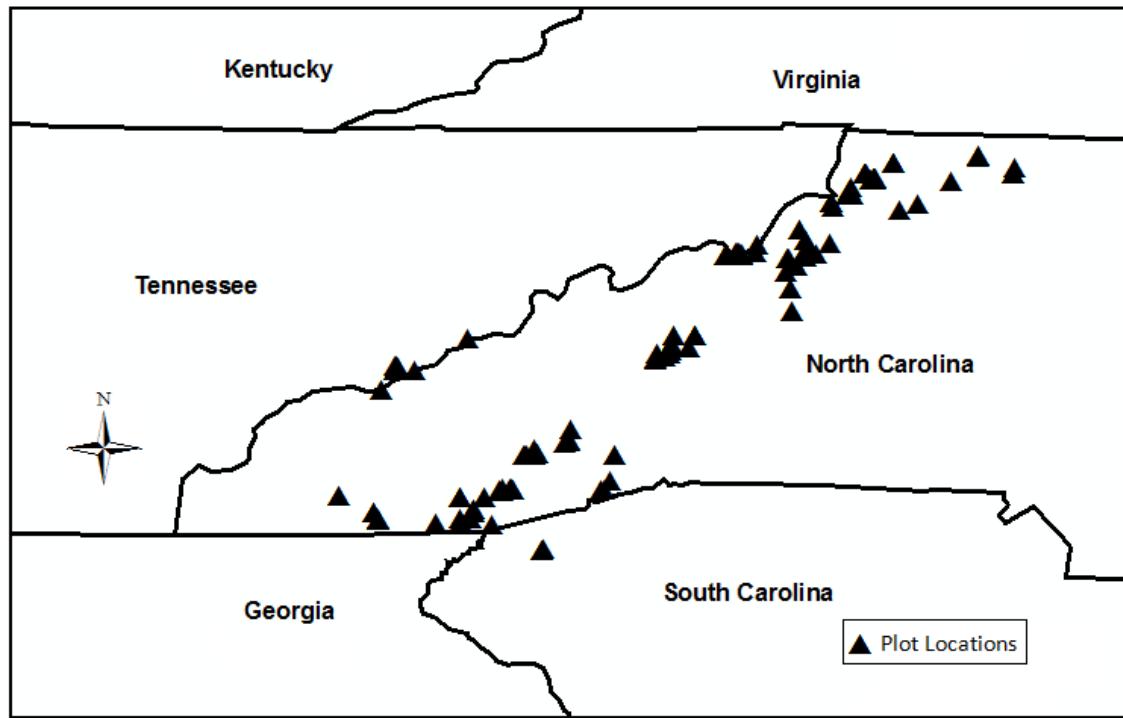


Figure 4.6. Location of high elevation rock outcrop and mountain bog vegetation survey plots in the Southern Appalachian Mountains, USA, analyzed using Multilevel Ordered Coalition Clustering (MOCC).

Table 4.1. Multilevel Coalition Clusters obtained by applying the MOCC algorithm to a plant community dataset of 219 100 m² vegetation plots taken from the Carolina Vegetation Survey, representing high elevation rock outcrops and mountain bogs.

Bog Cluster		Rock Outcrop Cluster 1	
Taxon	Rank	Taxon	Rank
<i>Acer rubrum</i>	Species	<i>Andropogoneae</i>	Tribe
<i>Carex leptalea</i>	Species	<i>Carex umbellata</i>	Species
<i>Clematis virginiana</i>	Species	<i>Danthonia sericea</i>	Species
<i>Drosera rotundifolia</i> var. <i>rotundifolia</i>	Variety	<i>Dichanthelium</i>	Genus
<i>Eleocharis</i>	Subgenus	<i>Houstonia longifolia</i>	Species
<i>Eriophorum virginicum</i>	Species	<i>Hypericum gentianoides</i>	Species
<i>Eupatorium perfoliatum</i>	Species	<i>Selaginella</i>	Genus
<i>Galium</i>	Genus		
<i>Glyceria</i>	Genus		
<i>Impatiens</i>	Genus		
<i>Juncus</i>	Genus		
<i>Lycopus</i>	Genus		
<i>Lyonia</i>	Genus		
<i>Myrtales</i>	Order		
<i>Orchidaceae</i>	Family		
<i>Osmunda</i>	Genus		
<i>Ovales</i>	Section		
<i>Packera aurea</i>	Species		
<i>Persicaria</i>	Genus		
<i>Rhynchospora</i>	Genus		
<i>Rosa</i>	Genus		
<i>Rubus hispida</i>	Species		
<i>Salix sericea</i>	Species		
<i>Scirpus</i>	Genus		
<i>Solidago altissima</i> var. <i>altissima</i>	Variety		
<i>Solidago patula</i>	Species		
<i>Stellulatae</i>	Section		
<i>Sympyotrichum puniceum</i>	Species		
<i>Thelypteris</i>	Genus		
<i>Trigynobrathys</i>	Section		
<i>Vaccinium macrocarpon</i>	Species		
<i>Vesicariae</i>	Section		
<i>Viola</i>	Genus		
Rock Outcrop Cluster 2			
Taxon	Rank		
<i>Alnus viridis</i> var. <i>crispa</i>	Variety		
<i>Deschampsia</i>	Genus		
<i>Minuartia groenlandica</i>	Species		
General Bog + Rock Outcrop Cluster			
Taxon	Rank		
<i>Cyanococcus</i>	Section		
<i>Hymenanthes</i>	Subgenus		
<i>Kalmia</i>	Genus		

APPENDIX 2.1

Taxa recorded by the 1999-2000 USGS-NPS vegetation survey of Zion National Park, Utah (Cogan et al. 2004), following nomenclature of Kartesz (1999), with translations to equivalent taxonomic concepts of the 1987-89 BYU-RMRS survey (Harper et al. 2001), updated to nomenclature of Welsh et al. (2003). USGS-NPS names are as they appear in the publicly-available database (USGS 2004); only taxa identified to species or genus are shown. Blanks in the BYU-RMRS column indicate taxa not recorded by the 1987-89 vegetation survey or (in the case of some genera) translating to multiple ecologically heterogeneous congeners. #Occ.=number of plot occurrences out of 346 (USGS-NPS) and 288 (BYU-RMRS) plots total.

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Abies concolor</i> (Gord. & Glend.) Lindl. ex Hildebr.	39	<i>Abies concolor</i>	19
<i>Abronia fragrans</i> Nutt. ex Hook.	4	<i>Abronia fragrans</i>	20
<i>Acer grandidentatum</i> Nutt.	35	<i>Acer grandidentatum</i>	29
<i>Acer negundo</i> L.	26	<i>Acer negundo</i>	18
<i>Achillea millefolium</i> L.	24	<i>Achillea millefolium</i>	6
<i>Achnatherum contractum</i> (B.L. Johnson) Barkworth	1	<i>Stipa hymenoides</i>	65
<i>Achnatherum hymenoides</i> (Roemer & J.A. Schultes) Barkworth	63	<i>Stipa hymenoides</i>	65
<i>Achnatherum lettermanii</i> (Vasey) Barkworth	12	<i>Stipa lettermanii</i>	12
<i>Achnatherum nelsonii</i> ssp. <i>nelsonii</i> (Scribn.) Barkworth	2	<i>Stipa nelsonii</i>	5
<i>Achnatherum speciosum</i> (Trin. & Rupr.) Barkworth	5	<i>Stipa speciosa</i>	15
<i>Agastache urticifolia</i> (Benth.) Kuntze	3	<i>Agastache urticifolia</i>	1
<i>Agoseris</i> Raf.	3		
<i>Agropyron cristatum</i> (L.) Gaertn.	3	<i>Agropyron cristatum</i>	1
<i>Agrostis exarata</i> Trin.	3	<i>Agrostis exarata</i>	3
<i>Agrostis</i> L.	2		
<i>Agrostis stolonifera</i> L.	4	<i>Agrostis stolonifera</i>	1
<i>Allionia incarnata</i> L.	1		
<i>Allium</i> L.	7		
<i>Ambrosia acanthicarpa</i> Hook.	1		
<i>Amelanchier alnifolia</i> (Nutt.) Nutt. ex M. Roemer	17	<i>Amelanchier alnifolia</i>	11
<i>Amelanchier utahensis</i> Koehne	139	<i>Amelanchier utahensis</i>	141
<i>Andropogon gerardii</i> Vitman	1	<i>Andropogon gerardii</i>	3
<i>Antennaria dimorpha</i> (Nutt.) Torr. & Gray	6		
<i>Antennaria</i> Gaertn.	8		
<i>Apocynum androsaemifolium</i> L.	1	<i>Apocynum androsaemifolium</i>	2
<i>Aquilegia</i> L.	2		
<i>Arabis holboellii</i> Hornem.	9		
<i>Arabis</i> L.	19	<i>Arabis hirsuta + perennans</i>	124
<i>Arabis perennans</i> S. Wats.	1	<i>Arabis perennans</i>	124
<i>Arctostaphylos patula</i> Greene	94	<i>Arctostaphylos patula</i>	94
<i>Arctostaphylos pungens</i> Kunth	12		
<i>Arenaria fendleri</i> Gray	26	<i>Arenaria fendleri</i>	23
<i>Arenaria</i> L.	3		
<i>Arenaria macradenia</i> S. Wats.	15	<i>Arenaria macradenia</i>	46

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Aristida purpurascens</i> Poir.	2		
<i>Aristida purpurea</i> Nutt.	19	<i>Aristida purpurea</i>	26
<i>Arnica</i> L.	1		
<i>Artemisia campestris</i> L.	16	<i>Artemisia campestris</i>	14
<i>Artemisia dracunculus</i> L.	13	<i>Artemisia dracunculus</i>	12
<i>Artemisia dracunculus</i> ssp. <i>dracunculus</i> L.	1	<i>Artemisia dracunculus</i>	12
<i>Artemisia filifolia</i> Torr.	11	<i>Artemisia filifolia</i>	3
<i>Artemisia ludoviciana</i> Nutt.	52	<i>Artemisia ludoviciana</i>	51
<i>Artemisia nova</i> A. Nels.	24	<i>Artemisia arbuscula</i>	4
<i>Artemisia tridentata</i> Nutt.	70	<i>Artemisia tridentata</i>	55
<i>Artemisia tridentata</i> ssp. <i>tridentata</i> Nutt.	2	<i>Artemisia tridentata</i>	55
<i>Artemisia tridentata</i> ssp. <i>vaseyana</i> (Rydb.) Beetle	9	<i>Artemisia tridentata</i>	55
<i>Asclepias subverticillata</i> (Gray) Vail	2	<i>Asclepias subverticillata</i>	9
<i>Asplenium</i> L.	1		
<i>Aster ascendens</i> Lindl.	1		
<i>Aster glaucodes</i> Blake	1	<i>Aster glaucodes</i>	6
<i>Aster</i> L.	9		
<i>Astragalus flavus</i> Nutt.	2	<i>Astragalus flavus</i>	1
<i>Astragalus</i> L.	33		
<i>Astragalus subcinereus</i> Gray	2	<i>Astragalus subcinereus</i>	1
<i>Atriplex canescens</i> (Pursh) Nutt.	12	<i>Atriplex canescens</i>	10
<i>Atriplex confertifolia</i> (Torr. & Frem.) S. Wats.	2	<i>Atriplex confertifolia</i>	2
<i>Atriplex</i> L.	1		
<i>Baccharis emoryi</i> Gray	9	<i>Baccharis salicina</i>	3
<i>Baccharis</i> L.	2		
<i>Baccharis salicifolia</i> (Ruiz & Pavon) Pers.	4	<i>Baccharis salicina</i>	3
<i>Baileya multiradiata</i> Harvey & Gray ex Gray	1	<i>Baileya multiradiata</i>	8
<i>Balsamorhiza sagittata</i> (Pursh) Nutt.	10	<i>Balsamorhiza sagittata</i>	8
<i>Betula occidentalis</i> Hook.	7	<i>Betula occidentalis</i>	2
<i>Bouteloua barbata</i> Lag.	1		
<i>Bouteloua gracilis</i> (Willd. ex Kunth) Lag. ex Griffiths	34	<i>Bouteloua gracilis</i>	19
<i>Bouteloua</i> Lag.	1		
<i>Brassica</i> L.	1		
<i>Brickellia atractyloides</i> Gray	3		
<i>Brickellia californica</i> (Torr. & Gray) Gray	3	<i>Brickellia californica</i>	4
<i>Brickellia</i> Ell.	9		
<i>Brickellia grandiflora</i> (Hook.) Nutt.	5	<i>Brickellia grandiflora</i>	10
<i>Brickellia longifolia</i> S. Wats.	7		
<i>Brickellia microphylla</i> (Nutt.) Gray	2	<i>Brickellia microphylla</i>	5
<i>Bromus anomalus</i> Rupr. ex Fourn.	8	<i>Bromus anomalus</i>	5
<i>Bromus diandrus</i> Roth	16	<i>Bromus diandrus</i>	10
<i>Bromus inermis</i> Leyss.	14		
<i>Bromus</i> L.	3		
<i>Bromus rubens</i> L.	6	<i>Bromus rubens</i>	43
<i>Bromus tectorum</i> L.	99	<i>Bromus tectorum</i>	130
<i>Calamagrostis</i> Adans.	1	<i>Calamagrostis scopulorum</i>	1
<i>Calochortus nuttallii</i> Torr. & Gray	8	<i>Calochortus nuttallii</i>	23

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Calochortus</i> Pursh	3		
<i>Carex geyeri</i> Boott	2	<i>Carex</i> sp.	7
<i>Carex</i> L.	19	<i>Carex</i> sp.+ <i>aurea</i> + <i>hystericina</i> + <i>occidentalis</i> + <i>rossii</i>	75
<i>Carex</i> L. (<i>occidentalis</i> ?)	1	<i>Carex occidentalis</i>	1
<i>Carex</i> L. (<i>rossii</i> ?)	1	<i>Carex rossii</i>	67
<i>Carex microptera</i> Mackenzie	5	<i>Carex</i> sp.	7
<i>Carex nebrascensis</i> Dewey	2	<i>Carex</i> sp.	7
<i>Carex occidentalis</i> Bailey	8	<i>Carex occidentalis</i>	1
<i>Carex rossii</i> Boott	11	<i>Carex rossii</i>	67
<i>Carex utriculata</i> Boott	2	<i>Carex</i> sp.	7
<i>Castilleja applegatei</i> ssp. <i>martinii</i> (Abrams) Chuang & Heckard	3	<i>Castilleja chromosa</i>	2
<i>Castilleja flava</i> var. <i>flava</i> S. Wats.	1		
<i>Castilleja linariifolia</i> Benth.	8	<i>Castilleja linariifolia</i>	7
<i>Castilleja</i> Mutis ex L. f.	8		
<i>Ceanothus fendleri</i> Gray	5		
<i>Ceanothus</i> L.	1		
<i>Ceanothus martinii</i> M.E. Jones	2	<i>Ceanothus martinii</i>	4
<i>Celtis laevigata</i> var. <i>reticulata</i> (Torr.) L. Benson	6	<i>Celtis reticulata</i>	2
<i>Cenchrus longispinus</i> (Hack.) Fern.	1	<i>Cenchrus longispinus</i>	2
<i>Cercocarpus intricatus</i> S. Wats.	23	<i>Cercocarpus intricatus</i>	40
<i>Cercocarpus ledifolius</i> Nutt.	2		
<i>Cercocarpus montanus</i> Raf.	46	<i>Cercocarpus montanus</i>	33
<i>Chaenactis</i> DC.	2	<i>Chaenactis douglasii</i> + <i>stevioides</i>	45
<i>Chaenactis</i> DC. (<i>douglasii</i> or <i>stevioides</i> ??)	1	<i>Chaenactis douglasii</i> + <i>stevioides</i>	45
<i>Chaenactis douglasii</i> (Hook.) Hook. & Arn.	9	<i>Chaenactis douglasii</i>	44
<i>Chaenactis stevioides</i> Hook. & Arn.	1	<i>Chaenactis stevioides</i>	1
<i>Chaetopappa ericoides</i> (Torr.) Nesom	2	<i>Chaetopappa ericoides</i>	1
<i>Chamaesyce albomarginata</i> (Torr. & Gray) Small	1	<i>Chamaesyce albomarginata</i>	23
<i>Chamaesyce fendleri</i> (Torr. & Gray) Small	8	<i>Chamaesyce fendleri</i>	3
<i>Chamaesyce glyptosperma</i> (Engelm.) Small	4		
<i>Chamaesyce parryi</i> (Engelm.) Rydb.	1		
<i>Chenopodium album</i> L.	2		
<i>Chenopodium fremontii</i> S. Wats.	4	<i>Chenopodium fremontii</i>	3
<i>Chenopodium</i> L.	1		
<i>Chimaphila umbellata</i> (L.) W. Bart.	1	<i>Chimaphila umbellata</i>	1
<i>Chrysanthmnus depressus</i> Nutt.	3	<i>Chrysanthmnus depressus</i>	5
<i>Chrysanthmnus</i> Nutt.	1		
<i>Chrysanthmnus viscidiflorus</i> (Hook.) Nutt.	15	<i>Chrysanthmnus viscidiflorus</i>	6
<i>Chrysanthmnus viscidiflorus</i> ssp. <i>axillaris</i> (Keck) L.C. Anders.	1	<i>Chrysanthmnus viscidiflorus</i>	6
<i>Cirsium arizonicum</i> (Gray) Petrak	1	<i>Cirsium arizonicum</i>	16
<i>Cirsium calcareum</i> (M.E. Jones) Woot. & Standl.	2		
<i>Cirsium</i> P. Mill.	18		
<i>Cirsium vulgare</i> (Savi) Ten.	1		
<i>Cirsium wheeleri</i> (Gray) Petrak	7	<i>Cirsium wheeleri</i>	8
<i>Claytonia perfoliata</i> ssp. <i>perfoliata</i> var. <i>nubigena</i> (Greene) Poelln.	4	<i>Montia perfoliata</i>	3
<i>Clematis columbiana</i> (Nutt.) Torr. & Gray	2	<i>Clematis columbiana</i>	3
<i>Clematis</i> L.	1		

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Clematis ligusticifolia</i> Nutt.	9		
<i>Coleogyne ramosissima</i> Torr.	12	<i>Coleogyne ramosissima</i>	13
<i>Collinsia parviflora</i> Lindl.	1	<i>Collinsia parviflora</i>	2
<i>Comandra umbellata</i> (L.) Nutt.	38	<i>Comandra umbellata</i>	35
<i>Comandra umbellata</i> ssp. <i>pallida</i> (A. DC.) Piehl	3	<i>Comandra umbellata</i>	35
<i>Convolvulus arvensis</i> L.	2	<i>Convolvulus arvensis</i>	1
<i>Cordylanthus kingii</i> S. Wats.	2	<i>Cordylanthus kingii</i>	19
<i>Cordylanthus parviflorus</i> (Ferris) Wiggins	4		
<i>Crepis intermedia</i> Gray	1		
<i>Cryptantha humilis</i> (Gray) Payson	2	<i>Cryptantha humilis</i>	1
<i>Cryptantha</i> Lehm. ex G. Don	15		
<i>Cupressus arizonica</i> Greene	1		
<i>Dactylis glomerata</i> L.	3	<i>Dactylis glomerata</i>	2
<i>Dalea searlsiae</i> (Gray) Barneby	3	<i>Dalea searlsiae</i>	13
<i>Datura</i> L.	1		
<i>Datura wrightii</i> Regel	5	<i>Datura wrightii</i>	3
<i>Descurainia pinnata</i> (Walt.) Britt.	1	<i>Descurainia pinnata</i>	28
<i>Descurainia</i> Webb & Berth.	1		
<i>Distichlis spicata</i> (L.) Greene	1	<i>Distichlis spicata</i>	1
<i>Draba</i> L.	4		
<i>Dracocephalum parviflorum</i> Nutt.	1	<i>Dracocephalum parviflorum</i>	1
<i>Echinocereus</i> Engelm.	3		
<i>Echinocereus engelmannii</i> (Parry ex Engelm.) Lem.	9	<i>Echinocereus engelmannii</i>	7
<i>Echinocereus triglochidiatus</i> Engelm.	11	<i>Echinocereus triglochidiatus</i>	22
<i>Elaeagnus angustifolia</i> L.	4		
<i>Eleocharis</i> R. Br.	1		
<i>Elymus canadensis</i> L.	3	<i>Elymus canadensis</i>	4
<i>Elymus elymoides</i> (Raf.) Swezey	59	<i>Elymus elymoides</i>	58
<i>Elymus elymoides</i> ssp. <i>elymoides</i> (Raf.) Swezey	1	<i>Elymus elymoides</i>	58
<i>Elymus glaucus</i> Buckl.	1		
<i>Elymus</i> L.	10		
<i>Elymus lanceolatus</i> (Scribn. & J.G. Sm.) Gould	11	<i>Elymus smithii</i>	3
<i>Elytrigia intermedia</i> (Host) Nevski	2	<i>Elymus hispidus</i>	1
<i>Elytrigia intermedia</i> ssp. <i>intermedia</i> (Host) Nevski	1	<i>Elymus hispidus</i>	1
<i>Elytrigia repens</i> (L.) Desv. ex B.D. Jackson	6		
<i>Ephedra nevadensis</i>	1	<i>Ephedra nevadensis</i>	18
<i>Ephedra nevadensis</i> S. Wats.	18	<i>Ephedra nevadensis</i>	18
<i>Ephedra viridis</i> Coville	25	<i>Ephedra viridis</i>	16
<i>Epilobium brachycarpum</i> K. Presl	7	<i>Epilobium brachycarpum</i>	3
<i>Epilobium canum</i> ssp. <i>garrettii</i> (A. Nels.) Raven	1	<i>Zauschneria latifolia</i>	15
<i>Epilobium canum</i> ssp. <i>latifolium</i> (Hook.) Raven	3	<i>Zauschneria latifolia</i>	15
<i>Equisetum arvense</i> L.	6	<i>Equisetum arvense</i>	1
<i>Equisetum hyemale</i> L.	3	<i>Equisetum hyemale</i>	8
<i>Equisetum</i> L.	3	<i>Equisetum arvense+hymenale+laevigatum</i>	8
<i>Equisetum laevigatum</i> A. Braun	6	<i>Equisetum laevigatum</i>	2
<i>Equisetum variegatum</i> Schleich. ex F. Weber & D.M.H. Mohr	4		
<i>Ericameria linearifolia</i> (DC.) Urbatsch & Wussow	4		

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Ericameria nauseosa</i> ssp. <i>nauseosa</i> var. <i>nauseosa</i>	43	<i>Chrysanthus nauseosus</i>	39
<i>Ericameria parryi</i> var. <i>parryi</i> (Gray) Nesom	1	<i>Chrysanthus parryi</i>	1
<i>Erigeron argentatus</i> Gray	1		
<i>Erigeron concinnus</i> var. <i>concinnus</i> (Hook. & Arn.) Torr. & Gray	1	<i>Erigeron pumilus</i>	1
<i>Erigeron divergens</i> Torr. & Gray	2	<i>Erigeron divergens</i>	17
<i>Erigeron</i> L.	37		
<i>Erigeron pumilus</i> Nutt.	1	<i>Erigeron pumilus</i>	1
<i>Erigeron speciosus</i> (Lindl.) DC.	2	<i>Erigeron speciosus</i>	6
<i>Erigeron speciosus</i> var. <i>macranthus</i> (Nutt.) Cronq.	1	<i>Erigeron speciosus</i>	6
<i>Erigeron utahensis</i> Gray	6	<i>Erigeron utahensis</i>	41
<i>Eriogonum corymbosum</i> Benth.	1	<i>Eriogonum corymbosum</i>	1
<i>Eriogonum fasciculatum</i> Benth.	1	<i>Eriogonum fasciculatum</i>	3
<i>Eriogonum flavum</i> Nutt.	1		
<i>Eriogonum inflatum</i> Torr. & Frem.	3	<i>Eriogonum inflatum</i>	3
<i>Eriogonum</i> Michx.	23		
<i>Eriogonum microthecum</i> Nutt.	5	<i>Eriogonum microthecum</i>	15
<i>Eriogonum ovalifolium</i> Nutt.	1		
<i>Eriogonum panguicense</i> (M.E. Jones) Reveal	2		
<i>Eriogonum racemosum</i> Nutt.	21	<i>Eriogonum racemosum</i>	59
<i>Eriogonum umbellatum</i> Torr.	27	<i>Eriogonum umbellatum</i>	21
<i>Eriogonum</i> 2 Michx.	2		
<i>Erioneuron pulchellum</i> (Kunth) Tateoka	1		
<i>Erodium cicutarium</i> (L.) L'Her. ex Ait.	6	<i>Erodium cicutarium</i>	10
<i>Erysimum capitatum</i> var. <i>argillosum</i> (Greene) R.J. Davis	8	<i>Erysimum asperum</i>	64
<i>Erysimum capitatum</i> var. <i>capitatum</i> (Dougl. ex Hook.) Greene	1	<i>Erysimum asperum</i>	64
<i>Euphorbia brachycera</i> Engelm.	1	<i>Euphorbia brachycera</i>	5
<i>Euphorbia</i> L.	14		
<i>Fendlerella utahensis</i> (S. Wats.) Heller	2	<i>Fendlerella utahensis</i>	1
<i>Festuca</i> L.	2		
<i>Festuca occidentalis</i> Hook.	1		
<i>Frasera speciosa</i> Dougl. ex Griseb.	11	<i>Swertia radiate</i>	11
<i>Fraxinus anomala</i> Torr. ex S. Wats.	34	<i>Fraxinus anomala</i>	45
<i>Fraxinus velutina</i> Torr.	9	<i>Fraxinus velutina</i>	6
<i>Gaillardia</i> Foug.	1		
<i>Galium aparine</i> L.	1	<i>Galium aparine</i>	6
<i>Galium</i> L.	6		
<i>Gayophytum</i> A. Juss.	1	<i>Gayophytum</i> sp.	14
<i>Geraea canescens</i> Torr. & Gray	1		
<i>Geranium caespitosum</i> James	1		
<i>Geranium</i> L.	1		
<i>Gilia Ruiz & Pavon</i>	1		
<i>Grayia spinosa</i> (Hook.) Moq.	2		
<i>Grindelia squarrosa</i> (Pursh) Dunal	1	<i>Grindelia squarrosa</i>	1
<i>Gutierrezia microcephala</i> (DC.) Gray	21	<i>Gutierrezia</i> sp.	94
<i>Gutierrezia sarothrae</i> (Pursh) Britt. & Rusby	81	<i>Gutierrezia</i> sp.	94
<i>Helianthella uniflora</i> (Nutt.) Torr. & Gray	5		
<i>Helianthus petiolaris</i> Nutt.	3		

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Helimeris multiflora</i> var. <i>multiflora</i> Nutt.	9	<i>Viguiera multiflora</i>	12
<i>Hesperodoria scopularum</i> (M.E. Jones) Greene	3	<i>Haplopappus scopulorum</i>	42
<i>Hesperostipa comata</i> ssp. <i>comata</i> (Trin. & Rupr.) Barkworth	34	<i>Stipa comata</i>	39
<i>Heterotheca villosa</i> (Pursh) Shinners	69	<i>Chrysopsis villosa</i>	74
<i>Heterotheca viscosa</i> (Gray) Harms	1	<i>Chrysopsis villosa</i>	74
<i>Heuchera rubescens</i> Torr.	1	<i>Heuchera rubescens</i>	12
<i>Heuchera rubescens</i> var. <i>versicolor</i> (Greene) M.G. Stewart	2	<i>Heuchera rubescens</i>	12
<i>Holodiscus dumosus</i> (Nutt. ex Hook.) Heller	2	<i>Holodiscus dumosus</i>	21
<i>Hordeum brachyantherum</i> Nevski	1		
<i>Hydrophyllum occidentale</i> (S. Wats.) Gray	3	<i>Hydrophyllum occidentale</i>	2
<i>Hymenopappus filifolius</i> Hook.	22	<i>Hymenopappus filifolius</i>	11
<i>Hymenopappus</i> L'Her.	1	<i>Hymenopappus filifolius</i>	11
<i>Hypochaeris radicata</i> L.	1		
<i>Ipomoea purpurea</i> (L.) Roth	1		
<i>Ipomopsis aggregata</i> (Pursh) V. Grant	3	<i>Ipomopsis aggregate</i>	5
<i>Ipomopsis arizonica</i> (Greene) Wherry	1	<i>Ipomopsis aggregata</i>	5
<i>Ipomopsis congesta</i> ssp. <i>congesta</i> (Hook.) V. Grant	5	<i>Ipomopsis congesta</i>	14
<i>Ipomopsis congesta</i> ssp. <i>frutescens</i> (Rydb.) Day	1	<i>Ipomopsis congesta</i>	14
<i>Ivesia sabulosa</i> (M.E. Jones) Keck	3	<i>Ivesia sabulosa</i>	7
<i>Juncus balticus</i> Willd.	6	<i>Juncus arcticus</i>	3
<i>Juncus ensifolius</i> Wikstr.	2	<i>Juncus ensifolius</i>	2
<i>Juncus</i> L.	3	<i>Juncus arcticus+ensifolius+tenuis</i>	4
<i>Juncus longistylis</i> Torr.	3		
<i>Juncus tenuis</i> Willd.	3	<i>Juncus tenuis</i>	1
<i>Juncus torreyi</i> Coville	1		
<i>Juniperus</i> L.	1		
<i>Juniperus osteosperma</i> (Torr.) Little	119	<i>Juniperus osteosperma</i>	115
<i>Juniperus scopulorum</i> Sarg.	50	<i>Juniperus scopulorum</i>	21
<i>Kelloggia galionoides</i> Torr.	4		
<i>Koeleria macrantha</i> (Ledeb.) J.A. Schultes	6	<i>Koeleria macrantha</i>	1
<i>Krameria</i> L.	1		
<i>Lactuca</i> L.	3		
<i>Lactuca serriola</i> L.	3	<i>Lactuca serriola</i>	13
<i>Lathyrus brachycalyx</i> Rydb.	1	<i>Lathyrus brachycalyx</i>	9
<i>Lathyrus</i> L.	3		
<i>Lathyrus</i> L. ? <i>Vicia</i> ?	1		
<i>Lepidium</i> L.	3		
<i>Leptodactylon pungens</i> (Torr.) Torr. ex Nutt.	1	<i>Leptodactylon pungens</i>	8
<i>Leptodactylon watsonii</i> (Gray) Rydb.	1	<i>Leptodactylon watsonii</i>	10
<i>Lesquerella</i> S. Wats.	1		
<i>Leymus cinereus</i> (Scribn. & Merr.) A. Love	1		
<i>Linanthus nuttallii</i> ssp. <i>nuttallii</i> (Gray) Greene ex Milliken	1	<i>Linanthastrum nuttallii</i>	19
<i>Linaria dalmatica</i> (L.) P. Mill.	2		
<i>Linum</i> L.	4		
<i>Linum perenne</i> L.	4	<i>Linum perenne</i>	6
<i>Lomatium graveolens</i> (S. Wats.) Dorn & Hartman	1	<i>Lomatium graveolens</i>	1
<i>Lomatium</i> Raf.	3		

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Lotus</i> L.	5		
<i>Lotus rigidus</i> (Benth.) Greene	2		
<i>Lotus utahensis</i> Ottley	26	<i>Lotus utahensis</i>	12
<i>Lupidium</i> species	1		
<i>Lupinus argenteus</i> Pursh	15	<i>Lupinus argenteus</i>	1
<i>Lupinus concinnus</i> J.G. Agardh	1	<i>Lupinus concinnus</i>	1
<i>Lupinus</i> L.	18		
<i>Lupinus sericeus</i> Pursh	3	<i>Lupinus sericeus</i>	9
<i>Luzula campestris</i> (L.) DC.	1		
<i>Luzula parviflora</i> (Ehrh.) Desv.	4		
<i>Lycium</i> L.	1		
<i>Lycium pallidum</i> Miers	4	<i>Lycium pallidum</i>	3
<i>Lygodesmia grandiflora</i> (Nutt.) Torr. & Gray	1	<i>Lygodesmia grandiflora</i>	2
<i>Lygodesmia juncea</i> (Pursh) D. Don ex Hook.	1		
<i>Machaeranthera canescens</i> (Pursh) Gray	21	<i>Machaeranthera canescens</i>	40
<i>Machaeranthera gracilis</i> (Nutt.) Shinners	2	<i>Haplopappus gracilis</i>	4
<i>Machaeranthera</i> Nees	6		
<i>Mahonia repens</i> (Lindl.) G. Don	40	<i>Mahonia repens</i>	29
<i>Maianthemum racemosum</i> ssp. <i>racemosum</i> (L.) Link	6	<i>Smilacina racemosa</i>	6
<i>Maianthemum stellatum</i> (L.) Link	9	<i>Smilacina stellata</i>	3
<i>Medicago lupulina</i> L.	2	<i>Medicago lupulina</i>	1
<i>Melilotus officinalis</i> (L.) Lam.	16	<i>Melilotus officinalis</i>	1
<i>Mentha arvensis</i> L.	3		
<i>Mentha X piperita</i> L. (pro sp.)	1		
<i>Mertensia arizonica</i> Greene	10	<i>Mertensia arizonica</i>	7
<i>Mertensia</i> Roth	3		
<i>Mirabilis multiflora</i> (Torr.) Gray	3	<i>Mirabilis multiflora</i>	7
<i>Monardella odoratissima</i> Benth.	1	<i>Monardella odoratissima</i>	6
<i>Muhlenbergia asperifolia</i> (Nees & Meyen ex Trin.) Parodi	3	<i>Muhlenbergia asperifolia</i>	2
<i>Muhlenbergia montana</i> (Nutt.) A.S. Hitchc.	8	<i>Muhlenbergia montana</i>	2
<i>Muhlenbergia pauciflora</i> Buckl.	1		
<i>Muhlenbergia porteri</i> Scribn. ex Beal	3		
<i>Muhlenbergia racemosa</i> (Michx.) B.S.P.	2	<i>Muhlenbergia racemosa</i>	3
<i>Muhlenbergia</i> Schreb.	5		
<i>Muhlenbergia thurberi</i> Rydb.	1	<i>Muhlenbergia thurberi</i>	12
<i>Muhlenbergia wrightii</i> Vasey ex Coulter	1		
<i>Nassella viridula</i> (Trin.) Barkworth	2		
<i>Nicotiana attenuata</i> Torr. ex S. Wats.	1	<i>Nicotiana attenuata</i>	1
<i>Oenothera cespitosa</i> Nutt.	1	<i>Oenothera caespitosa</i>	9
<i>Oenothera flava</i> (A. Nels.) Garrett	1		
<i>Oenothera howardii</i> (A. Nels.) W.L. Wagner	1		
<i>Oenothera</i> L.	8		
<i>Oenothera longissima</i> Rydb.	1	<i>Oenothera longissima</i>	2
<i>Oenothera pallida</i> Lindl.	6	<i>Oenothera pallida</i>	12
<i>Opuntia basilaris</i> Engelm. & Bigelow	9		
<i>Opuntia chlorotica</i> Engelm. & Bigelow	3		
<i>Opuntia echinocarpa</i> Engelm. & Bigelow	2		

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Opuntia engelmannii</i> Salm-Dyck	1	<i>Opuntia phaeacantha</i>	9
<i>Opuntia erinacea</i> Engelm. & Bigelow ex Engelm.	5	<i>Opuntia erinacea</i>	17
<i>Opuntia macrorhiza</i> Engelm.	103	<i>Opuntia macrorhiza</i>	120
<i>Opuntia</i> P. Mill.	4		
<i>Opuntia phaeacantha</i> Engelm.	36	<i>Opuntia phaeacantha</i>	9
<i>Opuntia polyacantha</i> Haw.	4	<i>Opuntia polyacantha</i>	3
<i>Opuntia whipplei</i> Engelm. & Bigelow	6	<i>Opuntia whipplei</i>	3
<i>Osmorrhiza depauperata</i> Phil.	2		
<i>Osmorrhiza occidentalis</i> (Nutt. ex Torr. & Gray) Torr.	6	<i>Osmorrhiza occidentalis</i>	4
<i>Osmorrhiza</i> Raf.	1		
<i>Oxytropis</i> DC.	1		
<i>Pascopyrum smithii</i> (Rydb.) A. Love	10	<i>Elymus smithii</i>	3
<i>Paxistima myrsinoides</i> (Pursh) Raf.	39	<i>Pachystima myrsinoides</i>	54
<i>Penstemon caespitosus</i> Nutt. ex Gray	10		
<i>Penstemon comarrhenus</i> Gray	1	<i>Penstemon comarrhenus</i>	2
<i>Penstemon eatonii</i> Gray	14	<i>Penstemon eatonii</i>	23
<i>Penstemon humilis</i> Nutt. ex Gray	3	<i>Penstemon humilis</i>	27
<i>Penstemon latus</i> Pennell	1		
<i>Penstemon leonardii</i> Rydb.	6	<i>Penstemon higginsii</i>	5
<i>Penstemon leonardii</i> var. <i>higginsii</i> Neese	1	<i>Penstemon higginsii</i>	5
<i>Penstemon linarioides</i> Gray	9	<i>Penstemon linarioides</i>	13
<i>Penstemon pachyphyllus</i> Gray ex Rydb.	2	<i>Penstemon pachyphyllus</i>	1
<i>Penstemon palmeri</i> Gray	16	<i>Penstemon palmeri</i>	19
<i>Penstemon rostriflorus</i> Kellogg	2	<i>Penstemon rostriflorus</i>	30
<i>Penstemon</i> Schmidel	38		
<i>Penstemon tusharensis</i> N. Holmgren	1		
<i>Peraphyllum ramosissimum</i> Nutt.	10	<i>Peraphyllum ramosissimum</i>	10
<i>Petradoria pumila</i> (Nutt.) Greene	7	<i>Petradoria pumila</i>	2
<i>Petrophyton caespitosum</i> (Nutt.) Rydb.	3	<i>Petrophytum caespitosum</i>	12
<i>Phacelia heterophylla</i> Pursh	21	<i>Phacelia heterophylla</i>	42
<i>Phacelia</i> Juss.	5		
<i>Phleum pratense</i> L.	1		
<i>Phlox austromontana</i> Coville	32	<i>Phlox austromontana</i>	54
<i>Phlox caespitosa</i> Nutt.	1		
<i>Phlox hoodii</i> Richards.	6		
<i>Phlox</i> L.	7		
<i>Phlox longifolia</i> Nutt.	1	<i>Phlox longifolia</i>	11
<i>Phoradendron juniperinum</i> Engelm.	2	<i>Phoradendron juniperinum</i>	14
<i>Phragmites australis</i> (Cav.) Trin. ex Steud.	1	<i>Phragmites australis</i>	2
<i>Physalis hederifolia</i> Gray	1	<i>Physalis hederifolia</i>	8
<i>Physalis heterophylla</i> Nees	1		
<i>Physaria</i> (Nutt. ex Torr. & Gray) Gray	2		
<i>Physaria chambersii</i> Rollins	6	<i>Physaria chambersii</i>	3
<i>Physaria newberryi</i> Gray	1	<i>Physaria newberryi</i>	7
<i>Pinus edulis</i> Engelm.	48	<i>Pinus edulis</i>	53
<i>Pinus edulis</i> x <i>monophylla</i> Engelm.	4	<i>Pinus edulis+monophylla</i>	115
<i>Pinus monophylla</i> Torr. & Frem.	58	<i>Pinus monophylla</i>	64

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Pinus ponderosa</i> P. & C. Lawson	74	<i>Pinus ponderosa</i>	60
<i>Plagiobothrys tenellus</i> (Nutt. ex Hook.) Gray	1		
<i>Plantago patagonica</i> Jacq.	3	<i>Plantago patagonica</i>	13
<i>Pleuraphis jamesii</i> Torr.	26	<i>Hilaria</i> sp.	41
<i>Pluchea sericea</i> (Nutt.) Coville	2		
<i>Poa fendleriana</i> (Steud.) Vasey	155	<i>Poa fendleriana</i>	190
<i>Poa</i> L.	4		
<i>Poa pratensis</i> L.	44	<i>Poa pratensis</i>	14
<i>Poa secunda</i> J. Presl	14	<i>Poa secunda</i>	4
<i>Poa</i> 2 L.	1		
<i>Polygonum douglasii</i> Greene	8	<i>Polygonum douglasii</i>	39
<i>Polygonum</i> L.	3		
<i>Polypodium hesperium</i> Maxon	1	<i>Polypodium hesperium</i>	6
<i>Polypogon</i> Desf.	1		
<i>Populus angustifolia</i> James	1		
<i>Populus fremontii</i> S. Wats.	22	<i>Populus fremontii</i>	6
<i>Populus tremuloides</i> Michx.	11	<i>Populus tremuloides</i>	2
<i>Potentilla glandulosa</i> Lindl.	1		
<i>Prosopis glandulosa</i> Torr.	1		
<i>Prunus</i> L.	1		
<i>Prunus virginiana</i> L.	14	<i>Prunus virginiana</i>	10
<i>Pseudoroegneria spicata</i> ssp. <i>spicata</i> (Pursh) A. Love	1	<i>Elymus spicatus</i>	6
<i>Pseudostellaria jamesiana</i> (Torr.) W.A. Weber & R.L. Hartman	3	<i>Stellaria jamesiana</i>	18
<i>Pseudotsuga menziesii</i> (Mirbel) Franco	19	<i>Pseudotsuga menziesii</i>	18
<i>Psoralidium</i> Rydb.	1		
<i>Psorothamnus fremontii</i> (Torr. ex Gray) Barneby	8	<i>Psorothamnus fremontii</i>	11
<i>Psorothamnus fremontii</i> var. <i>fremontii</i> (Torr. ex Gray) Barneby	1	<i>Psorothamnus fremontii</i>	11
<i>Pteridium aquilinum</i> (L.) Kuhn	4	<i>Pteridium aquilinum</i>	3
<i>Pterospora andromedea</i> Nutt.	1	<i>Pterospora andromedea</i>	2
<i>Puccinellia distans</i> (Jacq.) Parl.	1		
<i>Purshia</i> DC. ex Poir.	1		
<i>Purshia mexicana</i> (D. Don) Henrickson	11	<i>Purshia mexicana</i>	15
<i>Purshia stansburiana</i> (Torr.) Henrickson	6	<i>Purshia mexicana</i>	15
<i>Purshia tridentata</i> (Pursh) DC.	51	<i>Purshia tridentata</i>	36
<i>Quercus gambelii</i> Nutt.	166	<i>Quercus gambelii</i>	135
<i>Quercus turbinella</i> Greene	40	<i>Quercus turbinella</i>	50
<i>Rhus aromatica</i> Ait.	9	<i>Rhus aromatica</i>	23
<i>Rhus trilobata</i> Nutt.	16	<i>Rhus aromatica</i>	23
<i>Rhus trilobata</i> var. <i>trilobata</i> Nutt.	10	<i>Rhus aromatica</i>	23
<i>Rorippa nasturtium-aquaticum</i> (L.) Hayek	1	<i>Nasturtium officinale</i>	1
<i>Rosa woodsii</i> Lindl.	19	<i>Rosa woodsii</i>	12
<i>Rumex acetosella</i> L.	2		
<i>Rumex hymenosepalus</i> Torr.	2	<i>Rumex hymenosepalus</i>	2
<i>Salix exigua</i> Nutt.	12	<i>Salix exigua</i>	3
<i>Salix gooddingii</i> Ball	2		
<i>Salix</i> L.	4		
<i>Salix lasiolepis</i> Benth.	1		

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Salix ligulifolia</i> (Ball) Ball ex Schneid.	1		
<i>Salix lucida</i> Muhl.	1		
<i>Salix lucida</i> ssp. <i>lasiandra</i> (Benth.) E. Murr.	1		
<i>Salix scouleriana</i> Barratt ex Hook.	2	<i>Salix scouleriana</i>	2
<i>Salsola kali</i> ssp. <i>tragus</i> (L.) Celak.	1		
<i>Salvia dorrii</i> (Kellogg) Abrams	5	<i>Salvia dorrii</i>	3
<i>Sambucus</i> L.	1		
<i>Schizachyrium scoparium</i> (Michx.) Nash	2		
<i>Schoenoplectus americanus</i> (Pers.) Volk. ex Schinz & R. Keller	1		
<i>Schoenoplectus tabernaemontani</i> (K.C. Gmel.) Palla	1		
<i>Scirpus</i> L.	1		
<i>Senecio eremophilus</i> Richards.	3	<i>Senecio eremophilus</i>	3
<i>Senecio integrerrimus</i> Nutt.	1		
<i>Senecio</i> L.	14		
<i>Senecio multilobatus</i> Torr. & Gray ex Gray	31	<i>Senecio multilobatus</i>	99
<i>Senecio spartioides</i> Torr. & Gray	19	<i>Senecio spartioides</i>	26
<i>Shepherdia rotundifolia</i> Parry	12	<i>Shepherdia rotundifolia</i>	17
<i>Sisymbrium altissimum</i> L.	3	<i>Sisymbrium altissimum</i>	4
<i>Sisymbrium</i> L.	1	<i>Sisymbrium altissimum+irio</i>	6
<i>Sisyrinchium demissum</i> Greene	2		
skeletonweed	1		
<i>Solanum elaeagnifolium</i> Cav.	1		
<i>Solidago</i> L.	1		
<i>Solidago velutina</i> DC.	9	<i>Solidago velutina</i>	38
<i>Sorghastrum nutans</i> (L.) Nash	2		
<i>Sphaeralcea ambigua</i> Gray	2	<i>Sphaeralcea ambigua</i>	2
<i>Sphaeralcea coccinea</i> (Nutt.) Rydb.	12		
<i>Sphaeralcea grossulariifolia</i> (Hook. & Arn.) Rydb.	4	<i>Sphaeralcea grossulariifolia</i>	14
<i>Sphaeralcea</i> St.-Hil.	6		
<i>Sphaeromeria ruthiae</i> Holmgren, Shultz & Lowrey	2	<i>Sphaeromeria ruthiae</i>	3
<i>Sporobolus cryptandrus</i> (Torr.) Gray	60	<i>Sporobolus cryptandrus</i>	50
<i>Sporobolus</i> R. Br.	1		
<i>Stanleya pinnata</i> (Pursh) Britt.	3	<i>Stanleya pinnata</i>	2
<i>Stellaria</i> L.	2		
<i>Stellaria media</i> (L.) Vill.	2		
<i>Stephanomeria exigua</i> Nutt.	1	<i>Stephanomeria exigua</i>	1
<i>Stephanomeria</i> Nutt.	2		
<i>Stephanomeria tenuifolia</i> (Raf.) Hall	1	<i>Stephanomeria tenuifolia</i>	51
<i>Stipa</i> L.	6		
<i>Stipa</i> L. (comata or lettermanii)	1		
<i>Streptanthus cordatus</i> Nutt.	3	<i>Streptanthus cordatus</i>	38
<i>Symporicarpos longiflorus</i> Gray	4		
<i>Symporicarpos oreophilus</i> Gray	79	<i>Symporicarpos oreophilus</i>	58
<i>Tamarix ramosissima</i> Ledeb.	10	<i>Tamarix chinensis</i>	5
<i>Taraxacum officinale</i> G.H. Weber ex Wiggers	7	<i>Taraxacum officinale</i>	11
<i>Tetradymia axillaris</i> A. Nels.	1		
<i>Tetradymia canescens</i> DC.	10	<i>Tetradymia canescens</i>	9

USGS-NPS Name (Kartesz 1999)	#Occ	BYU-RMRS Name (Welsh et al. 2003)	#Occ
<i>Thalictrum fendleri</i> Engelm. ex Gray	15	<i>Thalictrum fendleri</i>	12
<i>Thalictrum</i> L.	1		
<i>Thermopsis montana</i> Nutt.	2	<i>Thermopsis montana</i>	4
<i>Townsendia</i> Hook.	2		
<i>Toxicodendron rydbergii</i> (Small ex Rydb.) Greene	2	<i>Toxicodendron rydbergii</i>	2
<i>Tradescantia occidentalis</i> (Britt.) Smyth	14	<i>Tradescantia occidentalis</i>	22
<i>Tragia ramosa</i> Torr.	1		
<i>Tragopogon dubius</i> Scop.	22	<i>Tragopogon dubius</i>	8
<i>Tribulus terrestris</i> L.	2	<i>Tribulus terrestris</i>	2
<i>Trifolium gymnocarpon</i> Nutt.	1		
<i>Trifolium</i> L.	3		
<i>Trifolium longipes</i> Nutt.	3	<i>Trifolium longipes</i>	10
<i>Triticum aestivum</i> L.	1		
<i>Typha angustifolia</i> L.	2	<i>Typha latifolia</i>	1
<i>Typha domingensis</i> Pers.	1	<i>Typha latifolia</i>	1
<i>Verbascum thapsus</i> L.	14	<i>Verbascum thapsus</i>	5
<i>Verbena bracteata</i> Lag. & Rodr.	3		
<i>Vicia americana</i> Muhl. ex Willd.	50	<i>Vicia americana</i>	43
<i>Vicia</i> L.	5		
<i>Viola charlestonensis</i> M.S. Baker & J.C. Clausen ex Clokey	1	<i>Viola purpurea</i>	10
<i>Viola</i> L.	1		
<i>Vitis arizonica</i> Engelm.	4	<i>Vitis arizonica</i>	1
<i>Vulpia octoflora</i> var. <i>octoflora</i> (Walt.) Rydb.	4	<i>Festuca octoflora</i>	74
<i>Wyethia arizonica</i> Gray	2	<i>Wyethia arizonica</i>	5
<i>Yucca baccata</i> Torr.	20	<i>Yucca baccata</i>	17
<i>Yucca elata</i> var. <i>utahensis</i> (McKelvey) Reveal	79	<i>Yucca angustissima</i>	41

APPENDIX 2.2

Vascular plant species/taxa recorded by the 1987-89 vegetation survey of Zion National Park, Utah (Harper 1993), and their affinities to coalition species groups. #Occ.=number of plot occurrences out of 288 plots total. Affinities are mean pairwise association values (in units of the phi coefficient) between a species and members of a coalition group (see also Table 2.2). Green and red shadings for low and high affinity values, respectively, are scaled separately for each column. Nomenclature follows Welsh et al. (2003) and names are cross-walked with Cogan et al. (2004) and Alexander and Fertig (2009) (see footnotes).

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Abies concolor</i>	Pinaceae	19	-0.0226	-0.0893	-0.0770	-0.0559	0.0865	0.0603	0.0229	0.1859	0.2236	0.0410
<i>Abronia fragrans</i>	Nyctaginaceae	20	-0.0448	-0.0051	0.0253	0.2521	0.0395	-0.0448	-0.0382	-0.0325	-0.0349	-0.0360
<i>Acer glabrum*</i>	Aceraceae	6	-0.0269	-0.0484	-0.0117	-0.0307	0.1020	0.0035	-0.0116	0.0185	0.1461	0.0090
<i>Acer grandidentatum</i>	Aceraceae	29	-0.0556	-0.0988	-0.0451	-0.0489	0.0587	0.0312	0.0003	0.1139	0.2614	0.0429
<i>Acer negundo</i>	Aceraceae	18	-0.0226	-0.0711	-0.0929	-0.0678	0.0736	-0.0447	-0.0579	0.0499	0.3163	0.2513
<i>Achillea millefolium</i>	Compositae	6	-0.0003	-0.0443	-0.0386	0.0190	0.0046	-0.0178	0.0136	0.2048	0.0272	0.0380
<i>Actaea rubra*</i>	Ranunculaceae	2	-0.0154	-0.0290	-0.0467	-0.0189	0.0224	0.0006	-0.0186	0.0479	0.2531	0.0817
<i>Adiantum capillus-veneris*</i>	Polypodiaceae	2	-0.0154	-0.0290	-0.0466	-0.0340	-0.0028	-0.0352	-0.0308	0.0132	0.1540	-0.0025
<i>Adiantum pedatum*</i>	Polypodiaceae	1	-0.0109	-0.0204	-0.0291	-0.0027	0.0370	0.0140	-0.0138	0.0581	0.2611	0.1104
<i>Agastache urticifolia</i>	Labiatae	1	-0.0109	-0.0133	-0.0368	-0.0149	-0.0163	-0.0150	-0.0040	0.2240	-0.0098	0.0533
<i>Agoseris aurantiaca*</i>	Compositae	1	-0.0109	-0.0071	-0.0067	-0.0149	-0.0163	-0.0051	-0.0059	0.0257	-0.0012	0.0067
<i>Agoseris glauca*</i>	Compositae	1	0.0196	0.0051	0.0044	-0.0094	-0.0056	-0.0150	-0.0138	-0.0032	-0.0098	0.0094
<i>Agoseris retrorsa*</i>	Compositae	1	0.0537	-0.0144	-0.0290	-0.0073	0.0010	-0.0033	0.0053	0.0870	-0.0098	0.0390
<i>Agropyron cristatum€</i>	Gramineae	1	0.0689	-0.0204	-0.0142	-0.0112	-0.0099	-0.0150	0.0458	0.0565	-0.0012	-0.0085
<i>Agrostis exarata</i>	Gramineae	3	-0.0189	-0.0164	-0.0469	-0.0211	0.0083	-0.0207	-0.0332	0.0416	0.1570	0.3453
<i>Agrostis scabra*</i>	Gramineae	1	-0.0109	-0.0204	-0.0291	-0.0027	0.0370	0.0140	-0.0138	0.0581	0.2611	0.1104
<i>Agrostis stolonifera€</i>	Gramineae	2	-0.0109	-0.0183	-0.0329	-0.0240	-0.0163	-0.0248	-0.0217	0.0068	0.0093	0.2268
<i>Allium acuminatum*</i>	Liliaceae	9	-0.0113	-0.0403	-0.0317	0.0492	-0.0213	-0.0161	0.0051	0.0793	-0.0269	0.0119
<i>Allium macropetalum*†</i>	Liliaceae	1	-0.0109	-0.0156	-0.0064	0.0599	0.1228	-0.0033	-0.0045	-0.0055	-0.0098	-0.0085
<i>Allium nevadense*</i>	Liliaceae	1	-0.0109	-0.0204	-0.0216	0.0856	-0.0029	-0.0024	-0.0019	-0.0127	-0.0098	-0.0085
<i>Allium sp.</i>	Liliaceae	2	0.0062	-0.0075	-0.0014	0.0303	0.1292	-0.0282	-0.0252	-0.0061	0.0107	0.0006
<i>Amelanchier alnifolia</i>	Rosaceae	11	-0.0245	-0.0668	-0.0809	-0.0447	0.0231	0.0570	0.0507	0.2711	0.1498	0.0627

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Amelanchier utahensis</i>	Rosaceae	141	-0.0900	-0.0968	0.1661	0.0340	-0.0081	0.1990	0.1598	-0.0446	-0.0817	-0.1249
<i>Andropogon gerardii</i>	Gramineae	3	0.0543	-0.0164	-0.0358	-0.0027	0.0669	-0.0177	-0.0232	-0.0220	0.0281	0.1007
<i>Androstaphium breviflorum*</i>	Liliaceae	18	0.1286	0.0230	0.0268	0.0390	-0.0266	-0.0414	0.0075	-0.0162	-0.0299	-0.0159
<i>Angelica pinnata*</i>	Umbelliferae	1	-0.0109	-0.0204	-0.0291	-0.0027	0.0370	0.0140	-0.0138	0.0581	0.2611	0.1104
<i>Antennaria microphylla*</i>	Compositae	1	-0.0109	-0.0159	-0.0063	0.0034	-0.0035	-0.0150	-0.0138	-0.0127	-0.0098	-0.0085
<i>Antennaria parvifolia*</i>	Compositae	2	-0.0154	0.0083	0.0049	0.0489	0.0019	-0.0122	-0.0054	-0.0136	-0.0139	-0.0121
<i>Antennaria rosulata*‡</i>	Compositae	3	-0.0189	-0.0276	-0.0021	-0.0057	0.0238	0.0195	0.0148	0.0635	0.1447	0.0540
<i>Apocynum androsaemifolium</i>	Apocynaceae	2	-0.0154	-0.0290	-0.0414	-0.0189	0.0270	0.0152	-0.0030	0.0476	0.2110	0.0817
<i>Aquilegia chrysantha*</i>	Ranunculaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	0.0179	-0.0085
<i>Aquilegia formosa*</i>	Ranunculaceae	3	-0.0189	-0.0355	-0.0509	-0.0269	0.0919	-0.0250	-0.0232	-0.0163	0.0585	-0.0070
<i>Aquilegia</i> sp.	Ranunculaceae	2	-0.0154	-0.0290	-0.0466	-0.0286	0.0010	-0.0147	-0.0238	0.0385	0.1668	-0.0025
<i>Arabis hirsuta*‡</i>	Cruciferae	1	-0.0109	-0.0204	0.0010	-0.0035	-0.0109	0.0065	0.0428	0.0474	-0.0098	-0.0085
<i>Arabis perennans</i>	Cruciferae	124	-0.0953	0.0295	0.2379	0.0173	0.0178	0.0472	0.0240	-0.0499	-0.0301	-0.0931
<i>Aralia racemosa*</i>	Araliaceae	3	-0.0189	-0.0331	-0.0507	-0.0417	-0.0017	-0.0187	-0.0151	0.0313	0.2012	0.0469
<i>Arceuthobium abietinum*‡</i>	Viscaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0054	-0.0132	-0.0124	0.0096	0.0963	0.0050
<i>Arceuthobium divaricatum*</i>	Viscaceae	5	-0.0245	-0.0183	0.1003	0.0123	-0.0047	0.0106	0.0083	-0.0202	-0.0220	-0.0192
<i>Arceuthobium vaginatum*</i>	Viscaceae	1	-0.0109	-0.0204	-0.0368	-0.0097	-0.0109	0.0411	0.0110	0.0214	-0.0016	-0.0085
<i>Arctostaphylos patula</i>	Ericaceae	94	-0.1013	-0.0593	0.1132	0.2147	0.0767	0.0893	0.0301	-0.0959	-0.0837	-0.0949
<i>Arenaria fendleri</i>	Caryophyllaceae	23	-0.0544	-0.0513	-0.0408	0.1880	0.2407	-0.0605	-0.0522	-0.0512	-0.0203	-0.0427
<i>Arenaria macradenia</i>	Caryophyllaceae	46	-0.0804	-0.0607	0.1271	0.0990	0.1502	-0.0052	-0.0069	-0.0401	-0.0037	-0.0588
<i>Arenaria pusilla*</i>	Caryophyllaceae	1	-0.0109	0.0189	-0.0059	0.0579	0.0148	-0.0132	-0.0124	-0.0127	-0.0098	-0.0085
<i>Argemone munita*</i>	Papaveraceae	1	0.0512	-0.0001	-0.0138	-0.0240	-0.0163	-0.0150	-0.0138	-0.0058	-0.0098	-0.0085
<i>Aristida purpurea*</i>	Gramineae	26	0.0216	0.1291	0.0011	-0.0051	-0.0026	-0.0986	-0.0888	-0.0646	-0.0393	0.0031
<i>Arnica cordifolia*</i>	Compositae	1	-0.0109	-0.0204	-0.0142	-0.0097	-0.0041	0.0778	0.0034	-0.0004	-0.0098	-0.0085
<i>Artemisia arbuscula¹</i>	Compositae	4	0.0383	-0.0411	-0.0399	-0.0278	-0.0146	0.0007	0.0998	0.1245	0.0005	-0.0095
<i>Artemisia campestris</i>	Compositae	14	-0.0334	-0.0471	-0.0323	0.2341	-0.0044	-0.0462	-0.0361	-0.0341	-0.0315	-0.0327
<i>Artemisia dracunculus</i>	Compositae	12	0.0695	-0.0227	-0.0022	-0.0069	-0.0291	-0.0208	-0.0178	-0.0225	-0.0260	-0.0138
<i>Artemisia filifolia</i>	Compositae	3	-0.0189	0.0497	-0.0347	-0.0349	-0.0250	-0.0431	-0.0308	-0.0220	-0.0170	-0.0016
<i>Artemisia frigida*</i>	Compositae	1	-0.0109	-0.0204	-0.0216	-0.0115	0.0069	0.0247	0.0354	0.0580	-0.0016	-0.0085
<i>Artemisia ludoviciana</i>	Compositae	51	-0.0089	0.0007	0.1161	0.0261	0.0346	-0.0376	-0.0201	0.0195	0.0962	0.1026
<i>Artemisia</i> sp.	Compositae	2	-0.0154	-0.0198	-0.0370	-0.0290	-0.0196	-0.0352	-0.0308	0.0197	0.0312	0.4632

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Artemisia tridentata</i>	Compositae	55	0.0211	0.0237	0.0140	0.0087	-0.0941	-0.0650	-0.0307	-0.0021	-0.0710	-0.0200
<i>Asclepias subverticillata</i>	Asclepiadaceae	9	0.0355	0.0465	-0.0005	-0.0368	-0.0372	0.0539	-0.0149	-0.0385	-0.0264	-0.0193
<i>Aster glaucodes</i>	Compositae	6	-0.0269	-0.0144	0.0115	-0.0389	-0.0091	-0.0054	-0.0173	0.0135	0.1160	0.1156
<i>Aster sp.</i>	Compositae	1	0.0207	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	-0.0016	0.0906
<i>Aster spathulatus*†‡</i>	Compositae	1	0.0290	-0.0204	-0.0220	-0.0177	-0.0099	0.0355	0.0251	0.0679	0.0050	-0.0085
<i>Aster welshii*</i>	Compositae	6	-0.0269	-0.0449	-0.0609	-0.0507	-0.0144	-0.0372	-0.0375	0.0224	0.1633	0.1134
<i>Astragalus flavus</i>	Leguminosae	1	-0.0109	0.0218	-0.0287	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Astragalus lancearius*</i>	Leguminosae	1	-0.0109	-0.0084	0.0175	0.0125	0.0009	-0.0051	-0.0059	0.0109	-0.0043	-0.0085
<i>Astragalus mollissimus*</i>	Leguminosae	1	-0.0109	0.0220	-0.0290	-0.0240	-0.0163	-0.0248	-0.0098	-0.0127	-0.0098	0.0144
<i>Astragalus nuttallianus*</i>	Leguminosae	9	0.0317	0.2167	-0.0609	-0.0709	-0.0475	-0.0755	-0.0594	-0.0385	-0.0297	-0.0260
<i>Astragalus piutensis*</i>	Leguminosae	6	0.0388	-0.0222	-0.0177	-0.0238	-0.0305	0.0101	0.0300	0.0813	-0.0118	0.0003
<i>Astragalus sp.</i>	Leguminosae	27	-0.0026	0.0839	0.0152	0.0117	-0.0543	-0.0184	0.0463	-0.0225	-0.0394	0.0038
<i>Astragalus subcinereus</i>	Leguminosae	1	-0.0109	-0.0161	0.0011	-0.0175	-0.0163	0.0356	0.0140	-0.0065	-0.0098	-0.0085
<i>Astragalus zionis*</i>	Leguminosae	22	-0.0192	-0.0059	0.0676	0.0727	0.0065	0.0252	0.0815	0.0117	-0.0376	-0.0311
<i>Atriplex canescens</i>	Chenopodiaceae	10	0.2722	0.0716	-0.0649	-0.0708	-0.0447	-0.0797	-0.0634	-0.0407	-0.0314	-0.0275
<i>Atriplex confertifolia</i>	Chenopodiaceae	2	0.0099	0.0251	-0.0303	-0.0295	-0.0231	-0.0352	-0.0308	-0.0179	-0.0139	-0.0121
<i>Baccharis salicina</i>	Compositae	3	-0.0189	-0.0139	-0.0469	-0.0335	-0.0226	-0.0431	-0.0377	0.0088	0.0332	0.4594
<i>Baileya multiradiata</i>	Compositae	8	0.0310	0.2654	-0.0188	-0.0432	-0.0368	-0.0711	-0.0622	-0.0363	-0.0280	-0.0245
<i>Balsamorhiza sagittata</i>	Compositae	8	-0.0169	-0.0466	0.0013	-0.0108	-0.0278	0.1063	0.2200	0.0523	-0.0256	-0.0245
<i>Berula erecta*</i>	Umbelliferae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Betula occidentalis*</i>	Betulaceae	2	-0.0154	-0.0224	-0.0114	-0.0192	0.0012	-0.0007	-0.0086	0.0070	0.0529	0.0360
<i>Bouteloua curtipendula*</i>	Gramineae	2	-0.0154	-0.0189	-0.0342	-0.0030	0.0629	-0.0352	-0.0308	-0.0179	-0.0089	-0.0121
<i>Bouteloua eriopoda*</i>	Gramineae	5	0.0220	0.2274	-0.0295	-0.0540	-0.0316	-0.0514	-0.0453	-0.0285	-0.0220	-0.0192
<i>Bouteloua gracilis</i>	Gramineae	19	-0.0244	-0.0350	-0.0107	0.1887	-0.0281	-0.0236	0.0192	-0.0029	-0.0361	-0.0306
<i>Brickellia californica</i>	Compositae	4	-0.0066	0.0351	0.0451	-0.0051	0.0036	-0.0400	-0.0357	-0.0125	-0.0159	-0.0082
<i>Brickellia grandiflora</i>	Compositae	10	-0.0350	-0.0516	-0.0105	0.0153	0.1977	0.0232	-0.0173	0.0078	0.1723	0.0031
<i>Brickellia microphylla</i>	Compositae	5	-0.0245	0.0370	0.0673	-0.0131	0.0217	-0.0240	-0.0234	-0.0285	-0.0092	-0.0192
<i>Brickellia oblongifolia*†‡</i>	Compositae	2	-0.0154	-0.0055	-0.0153	-0.0251	0.0119	-0.0192	-0.0252	0.0050	0.0747	-0.0025
<i>Bromus anomalus</i>	Gramineae	5	0.0436	-0.0421	-0.0621	-0.0175	-0.0099	-0.0147	0.0014	0.1586	0.1089	0.0533
<i>Bromus carinatus*</i>	Gramineae	9	-0.0196	-0.0434	-0.0574	-0.0456	-0.0372	-0.0416	-0.0107	0.2714	0.0118	0.2511
<i>Bromus ciliatus*</i>	Gramineae	12	0.0293	-0.0670	-0.0665	-0.0486	0.0757	-0.0176	-0.0460	0.0473	0.2602	0.0995

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Bromus diandrus</i> €	Gramineae	10	0.0312	-0.0336	-0.0857	-0.0613	-0.0407	-0.0734	-0.0647	-0.0126	0.0071	0.3353
<i>Bromus rubens</i> €	Gramineae	43	0.0339	0.3680	0.0731	-0.0990	-0.0855	-0.1439	-0.1239	-0.0865	-0.0642	-0.0544
<i>Bromus tectorum</i> €	Gramineae	130	0.1156	0.2135	0.0710	-0.0590	-0.1214	-0.1832	-0.1512	-0.1083	-0.0873	0.0262
<i>Bromus vulgaris</i> *‡	Gramineae	1	0.0606	-0.0162	-0.0368	-0.0149	-0.0099	-0.0033	0.0053	0.0953	-0.0016	-0.0085
<i>Calamagrostis scopulorum</i>	Gramineae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	0.0179	-0.0085
<i>Calochortus flexuosus</i> *	Liliaceae	7	0.0171	0.1303	-0.0365	-0.0562	-0.0395	-0.0626	-0.0550	-0.0302	-0.0261	-0.0160
<i>Calochortus nuttallii</i>	Liliaceae	23	0.0711	0.0121	0.0399	0.0553	-0.0340	-0.0119	-0.0077	0.0074	-0.0329	-0.0153
<i>Camissonia multifluga</i> *	Onagraceae	1	-0.0109	0.0127	0.0000	-0.0240	-0.0100	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Camissonia parvula</i> *	Onagraceae	2	-0.0154	0.0040	0.0322	0.0436	-0.0006	0.0107	0.0060	-0.0112	-0.0100	-0.0121
<i>Camissonia</i> sp.*	Onagraceae	1	-0.0109	-0.0117	-0.0232	-0.0169	-0.0113	-0.0248	-0.0217	0.0263	0.0371	0.3576
<i>Cardamine cordifolia</i> *‡‡	Cruciferae	1	0.0606	-0.0204	-0.0368	-0.0169	0.0454	-0.0248	-0.0217	-0.0127	0.0547	0.0050
<i>Carex aurea</i> *	Cyperaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0099	0.0014	-0.0138	0.0403	0.0491	0.0067
<i>Carex hystericina</i> *	Cyperaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	0.0179	-0.0085
<i>Carex occidentalis</i>	Cyperaceae	1	-0.0109	-0.0204	-0.0213	0.0098	0.0029	-0.0248	-0.0217	0.0176	0.0465	0.0870
<i>Carex rossii</i>	Cyperaceae	67	-0.0335	-0.1431	-0.0482	0.0505	0.1222	0.2114	0.1553	0.1027	0.0042	-0.0554
<i>Carex</i> sp.	Cyperaceae	7	-0.0291	-0.0347	-0.0301	0.0315	0.0308	-0.0238	-0.0291	-0.0045	0.0800	0.0708
<i>Castilleja chromosa</i>	Scrophulariaceae	2	-0.0154	-0.0260	-0.0198	0.0174	-0.0022	-0.0212	-0.0196	0.0086	0.0260	0.0556
<i>Castilleja linariifolia</i>	Scrophulariaceae	7	0.0512	-0.0182	-0.0071	-0.0227	-0.0003	0.0218	0.1229	0.0230	0.0120	-0.0101
<i>Castilleja scabrida</i> *	Scrophulariaceae	40	-0.0392	-0.0544	-0.0185	0.0831	0.2903	-0.0401	-0.0609	-0.0449	0.0430	-0.0458
<i>Caulanthus cooperi</i> *‡	Cruciferae	1	0.1112	-0.0117	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Caulanthus crassicaulis</i> *	Cruciferae	1	-0.0109	-0.0154	-0.0030	-0.0240	-0.0163	0.0103	0.0039	0.0036	-0.0098	-0.0085
<i>Ceanothus martinii</i>	Rhamnaceae	4	0.0182	-0.0389	0.0003	-0.0180	-0.0174	0.1121	0.0788	0.0641	-0.0055	-0.0172
<i>Celtis reticulata</i>	Ulmaceae	2	0.0233	-0.0197	-0.0411	-0.0143	-0.0196	-0.0282	-0.0252	-0.0179	-0.0139	0.0235
<i>Cenchrus longispinus</i>	Gramineae	2	0.0017	-0.0198	-0.0425	-0.0290	-0.0196	-0.0352	-0.0308	0.0097	0.0264	0.2903
<i>Centaurium exaltatum</i> *	Gentianaceae	1	0.0196	0.0051	0.0044	-0.0094	-0.0056	-0.0150	-0.0138	-0.0032	-0.0098	0.0094
<i>Centrostegia thurberi</i> *	Polygonaceae	3	-0.0189	0.1220	-0.0116	-0.0303	-0.0221	-0.0431	-0.0263	-0.0220	-0.0170	-0.0149
<i>Cercocarpus intricatus</i>	Rosaceae	40	-0.0741	-0.0732	-0.0008	0.0948	0.3022	-0.0254	-0.0425	-0.0540	0.0241	-0.0535
<i>Cercocarpus montanus</i>	Rosaceae	33	-0.0411	-0.0845	0.0125	-0.0766	-0.0614	0.2588	0.1440	-0.0299	-0.0468	-0.0521
<i>Chaenactis douglasii</i>	Compositae	44	-0.0625	-0.0719	0.1114	0.2527	0.1140	-0.0008	0.0032	-0.0622	-0.0409	-0.0586
<i>Chaenactis stevioides</i>	Compositae	1	-0.0109	0.0466	-0.0212	-0.0177	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Chaetopappa ericoides</i>	Compositae	1	-0.0109	-0.0115	0.0011	-0.0177	-0.0163	0.1087	0.0330	-0.0065	-0.0098	-0.0085

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Chamaesyce albomarginata</i>	Euphorbiaceae	23	0.2085	0.0081	-0.0514	-0.0601	-0.0683	0.0486	0.0045	-0.0518	-0.0427	-0.0065
<i>Chamaesyce fendleri</i>	Euphorbiaceae	3	-0.0189	0.0881	0.0804	0.0042	-0.0212	-0.0075	-0.0240	-0.0220	-0.0170	-0.0149
<i>Chenopodium fremontii</i>	Chenopodiaceae	3	-0.0189	-0.0138	-0.0414	-0.0259	-0.0284	-0.0203	0.0029	0.1983	-0.0100	0.0605
<i>Chimaphila menziesii</i> *	Pyrolaceae	5	-0.0245	-0.0460	-0.0491	-0.0310	0.0634	0.0085	-0.0139	0.0322	0.1943	0.0404
<i>Chimaphila umbellata</i> ‡	Pyrolaceae	1	-0.0109	-0.0204	-0.0291	-0.0027	0.0370	0.0140	-0.0138	0.0581	0.2611	0.1104
<i>Chorispora tenella</i> *€	Cruciferae	1	0.1195	0.0037	-0.0215	-0.0240	-0.0163	-0.0150	-0.0138	-0.0127	0.0002	0.0114
<i>Chrysopsis jonesii</i> *	Compositae	1	-0.0109	-0.0204	-0.0290	-0.0054	0.0507	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Chrysopsis villosa</i>	Compositae	74	-0.0433	-0.0529	0.0206	0.1933	0.1966	-0.0952	-0.0770	-0.0592	0.0364	0.0487
<i>Chrysothamnus depressus</i>	Compositae	5	0.0294	-0.0360	-0.0271	-0.0197	-0.0127	0.0345	0.1618	0.0621	-0.0145	-0.0192
<i>Chrysothamnus greenei</i> *‡	Compositae	1	-0.0109	0.0045	0.0285	-0.0112	-0.0104	-0.0023	-0.0037	-0.0127	-0.0098	-0.0085
<i>Chrysothamnus nauseosus</i>	Compositae	39	-0.0081	0.0136	0.0303	0.1202	-0.0282	-0.0941	-0.0726	-0.0711	-0.0435	0.0411
<i>Chrysothamnus parryi</i>	Compositae	1	-0.0109	-0.0204	-0.0142	0.0130	-0.0109	0.0987	0.0279	-0.0032	-0.0098	-0.0085
<i>Chrysothamnus viscidiflorus</i>	Compositae	6	0.0518	0.0138	0.0050	0.0321	-0.0334	0.0095	0.0182	0.0337	-0.0206	-0.0211
<i>Cirsium arizonicum</i>	Compositae	16	0.0236	-0.0538	0.0075	0.0434	0.1052	0.0028	-0.0078	0.0262	0.0954	0.0911
<i>Cirsium wheeleri</i>	Compositae	8	0.2224	-0.0308	-0.0508	-0.0337	-0.0135	0.0147	0.0741	0.0514	0.0035	-0.0196
<i>Clematis columbiana</i>	Ranunculaceae	3	-0.0006	-0.0355	-0.0552	-0.0252	0.0046	0.0343	-0.0085	0.0509	0.0859	0.0117
<i>Coleogyne ramosissima</i>	Rosaceae	13	0.0145	0.2448	-0.0393	-0.0830	-0.0601	-0.0914	-0.0800	-0.0466	-0.0360	-0.0315
<i>Collinsia parviflora</i>	Scrophulariaceae	2	-0.0154	-0.0255	-0.0199	0.0254	0.0755	-0.0060	-0.0005	0.0123	-0.0139	-0.0121
<i>Collomia grandiflora</i> *	Polemoniaceae	4	-0.0219	-0.0178	0.0063	0.0087	0.0116	0.0250	0.0349	0.0067	0.0337	-0.0095
<i>Collomia linearis</i> *‡‡	Polemoniaceae	2	0.0286	0.0019	-0.0082	-0.0276	-0.0231	-0.0142	-0.0006	0.0544	-0.0139	0.0120
<i>Comandra umbellata</i>	Santalaceae	35	-0.0686	-0.0696	0.0777	0.1314	0.0282	0.0257	0.0821	-0.0146	-0.0470	-0.0505
<i>Convolvulus arvensis</i> €	Convolvulaceae	1	-0.0109	0.0235	-0.0212	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Cordylanthus kingii</i>	Scrophulariaceae	19	-0.0072	0.0745	0.0772	-0.0369	-0.0655	0.0145	0.0030	-0.0393	-0.0440	-0.0342
<i>Cordylanthus</i> sp.	Scrophulariaceae	1	-0.0109	0.0330	0.0669	-0.0240	-0.0163	-0.0150	-0.0138	-0.0127	-0.0098	-0.0085
<i>Cornus sericea</i> *	Cornaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0054	-0.0132	-0.0124	0.0096	0.0963	0.0050
<i>Crepis acuminata</i> *‡‡	Compositae	3	-0.0189	-0.0118	0.0150	0.0580	0.0018	-0.0193	-0.0003	-0.0178	-0.0170	-0.0149
<i>Crepis occidentalis</i> *	Compositae	1	-0.0109	0.0151	0.0314	-0.0097	-0.0104	0.0103	-0.0059	-0.0127	-0.0098	-0.0085
<i>Crepis runcinata</i> *‡	Compositae	2	-0.0154	-0.0156	-0.0085	0.0443	-0.0092	-0.0212	-0.0112	-0.0129	-0.0139	-0.0121
<i>Crepis</i> sp.	Compositae	2	0.0411	-0.0224	-0.0006	-0.0101	-0.0065	-0.0166	0.0171	0.0406	0.0260	0.0264
<i>Cryptantha cinerea</i> *	Boraginaceae	17	-0.0462	-0.0177	0.0358	0.2622	-0.0157	-0.0466	-0.0341	-0.0226	-0.0398	-0.0325
<i>Cryptantha circumscissa</i> *‡‡	Boraginaceae	3	0.0018	0.0247	-0.0137	0.0253	-0.0223	-0.0301	-0.0204	-0.0178	-0.0170	-0.0149

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Cryptantha confertiflora</i> *	Boraginaceae	31	-0.0351	0.0704	0.1252	0.0887	0.0972	-0.0691	-0.0637	-0.0683	-0.0403	-0.0469
<i>Cryptantha fendleri</i> *	Boraginaceae	2	0.0286	-0.0031	-0.0284	-0.0251	-0.0231	-0.0282	-0.0252	-0.0131	-0.0139	-0.0121
<i>Cryptantha humilis</i>	Boraginaceae	1	-0.0109	-0.0111	-0.0095	-0.0240	-0.0163	-0.0248	-0.0028	-0.0127	-0.0098	-0.0085
<i>Cryptantha</i> sp. (annual)	Boraginaceae	60	-0.0026	0.2085	0.1317	0.0211	-0.0596	-0.1192	-0.0796	-0.0746	-0.0776	-0.0639
<i>Cucurbita foetidissima</i> *	Cucurbitaceae	1	0.1155	-0.0117	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Cymopterus newberryi</i> *	Umbelliferae	1	-0.0109	-0.0106	0.0414	0.1042	-0.0083	-0.0248	-0.0119	0.0007	-0.0098	-0.0085
<i>Cymopterus purpureus</i> *	Umbelliferae	15	-0.0240	0.0333	0.0497	-0.0356	-0.0408	0.1293	0.0713	-0.0351	-0.0388	-0.0339
<i>Cymopterus</i> sp.*	Umbelliferae	1	0.0133	0.0499	-0.0212	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Cystopteris fragilis</i> *	Polypodiaceae	6	0.0155	-0.0505	-0.0547	-0.0311	0.0833	-0.0160	-0.0366	0.0764	0.3184	0.0691
<i>Dactylis glomerata</i> €	Gramineae	2	-0.0154	-0.0239	-0.0466	-0.0211	-0.0231	-0.0129	0.0009	0.3368	-0.0020	0.0686
<i>Dalea searlsiae</i>	Leguminosae	13	0.0076	0.1213	0.0237	-0.0688	-0.0565	-0.0649	-0.0570	-0.0412	-0.0360	-0.0315
<i>Datura wrightii</i>	Solanaceae	3	-0.0049	-0.0306	-0.0595	-0.0417	-0.0284	-0.0374	-0.0332	-0.0138	-0.0073	0.2023
<i>Delphinium andersonii</i> *	Ranunculaceae	1	-0.0109	-0.0106	0.0414	0.1042	-0.0083	-0.0248	-0.0119	0.0007	-0.0098	-0.0085
<i>Delphinium nuttallianum</i> *	Ranunculaceae	7	-0.0061	-0.0339	-0.0169	-0.0119	-0.0293	0.0741	0.0510	0.0040	-0.0172	-0.0141
<i>Descurainia californica</i> *	Cruciferae	1	0.0779	-0.0084	-0.0142	-0.0007	-0.0045	0.0076	0.0500	0.0264	-0.0098	-0.0085
<i>Descurainia pinnata</i>	Cruciferae	28	-0.0037	0.1716	0.0713	-0.0494	-0.0819	-0.0556	-0.0588	0.0116	-0.0520	-0.0214
<i>Descurainia sophia</i> *€	Cruciferae	1	-0.0109	-0.0204	-0.0279	-0.0165	-0.0163	-0.0150	0.0115	0.0787	-0.0016	0.0103
<i>Disporum trachycarpum</i> *	Liliaceae	4	-0.0219	-0.0411	-0.0482	-0.0337	0.0665	0.0404	-0.0081	0.0632	0.2342	0.0494
<i>Distichlis spicata</i>	Gramineae	1	-0.0109	0.0039	-0.0287	-0.0169	-0.0113	-0.0248	-0.0217	-0.0127	0.0131	0.1908
<i>Dodecatheon pulchellum</i> *	Primulaceae	2	-0.0154	-0.0290	-0.0469	-0.0340	-0.0108	-0.0123	-0.0086	-0.0025	0.0387	-0.0025
<i>Draba asprella</i> *	Cruciferae	33	-0.0516	-0.0823	0.0124	0.0971	0.2068	0.0802	0.0067	-0.0115	0.1409	-0.0142
<i>Draba cuneifolia</i> *	Cruciferae	1	0.0196	0.0051	0.0044	-0.0094	-0.0056	-0.0150	-0.0138	-0.0032	-0.0098	0.0094
<i>Draba verna</i> *‡‡€	Cruciferae	40	0.0217	0.2595	0.0867	-0.0141	-0.0704	-0.0816	-0.0745	-0.0737	-0.0615	-0.0189
<i>Dracocephalum parviflorum</i>	Labiate	1	0.0207	-0.0110	-0.0216	0.0050	-0.0113	0.0014	-0.0040	0.0148	-0.0098	-0.0085
<i>Dryopteris filix-mas</i> *	Polypodiaceae	2	-0.0154	-0.0290	-0.0467	-0.0189	0.0224	0.0006	-0.0186	0.0479	0.2531	0.0817
<i>Echinocereus engelmannii</i>	Cactaceae	7	0.0099	0.0998	0.0162	-0.0347	-0.0267	-0.0588	-0.0520	-0.0302	-0.0261	-0.0229
<i>Echinocereus triglochidiatus</i>	Cactaceae	22	-0.0407	0.1003	0.1300	0.0608	0.0065	-0.0478	-0.0232	-0.0488	-0.0394	-0.0377
<i>Eleocharis palustris</i> *	Cyperaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Eleocharis parishii</i> *	Cyperaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Elymus canadensis</i>	Gramineae	4	-0.0219	-0.0346	-0.0516	-0.0170	0.0062	-0.0254	-0.0357	0.0581	0.2267	0.2742
<i>Elymus elymoides</i>	Gramineae	58	0.0162	0.0923	0.0099	0.0133	0.0141	-0.0536	-0.0274	-0.0206	-0.0247	-0.0314

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Elymus hispidus</i> €	Gramineae	1	-0.0109	-0.0116	0.0146	-0.0026	-0.0053	-0.0051	-0.0059	-0.0127	-0.0043	-0.0085
<i>Elymus smithii</i> †‡‡ ²	Gramineae	3	0.1929	-0.0112	-0.0420	-0.0185	-0.0115	-0.0119	0.0309	0.0583	-0.0170	0.0127
<i>Elymus</i> sp.	Gramineae	2	-0.0109	-0.0139	-0.0177	-0.0046	-0.0111	-0.0100	-0.0099	-0.0091	-0.0098	-0.0085
<i>Elymus spicatus</i>	Gramineae	6	0.0722	-0.0048	-0.0195	-0.0271	-0.0193	0.0027	0.0116	0.0236	-0.0167	-0.0055
<i>Elymus trachycaulis</i> *	Gramineae	10	-0.0042	-0.0334	-0.0334	-0.0351	-0.0399	-0.0503	-0.0244	0.1829	0.0698	0.2409
<i>Encelia frutescens</i> *	Compositae	4	0.0475	0.1980	-0.0583	-0.0451	-0.0328	-0.0499	-0.0436	-0.0255	-0.0197	-0.0172
<i>Ephedra nevadensis</i>	Ephedraceae	18	0.0478	0.2637	0.0091	-0.0872	-0.0630	-0.0888	-0.0854	-0.0554	-0.0394	-0.0374
<i>Ephedra viridis</i>	Ephedraceae	16	-0.0215	0.0204	0.0730	0.0153	-0.0456	-0.0459	-0.0326	-0.0466	-0.0366	-0.0303
<i>Epilobium brachycarpum</i>	Onagraceae	3	-0.0189	-0.0331	-0.0461	0.0110	-0.0058	-0.0317	-0.0286	0.0187	0.1830	0.0390
<i>Epilobium ciliatum</i> *	Onagraceae	3	0.0225	-0.0355	-0.0595	-0.0253	0.0420	-0.0055	-0.0286	0.0497	0.1889	0.0707
<i>Epilobium glandulosum</i> *‡	Onagraceae	3	-0.0006	-0.0355	-0.0417	-0.0060	0.0404	0.0018	-0.0286	0.0690	0.2225	0.1360
<i>Epilobium</i> sp.	Onagraceae	1	-0.0109	-0.0204	-0.0279	-0.0165	-0.0163	-0.0150	0.0115	0.0787	-0.0016	0.0103
<i>Epipactis gigantea</i> *	Orchidaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	0.0179	-0.0085
<i>Equisetum arvense</i>	Equisetaceae	1	-0.0109	-0.0117	-0.0232	-0.0169	-0.0113	-0.0248	-0.0217	0.0263	0.0371	0.3576
<i>Equisetum hyemale</i>	Equisetaceae	8	0.0242	-0.0422	-0.0812	-0.0464	-0.0190	-0.0501	-0.0537	0.0372	0.1603	0.3744
<i>Equisetum laevigatum</i>	Equisetaceae	2	-0.0154	-0.0260	-0.0466	-0.0340	-0.0231	-0.0352	-0.0308	0.0097	0.0132	0.3213
<i>Eriastrum eremicum</i> *	Polemoniaceae	7	-0.0062	0.2058	0.0095	-0.0395	-0.0417	-0.0626	-0.0550	-0.0339	-0.0261	-0.0229
<i>Eriastrum sparsiflorum</i> *	Polemoniaceae	13	0.0561	0.1752	-0.0212	-0.0568	-0.0495	-0.0914	-0.0710	-0.0466	-0.0360	-0.0250
<i>Erigeron canaani</i> *	Compositae	16	-0.0448	-0.0719	-0.0209	0.1198	0.2338	0.0168	-0.0131	-0.0192	0.0259	-0.0316
<i>Erigeron divergens</i>	Compositae	17	-0.0097	-0.0041	0.0203	-0.0308	-0.0587	-0.0115	0.0173	0.0552	-0.0393	-0.0170
<i>Erigeron flagellaris</i> *	Compositae	2	-0.0154	-0.0198	-0.0127	-0.0103	0.0322	-0.0077	-0.0252	0.0184	0.0885	0.2472
<i>Erigeron pumilus</i>	Compositae	1	-0.0109	-0.0116	0.0146	-0.0026	-0.0053	-0.0051	-0.0059	-0.0127	-0.0043	-0.0085
<i>Erigeron religious</i> *	Compositae	1	-0.0109	-0.0001	-0.0138	0.0062	0.0029	0.0233	0.0284	0.0007	-0.0098	-0.0085
<i>Erigeron sionis</i> *	Compositae	4	-0.0219	-0.0387	-0.0358	0.0211	0.2217	0.0074	-0.0145	0.0187	0.1662	0.0426
<i>Erigeron</i> sp.	Compositae	14	0.0183	-0.0196	0.0122	0.0569	-0.0271	0.0253	0.0560	-0.0216	-0.0246	0.0674
<i>Erigeron speciosus</i>	Compositae	6	0.0224	-0.0409	-0.0374	-0.0328	-0.0154	0.0479	0.0804	0.1538	0.0046	0.0143
<i>Erigeron utahensis</i>	Compositae	41	-0.0341	0.1342	0.2246	0.0464	-0.0274	-0.0700	-0.0681	-0.0713	-0.0588	-0.0560
<i>Eriogonum alatum</i> *	Polygonaceae	6	-0.0269	-0.0291	-0.0297	0.2047	0.0216	-0.0118	-0.0125	-0.0224	-0.0147	-0.0211
<i>Eriogonum corymbosum</i>	Polygonaceae	4	0.0057	0.0237	-0.0251	-0.0224	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Eriogonum davidsonii</i> *	Polygonaceae	32	-0.0379	0.1334	0.1227	0.0399	-0.0310	-0.0761	-0.0554	-0.0592	-0.0503	-0.0478
<i>Eriogonum fasciculatum</i>	Polygonaceae	3	-0.0189	0.0211	0.0748	-0.0180	-0.0206	0.0043	-0.0128	-0.0220	-0.0170	-0.0149

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Eriogonum heracleoides</i> *‡	Polygonaceae	1	-0.0109	-0.0204	-0.0216	0.0856	-0.0029	-0.0024	-0.0019	-0.0127	-0.0098	-0.0085
<i>Eriogonum inflatum</i>	Polygonaceae	3	0.0610	0.1657	-0.0459	-0.0381	-0.0284	-0.0431	-0.0377	-0.0220	-0.0170	-0.0149
<i>Eriogonum jamesii</i> *	Polygonaceae	10	-0.0350	-0.0338	-0.0469	0.0466	0.2322	-0.0553	-0.0502	-0.0361	-0.0065	-0.0275
<i>Eriogonum microthecum</i>	Polygonaceae	15	0.0519	0.0652	0.0645	-0.0408	-0.0378	-0.0589	-0.0519	-0.0245	-0.0305	-0.0290
<i>Eriogonum palmerianum</i> *	Polygonaceae	15	0.0092	0.2175	0.0316	-0.0719	-0.0476	-0.0797	-0.0779	-0.0503	-0.0369	-0.0339
<i>Eriogonum racemosum</i>	Polygonaceae	59	-0.0291	-0.0898	0.0990	0.1001	0.0704	-0.0038	0.0302	0.0394	0.0517	-0.0253
<i>Eriogonum sp.</i>	Polygonaceae	7	-0.0291	0.0333	0.0162	-0.0002	-0.0297	-0.0475	-0.0430	-0.0339	-0.0261	-0.0229
<i>Eriogonum umbellatum</i>	Polygonaceae	21	-0.0382	-0.0305	0.0243	0.1447	0.0862	-0.0200	0.0085	-0.0145	0.0074	-0.0264
<i>Erioneuron pilosum</i> *‡	Gramineae	2	0.0099	0.1098	-0.0356	-0.0208	0.0332	-0.0352	-0.0308	-0.0179	-0.0139	-0.0121
<i>Erodium cicutarium</i> €	Geraniaceae	10	0.2865	0.1277	-0.0910	-0.0748	-0.0488	-0.0766	-0.0609	-0.0407	-0.0282	-0.0211
<i>Erysimum asperum</i>	Cruciferae	64	-0.0775	-0.0340	0.2064	0.1601	-0.0076	0.0100	0.0369	-0.0315	-0.0287	-0.0527
<i>Eupatorium herbaceum</i> *	Compositae	12	-0.0081	-0.0709	-0.0670	-0.0391	0.1492	0.0054	-0.0365	0.0630	0.3612	0.0462
<i>Euphorbia brachycera</i>	Euphorbiaceae	5	0.0414	-0.0382	-0.0518	0.0127	-0.0217	0.0515	0.0060	-0.0243	-0.0220	-0.0192
<i>Euphorbia spathulata</i> *	Euphorbiaceae	1	0.3561	0.0035	-0.0293	-0.0240	-0.0163	-0.0248	-0.0020	-0.0127	-0.0098	-0.0085
<i>Fendlerella utahensis</i>	Saxifragaceae	1	-0.0109	-0.0204	0.0064	-0.0240	0.0168	0.0473	0.0135	-0.0127	-0.0030	-0.0085
<i>Festuca octoflora</i>	Gramineae	74	0.0169	0.2480	0.0657	0.0993	-0.0069	-0.1467	-0.1123	-0.0946	-0.0819	-0.0469
<i>Fraxinus anomala</i>	Oleaceae	45	-0.0588	0.0431	0.2408	-0.0204	0.0183	0.0824	0.0020	-0.0571	-0.0092	-0.0520
<i>Fraxinus velutina</i>	Oleaceae	6	-0.0169	-0.0349	-0.0638	-0.0478	-0.0313	-0.0546	-0.0496	-0.0130	0.0263	0.1862
<i>Fritillaria atropurpurea</i> *	Liliaceae	9	-0.0225	-0.0276	0.0404	0.1442	-0.0179	-0.0199	0.0025	-0.0235	-0.0153	-0.0260
<i>Gaillardia pinnatifida</i> *	Compositae	2	0.0770	0.0830	-0.0413	-0.0340	-0.0231	-0.0282	-0.0252	-0.0179	-0.0068	0.0020
<i>Galium aparine</i>	Rubiaceae	6	-0.0144	-0.0180	-0.0198	-0.0275	-0.0202	-0.0195	-0.0191	0.0417	0.0227	0.0290
<i>Galium bifolium</i> *	Rubiaceae	2	0.0411	-0.0290	-0.0306	-0.0185	-0.0185	-0.0130	0.0362	0.2181	0.0041	0.0247
<i>Galium multiflorum</i> *	Rubiaceae	28	-0.0557	-0.0715	0.1091	0.0320	0.0953	0.0121	-0.0059	-0.0183	0.0520	-0.0249
<i>Galium sp.</i>	Rubiaceae	1	0.0196	0.0007	0.0788	-0.0020	-0.0163	0.0014	-0.0040	-0.0065	-0.0098	-0.0085
<i>Galium trifidum</i> *	Rubiaceae	2	-0.0154	-0.0194	0.0132	-0.0006	0.0406	0.0039	-0.0252	0.0003	0.0890	0.0264
<i>Galium triflorum</i> *	Rubiaceae	4	-0.0219	-0.0174	-0.0340	-0.0375	0.0169	-0.0196	-0.0310	0.0434	0.2616	0.0562
<i>Garrya flavescens</i> *	Garryaceae	9	-0.0331	-0.0363	0.0376	-0.0030	-0.0256	0.0300	-0.0135	-0.0232	-0.0226	-0.0260
<i>Gaura coccinea</i> *	Onagraceae	1	0.0249	0.1296	-0.0287	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Gayophytum sp.</i>	Onagraceae	14	0.0125	-0.0316	-0.0370	-0.0369	-0.0406	0.0238	0.0777	0.1380	-0.0272	0.0207
<i>Geranium richardsonii</i> ***‡‡	Geraniaceae	1	-0.0109	-0.0204	-0.0291	-0.0027	0.0370	0.0140	-0.0138	0.0581	0.2611	0.1104
<i>Gilia inconspicua</i> *	Polemoniaceae	84	-0.0137	0.2558	0.1513	0.0515	-0.1029	-0.1116	-0.0963	-0.1140	-0.0969	-0.0584

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Gilia scopulorum</i> *†††	Polemoniaceae	1	-0.0109	0.0004	0.0031	0.0269	-0.0163	0.0112	-0.0059	0.0014	-0.0098	-0.0085
<i>Glycyrrhiza lepidota</i> *	Leguminosae	1	0.1195	0.0037	-0.0215	-0.0240	-0.0163	-0.0150	-0.0138	-0.0127	0.0002	0.0114
<i>Gnaphalium wrightii</i> *	Compositae	9	-0.0331	-0.0003	0.0182	0.0485	0.0870	-0.0346	-0.0305	0.0031	0.1052	0.0465
<i>Goodyera oblongifolia</i> *	Orchidaceae	5	-0.0245	-0.0460	-0.0690	-0.0374	0.0260	0.0369	-0.0076	0.0882	0.2131	0.0404
<i>Grindelia squarrosa</i>	Compositae	1	0.0449	-0.0117	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Gutierrezia</i> sp. ³	Compositae	94	0.0989	0.2817	0.0670	-0.0696	-0.0984	-0.1590	-0.1114	-0.1043	-0.0962	-0.0086
<i>Hackelia patens</i> *	Boraginaceae	5	0.0219	-0.0146	0.0180	0.0033	-0.0109	-0.0110	-0.0057	0.0411	-0.0125	-0.0192
<i>Haplopappus armeriooides</i> *‡	Compositae	1	-0.0109	-0.0204	-0.0294	-0.0174	-0.0163	0.1376	0.0257	-0.0127	-0.0098	-0.0085
<i>Haplopappus gracilis</i>	Compositae	4	0.1243	-0.0202	-0.0699	-0.0411	-0.0278	-0.0449	-0.0397	-0.0255	-0.0081	0.1083
<i>Haplopappus laricifolius</i> ‡	Compositae	1	0.0207	0.0772	0.0151	-0.0240	-0.0106	-0.0150	-0.0138	-0.0127	-0.0098	-0.0085
<i>Haplopappus scopulorum</i>	Compositae	42	-0.0620	-0.0438	0.1804	0.0711	0.0046	0.0255	0.0324	-0.0509	-0.0055	-0.0515
<i>Helianthella quinquenervis</i> *	Compositae	1	-0.0109	-0.0161	-0.0052	-0.0165	-0.0099	0.0461	0.1300	-0.0058	-0.0098	-0.0085
<i>Helianthus annuus</i> *	Compositae	2	0.0241	-0.0166	-0.0522	-0.0340	-0.0231	-0.0352	-0.0308	-0.0179	-0.0139	0.0057
<i>Helianthus anomalous</i> *†††	Compositae	2	0.3310	-0.0058	-0.0468	-0.0340	-0.0231	-0.0352	-0.0168	-0.0179	-0.0139	-0.0121
<i>Helianthus nuttallii</i> *†††	Compositae	2	-0.0154	-0.0290	-0.0069	-0.0193	-0.0185	0.0860	0.1539	-0.0021	-0.0139	-0.0121
<i>Heuchera rubescens</i>	Saxifragaceae	12	-0.0292	-0.0709	-0.0461	-0.0292	0.1744	0.0382	-0.0019	0.0476	0.2533	-0.0007
<i>Hieracium fendleri</i> *†††	Compositae	1	-0.0109	-0.0156	-0.0138	0.0744	0.0327	0.0093	-0.0045	-0.0055	0.0131	-0.0085
<i>Hilaria</i> sp.	Gramineae	41	0.1280	0.2507	0.0402	-0.1195	-0.0871	-0.1262	-0.1138	-0.0683	-0.0675	-0.0510
<i>Holodiscus dumosus</i>	Rosaceae	21	-0.0356	-0.0921	-0.0438	0.0077	0.3078	-0.0046	-0.0308	-0.0027	0.1947	-0.0222
<i>Hordeum murinum</i> *€	Gramineae	1	0.3561	0.0035	-0.0293	-0.0240	-0.0163	-0.0248	-0.0020	-0.0127	-0.0098	-0.0085
<i>Hordeum</i> sp.	Gramineae	1	-0.0109	0.0044	-0.0049	0.0480	0.0072	-0.0248	-0.0119	-0.0065	-0.0098	-0.0085
<i>Hydrophyllum occidentale</i>	Hydrophyllaceae	2	-0.0154	-0.0290	-0.0359	-0.0251	0.0522	0.0165	-0.0130	0.0432	0.1573	-0.0025
<i>Hymenopappus filifolius</i>	Compositae	11	-0.0368	-0.0381	-0.0048	0.2352	-0.0077	-0.0134	-0.0248	-0.0122	-0.0309	-0.0242
<i>Hymenoxys cooperi</i> *	Compositae	5	0.1070	0.0776	0.0185	-0.0390	-0.0263	-0.0291	-0.0196	0.0033	-0.0183	-0.0192
<i>Ipomopsis aggregata</i>	Polemoniaceae	5	-0.0027	0.0176	0.0435	0.0043	-0.0292	0.0541	0.1344	-0.0049	-0.0220	-0.0112
<i>Ipomopsis congesta</i>	Polemoniaceae	14	-0.0417	-0.0395	0.0396	0.1277	0.0315	-0.0169	-0.0247	-0.0114	-0.0070	-0.0024
<i>Iva axillaris</i> *	Compositae	1	-0.0109	-0.0117	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	0.0166
<i>Ivesia sabulosa</i>	Rosaceae	7	-0.0291	-0.0435	-0.0373	0.0594	0.2693	-0.0287	-0.0318	-0.0284	0.0105	-0.0229
<i>Jamesia americana</i> *	Saxifragaceae	2	0.0352	-0.0260	-0.0466	-0.0290	0.0206	-0.0282	-0.0252	0.0007	0.1444	0.0538
<i>Juncus arcticus</i>	Juncaceae	3	-0.0189	-0.0305	-0.0471	-0.0180	-0.0143	-0.0431	-0.0377	0.0325	0.0552	0.3103
<i>Juncus ensifolius</i>	Juncaceae	2	-0.0154	-0.0228	-0.0425	-0.0290	-0.0150	-0.0166	-0.0252	0.0472	0.0611	0.2580

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Juncus tenuis</i>	Juncaceae	1	-0.0109	-0.0117	-0.0232	-0.0169	-0.0113	-0.0248	-0.0217	0.0263	0.0371	0.3576
<i>Juniperus osteosperma</i>	Cupressaceae	115	0.0251	0.1706	0.1868	-0.0462	-0.1486	0.0226	0.0318	-0.1258	-0.1273	-0.1076
<i>Juniperus scopulorum</i>	Cupressaceae	21	-0.0178	-0.0775	-0.0461	-0.0170	-0.0283	0.0539	0.0409	0.1754	0.0134	0.0109
<i>Koeleria macrantha</i>	Gramineae	1	0.0689	-0.0159	-0.0217	0.0052	-0.0109	0.0235	0.0359	0.0344	-0.0098	-0.0085
<i>Lactuca serriola</i> €	Compositae	13	0.2237	-0.0045	-0.0836	-0.0662	-0.0233	-0.0521	-0.0387	0.0166	0.0243	0.0185
<i>Lactuca tatarica</i> *	Compositae	2	0.0069	-0.0223	-0.0359	0.0017	0.0182	0.0109	-0.0126	0.0516	0.1780	0.0721
<i>Lappula occidentalis</i> *	Boraginaceae	2	0.0352	-0.0143	-0.0326	-0.0223	-0.0116	-0.0129	0.0009	0.0628	-0.0028	-0.0121
<i>Lathyrus brachycalyx</i>	Leguminosae	9	-0.0331	-0.0063	0.0917	-0.0259	-0.0346	0.0058	-0.0064	-0.0323	-0.0297	-0.0260
<i>Lathyrus lanszwertii</i> *	Leguminosae	20	-0.0150	-0.0783	-0.0283	-0.0158	-0.0433	0.1446	0.2432	0.0463	-0.0253	-0.0321
<i>Layia glandulosa</i> *	Compositae	2	-0.0154	0.0319	0.0088	0.0161	-0.0090	-0.0122	-0.0124	-0.0112	-0.0100	-0.0121
<i>Lepidium campestre</i> *‡€	Cruciferae	1	-0.0109	-0.0204	-0.0294	-0.0240	-0.0163	-0.0033	0.0053	0.1108	0.0136	0.0067
<i>Lepidium densiflorum</i> *	Cruciferae	8	0.1484	0.1408	-0.0475	-0.0499	-0.0344	-0.0559	-0.0466	-0.0341	-0.0254	-0.0245
<i>Lepidium fremontii</i> *	Cruciferae	1	0.0196	0.0550	-0.0107	-0.0177	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Lepidium lasiocarpum</i> *	Cruciferae	1	0.1195	0.0037	-0.0215	-0.0240	-0.0163	-0.0150	-0.0138	-0.0127	0.0002	0.0114
<i>Lepidium montanum</i> *	Cruciferae	1	0.0537	-0.0144	-0.0290	-0.0073	0.0010	-0.0033	0.0053	0.0870	-0.0098	0.0390
<i>Leptodactylon pungens</i>	Polemoniaceae	8	-0.0312	-0.0489	-0.0113	0.0261	0.2160	0.0105	0.0036	-0.0213	0.0551	-0.0196
<i>Leptodactylon watsonii</i>	Polemoniaceae	10	-0.0350	0.0074	0.0209	0.0080	0.0732	-0.0228	-0.0311	-0.0149	0.0196	-0.0275
<i>Leucocrinum montanum</i> *‡	Liliaceae	1	-0.0109	0.0073	0.0119	0.0210	-0.0045	0.0076	0.0042	-0.0127	-0.0098	-0.0085
<i>Linanthastrum nuttallii</i>	Polemoniaceae	19	-0.0433	-0.0681	0.0315	0.1760	0.1762	-0.0110	-0.0309	-0.0092	0.0617	-0.0090
<i>Linanthus dichotomus</i> *	Polemoniaceae	14	0.0000	0.1643	0.0400	-0.0073	-0.0175	-0.0784	-0.0666	-0.0485	-0.0335	-0.0265
<i>Linum perenne</i> €	Linaceae	6	-0.0070	-0.0198	0.0699	0.0251	0.0469	0.0372	0.0333	-0.0255	0.0071	-0.0134
<i>Lithophragma tenellum</i> *	Saxifragaceae	1	-0.0109	-0.0071	-0.0067	-0.0149	-0.0163	-0.0051	-0.0059	0.0257	-0.0012	0.0067
<i>Lithospermum incisum</i> *	Boraginaceae	2	0.0017	0.0099	0.0067	-0.0246	-0.0231	0.0576	0.0026	-0.0131	-0.0139	-0.0121
<i>Lithospermum ruderale</i> *	Boraginaceae	2	0.0524	-0.0114	-0.0033	0.0115	-0.0150	-0.0027	0.0386	0.0680	-0.0080	0.0006
<i>Lomatium dissectum</i> *	Umbelliferae	4	-0.0219	-0.0389	-0.0354	-0.0296	-0.0269	-0.0035	0.0358	0.1080	-0.0067	0.0000
<i>Lomatium graveolens</i>	Umbelliferae	1	-0.0109	-0.0204	-0.0290	-0.0240	0.0469	0.0256	-0.0045	0.0413	0.1068	-0.0085
<i>Lomatium sp.</i>	Umbelliferae	1	-0.0109	-0.0159	-0.0063	0.0034	-0.0035	-0.0150	-0.0138	-0.0127	-0.0098	-0.0085
<i>Lotus denticulatus</i> *	Leguminosae	6	0.1489	0.0262	-0.0177	-0.0513	-0.0379	-0.0439	-0.0235	-0.0285	-0.0242	-0.0211
<i>Lotus plebeius</i> *	Leguminosae	17	-0.0171	0.1402	0.1080	0.0393	-0.0188	-0.0600	-0.0524	-0.0514	-0.0374	-0.0363
<i>Lotus utahensis</i> *	Leguminosae	12	0.0040	-0.0308	-0.0165	-0.0260	-0.0268	0.0389	0.1321	0.0350	-0.0305	-0.0162
<i>Lupinus argenteus</i>	Leguminosae	1	-0.0109	0.0903	0.0057	-0.0106	-0.0056	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Lupinus brevicaulis</i> *	Leguminosae	4	-0.0219	0.0659	-0.0162	-0.0376	-0.0303	-0.0449	-0.0298	-0.0255	-0.0197	-0.0172
<i>Lupinus concinnus</i>	Leguminosae	1	-0.0109	0.0722	0.0141	0.0133	-0.0056	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Lupinus kingii</i> *	Leguminosae	2	-0.0154	0.0136	-0.0176	-0.0142	-0.0231	-0.0002	0.0474	-0.0036	-0.0139	-0.0121
<i>Lupinus sericeus</i>	Leguminosae	9	0.0692	-0.0536	-0.0838	-0.0294	-0.0299	0.0016	0.0436	0.2937	-0.0064	0.0352
<i>Lychnis drummondii</i> *	Caryophyllaceae	1	-0.0109	-0.0114	-0.0124	0.0988	0.0029	-0.0248	-0.0217	0.0355	-0.0098	0.0067
<i>Lycium pallidum</i>	Solanaceae	3	0.3355	0.0067	-0.0506	-0.0376	-0.0255	-0.0431	-0.0263	-0.0220	-0.0170	-0.0149
<i>Lygodesmia grandiflora</i>	Compositae	2	-0.0154	0.0277	0.0124	0.0441	-0.0193	-0.0122	-0.0040	-0.0129	-0.0139	-0.0121
<i>Machaeranthera canescens</i>	Compositae	40	-0.0036	0.0284	0.0883	0.1432	-0.0142	-0.0414	-0.0210	-0.0066	-0.0534	-0.0318
<i>Mahonia repens</i>	Berberidaceae	29	-0.0231	-0.0947	-0.0511	-0.0508	-0.0046	0.1614	0.0969	0.0752	0.1310	0.0000
<i>Medicago lupulina</i> €	Leguminosae	1	-0.0109	-0.0162	-0.0290	-0.0240	-0.0163	-0.0248	-0.0217	0.0015	0.0070	0.2964
<i>Melilotus alba</i> *€	Leguminosae	2	0.0993	-0.0119	-0.0413	-0.0340	-0.0231	-0.0282	-0.0252	-0.0003	-0.0010	0.0723
<i>Melilotus officinalis</i> €	Leguminosae	1	-0.0109	0.0039	-0.0287	-0.0169	-0.0113	-0.0248	-0.0217	-0.0127	0.0131	0.1908
<i>Melilotus</i> sp. €	Leguminosae	2	-0.0154	-0.0198	-0.0370	-0.0290	-0.0196	-0.0352	-0.0308	0.0197	0.0312	0.4632
<i>Mentzelia integrifolia</i> *	Loasaceae	1	0.0554	0.0417	-0.0293	-0.0177	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Mertensia arizonica</i>	Boraginaceae	7	-0.0044	-0.0469	-0.0832	-0.0364	-0.0166	-0.0108	0.0021	0.2907	0.0961	0.1045
<i>Mertensia fusiformis</i> *	Boraginaceae	3	-0.0189	-0.0260	-0.0181	-0.0255	-0.0106	0.1384	0.1672	-0.0149	-0.0170	-0.0149
<i>Microseris lindleyi</i> *	Compositae	8	0.0141	0.0280	0.0754	0.0552	-0.0134	-0.0329	-0.0171	-0.0204	-0.0223	-0.0211
<i>Microsteris gracilis</i> *	Polemoniaceae	28	-0.0384	-0.0110	0.0462	0.0624	0.0171	-0.0096	0.0168	0.0556	-0.0135	-0.0144
<i>Mimulus cardinalis</i> *	Scrophulariaceae	2	-0.0154	-0.0260	-0.0466	-0.0340	-0.0231	-0.0282	-0.0252	0.0007	0.1183	0.0442
<i>Mimulus floribundus</i> *	Scrophulariaceae	1	-0.0109	0.0337	0.0295	0.0488	0.0036	-0.0150	-0.0138	-0.0127	-0.0098	-0.0085
<i>Mimulus guttatus</i> *	Scrophulariaceae	3	-0.0189	-0.0331	-0.0550	-0.0294	0.0063	0.0002	-0.0240	0.0649	0.2451	0.1089
<i>Mimulus parryi</i> *	Scrophulariaceae	1	-0.0109	0.0073	0.0119	0.0210	-0.0045	0.0076	0.0042	-0.0127	-0.0098	-0.0085
<i>Mirabilis multiflora</i>	Nyctaginaceae	7	0.1470	0.1333	-0.0178	-0.0565	-0.0365	-0.0588	-0.0442	-0.0339	-0.0223	-0.0152
<i>Mirabilis pumila</i> **†‡	Nyctaginaceae	1	0.0491	0.0152	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Monardella odoratissima</i>	Labiatae	6	0.0025	-0.0488	-0.0627	-0.0131	0.1961	-0.0348	-0.0324	-0.0244	0.0404	-0.0155
<i>Montia perfoliata</i>	Portulacaceae	3	-0.0189	-0.0110	0.0469	0.0374	0.0018	-0.0149	-0.0227	-0.0033	0.0138	0.0167
<i>Muhlenbergia asperifolia</i>	Gramineae	2	0.0770	-0.0089	-0.0358	-0.0340	-0.0231	-0.0282	-0.0252	-0.0079	0.0051	0.2180
<i>Muhlenbergia montana</i>	Gramineae	2	-0.0154	-0.0224	-0.0306	0.0769	0.0860	-0.0117	-0.0120	-0.0129	-0.0045	-0.0121
<i>Muhlenbergia pungens</i> *	Gramineae	5	-0.0245	-0.0001	-0.0069	0.1187	-0.0227	-0.0316	-0.0295	-0.0253	-0.0220	-0.0192
<i>Muhlenbergia racemosa</i>	Gramineae	3	-0.0006	-0.0355	-0.0507	-0.0256	0.0293	0.0018	-0.0286	0.0515	0.2055	0.0806
<i>Muhlenbergia</i> sp.	Gramineae	1	-0.0109	-0.0115	0.0011	-0.0177	-0.0163	0.1087	0.0330	-0.0065	-0.0098	-0.0085

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Muhlenbergia thurberi</i>	Gramineae	12	-0.0174	-0.0610	-0.0526	0.0809	0.2838	-0.0207	-0.0274	-0.0274	0.0811	-0.0182
<i>Myosurus cupulatus*</i>	Ranunculaceae	1	-0.0109	-0.0068	0.0019	-0.0177	-0.0163	0.0014	-0.0138	-0.0127	-0.0098	-0.0085
<i>Nasturtium officinale</i>	Cruciferae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Navarretia breweri*</i>	Polemoniaceae	5	0.0225	-0.0303	-0.0351	-0.0051	-0.0103	0.0073	0.0645	0.0414	-0.0220	0.0022
<i>Nicotiana attenuata</i>	Solanaceae	1	0.0512	-0.0001	-0.0138	-0.0240	-0.0163	-0.0150	-0.0138	-0.0058	-0.0098	-0.0085
<i>Oenothera caespitosa</i>	Onagraceae	9	-0.0033	-0.0030	0.0777	0.0068	-0.0066	0.0235	0.0220	-0.0001	-0.0194	-0.0139
<i>Oenothera longissima</i>	Onagraceae	2	-0.0154	-0.0198	-0.0370	-0.0290	-0.0196	-0.0352	-0.0308	0.0197	0.0312	0.4632
<i>Oenothera pallida</i>	Onagraceae	12	-0.0224	0.0081	0.0494	0.1056	-0.0116	-0.0433	-0.0436	-0.0284	-0.0092	0.1363
<i>Oenothera</i> sp.	Onagraceae	2	0.1635	-0.0057	-0.0413	-0.0340	-0.0231	-0.0282	-0.0252	-0.0179	-0.0068	0.0020
<i>Opuntia erinacea</i>	Cactaceae	17	-0.0068	0.2280	0.0095	-0.0750	-0.0492	-0.0720	-0.0542	-0.0497	-0.0380	-0.0261
<i>Opuntia macrorhiza</i>	Cactaceae	120	-0.0871	0.0051	0.1969	0.2100	0.0551	0.0113	0.0278	-0.0893	-0.0880	-0.1079
<i>Opuntia phaeacantha</i>	Cactaceae	9	0.1030	0.0449	0.0187	0.0060	-0.0339	-0.0589	-0.0420	-0.0291	-0.0297	-0.0200
<i>Opuntia polyacantha†‡</i>	Cactaceae	3	-0.0189	0.0721	-0.0325	-0.0381	-0.0284	-0.0431	-0.0377	-0.0220	-0.0170	-0.0149
<i>Opuntia whipplei</i>	Cactaceae	3	0.0158	0.0719	-0.0459	-0.0339	-0.0255	-0.0431	-0.0377	-0.0220	-0.0170	-0.0149
<i>Orobanche fasciculata*</i>	Orobanchaceae	2	-0.0154	-0.0220	0.0088	0.1714	0.0019	-0.0282	-0.0098	-0.0085	-0.0139	-0.0121
<i>Orobanche uniflora*†‡</i>	Orobanchaceae	1	-0.0109	-0.0026	0.0039	0.0039	-0.0163	0.0246	0.0886	0.0076	-0.0098	-0.0085
<i>Oryzopsis micrantha*</i>	Gramineae	2	-0.0154	-0.0254	0.0236	-0.0089	-0.0003	0.0515	-0.0018	0.0008	-0.0089	0.0012
<i>Osmorrhiza occidentalis</i>	Umbelliferae	4	-0.0060	-0.0258	-0.0429	-0.0358	0.0221	0.0098	-0.0047	0.1617	0.1010	0.0541
<i>Pachystima myrsinoides</i>	Celastraceae	54	-0.0573	-0.1407	-0.0118	0.0118	0.0706	0.2096	0.1310	0.0248	0.1121	-0.0386
<i>Panicum</i> sp.*	Gramineae	2	-0.0154	-0.0052	-0.0199	0.0220	-0.0029	-0.0352	-0.0238	0.0140	0.0194	0.2472
<i>Pectocarya setosa*</i>	Boraginaceae	3	0.0226	0.0666	-0.0321	-0.0339	-0.0255	-0.0431	-0.0263	-0.0220	-0.0170	-0.0149
<i>Pedicularis centranthera*</i>	Scrophulariaceae	10	-0.0194	-0.0305	0.0315	-0.0249	-0.0408	0.1849	0.1029	-0.0235	-0.0314	-0.0275
<i>Pediomelum mephiticum*</i>	Leguminosae	2	0.0069	0.0005	0.0142	-0.0157	-0.0231	0.0063	0.0156	-0.0179	-0.0139	-0.0121
<i>Pellaea truncata*</i>	Polypodiaceae	1	-0.0109	-0.0008	0.0154	-0.0174	-0.0163	-0.0033	-0.0045	-0.0058	-0.0098	-0.0085
<i>Penstemon angustifolius*‡</i>	Scrophulariaceae	38	-0.0262	-0.0539	0.0177	0.0474	0.0685	0.0513	0.0048	0.0071	0.0290	-0.0272
<i>Penstemon barbatus*</i>	Scrophulariaceae	6	-0.0269	-0.0216	0.0152	0.0110	0.0565	0.0150	0.0015	0.0004	0.1087	0.0279
<i>Penstemon comarrhenus</i>	Scrophulariaceae	2	0.0017	0.0004	0.0249	0.0411	-0.0196	0.0076	0.0306	0.0016	-0.0139	0.0006
<i>Penstemon confertus*</i>	Scrophulariaceae	3	-0.0189	-0.0032	0.0457	-0.0067	-0.0143	-0.0244	-0.0117	-0.0220	-0.0170	-0.0149
<i>Penstemon eatonii</i>	Scrophulariaceae	23	-0.0439	0.0296	0.1760	0.0390	-0.0203	-0.0281	-0.0238	-0.0371	-0.0302	0.0407
<i>Penstemon higginsii</i>	Scrophulariaceae	5	-0.0245	-0.0318	-0.0169	-0.0317	-0.0180	0.0574	0.0720	0.0924	-0.0099	-0.0124
<i>Penstemon humilis</i>	Scrophulariaceae	27	-0.0593	-0.0565	0.0790	0.1052	0.2021	-0.0007	-0.0125	-0.0374	0.0411	-0.0438

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Penstemon laevis</i> *	Scrophulariaceae	26	-0.0532	-0.0162	0.0773	0.2316	0.0846	-0.0416	-0.0335	-0.0542	-0.0342	-0.0456
<i>Penstemon linarioides</i>	Scrophulariaceae	13	0.0007	-0.0437	-0.0164	-0.0284	-0.0471	0.0904	0.2105	0.0331	-0.0360	-0.0167
<i>Penstemon pachyphyllus</i>	Scrophulariaceae	1	-0.0109	0.0759	0.0043	-0.0034	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Penstemon palmeri</i>	Scrophulariaceae	19	-0.0032	0.1018	0.0837	-0.0440	-0.0310	-0.0624	-0.0480	-0.0468	-0.0295	0.0380
<i>Penstemon rostriflorus</i>	Scrophulariaceae	30	-0.0179	-0.0415	0.0496	-0.0137	0.1297	0.0101	-0.0091	0.0460	0.1111	-0.0184
<i>Peraphyllum ramosissimum</i>	Rosaceae	10	-0.0016	-0.0473	-0.0258	-0.0263	-0.0351	0.1847	0.2389	0.0165	-0.0293	-0.0275
<i>Perityle tenella</i> *	Compositae	1	-0.0109	-0.0204	-0.0368	-0.0240	0.0041	-0.0248	-0.0217	-0.0127	0.0277	0.0050
<i>Peteria thompsoniae</i> *	Leguminosae	5	0.0215	0.1095	-0.0140	-0.0511	-0.0311	-0.0559	-0.0489	-0.0254	-0.0220	-0.0192
<i>Petradoria pumila</i>	Compositae	2	0.0128	-0.0290	-0.0066	0.0259	0.0074	-0.0060	0.0762	0.0069	-0.0139	-0.0121
<i>Petrophytum caespitosum</i>	Rosaceae	12	-0.0385	-0.0612	-0.0582	0.0268	0.3143	0.0020	-0.0243	-0.0130	0.1274	0.0088
<i>Phacelia fremontii</i> *	Hydrophyllaceae	6	-0.0144	0.1215	0.0457	-0.0356	-0.0354	-0.0399	-0.0365	-0.0313	-0.0242	-0.0211
<i>Phacelia heterophylla</i>	Hydrophyllaceae	42	-0.0498	-0.0651	0.0028	0.0973	0.0148	-0.0231	-0.0256	0.1026	0.0676	0.1188
<i>Phacelia ivesiana</i> *	Hydrophyllaceae	1	0.0196	0.0051	0.0044	-0.0094	-0.0056	-0.0150	-0.0138	-0.0032	-0.0098	0.0094
<i>Phacelia palmeri</i> *	Hydrophyllaceae	1	0.0554	0.0417	-0.0293	-0.0177	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Philadelphus microphyllus</i> *	Saxifragaceae	10	-0.0248	-0.0600	-0.0154	-0.0246	0.1294	0.0352	-0.0116	0.0565	0.2350	0.0047
<i>Phlox austromontana</i>	Polemoniaceae	54	-0.0767	-0.0520	0.1251	0.1886	0.0721	0.0491	0.0433	-0.0512	-0.0503	-0.0675
<i>Phlox longifolia</i> *	Polemoniaceae	11	0.0053	-0.0238	0.0293	-0.0061	-0.0282	0.1193	0.2187	-0.0028	-0.0293	-0.0234
<i>Phoradendron juniperinum</i>	Viscaceae	14	-0.0331	-0.0010	0.1106	0.0545	-0.0285	0.0484	0.0399	-0.0264	-0.0335	-0.0327
<i>Phragmites australis</i>	Gramineae	2	-0.0154	-0.0087	-0.0409	-0.0290	-0.0196	-0.0352	-0.0308	-0.0079	0.0143	0.3451
<i>Physalis hederifolia</i>	Solanaceae	8	0.0857	-0.0088	-0.0355	-0.0285	-0.0332	0.0003	-0.0346	-0.0226	-0.0044	0.0376
<i>Physalis longifolia</i> *	Solanaceae	1	0.1195	0.0037	-0.0215	-0.0240	-0.0163	-0.0150	-0.0138	-0.0127	0.0002	0.0114
<i>Physaria chambersii</i>	Cruciferae	3	-0.0049	-0.0200	0.0859	-0.0097	-0.0166	0.0871	0.0155	-0.0220	-0.0170	-0.0149
<i>Physaria ludoviciana</i> *	Cruciferae	3	-0.0189	0.0209	0.0691	0.1120	-0.0154	-0.0187	-0.0113	-0.0137	-0.0170	-0.0149
<i>Physaria newberryi</i>	Cruciferae	7	-0.0106	-0.0018	0.0068	-0.0185	-0.0416	0.1630	0.0418	-0.0279	-0.0261	-0.0229
<i>Physaria rectipes</i> *	Cruciferae	3	-0.0189	-0.0058	0.0326	0.1040	-0.0168	-0.0187	-0.0125	-0.0143	-0.0170	-0.0149
<i>Physaria wardii</i> *	Cruciferae	1	-0.0109	-0.0161	-0.0141	-0.0163	0.0010	0.0474	0.2013	-0.0065	-0.0098	-0.0085
<i>Physocarpus alternans</i> *	Rosaceae	1	-0.0109	-0.0204	0.0064	-0.0240	0.0168	0.0473	0.0135	-0.0127	-0.0030	-0.0085
<i>Pinus edulis</i>	Pinaceae	53	-0.0363	-0.0799	0.0969	0.0150	-0.0519	0.2160	0.1513	-0.0540	-0.0481	-0.0640
<i>Pinus monophylla</i>	Pinaceae	64	-0.0639	0.1617	0.1964	-0.0616	-0.1071	-0.1054	-0.0930	-0.0808	-0.0790	-0.0597
<i>Pinus ponderosa</i>	Pinaceae	60	-0.0574	-0.1236	-0.0675	0.1735	0.1993	0.0611	0.0717	-0.0050	0.0285	-0.0322
<i>Plantago patagonica</i>	Plantaginaceae	13	0.1238	0.1362	-0.0524	0.0399	-0.0229	-0.0564	-0.0376	-0.0292	-0.0360	-0.0315

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Poa bigelovii*</i>	Gramineae	8	0.0487	0.1065	-0.0240	-0.0542	-0.0388	-0.0675	-0.0593	-0.0329	-0.0280	-0.0181
<i>Poa bulbosa*€</i>	Gramineae	1	0.0848	-0.0162	-0.0368	-0.0169	-0.0113	-0.0248	-0.0217	-0.0127	-0.0016	0.0094
<i>Poa compressa*€</i>	Gramineae	3	0.0185	-0.0296	-0.0550	-0.0320	-0.0183	-0.0307	-0.0221	0.0583	0.0051	0.2434
<i>Poa fendleriana</i>	Gramineae	190	-0.1391	-0.0620	0.2132	0.1283	0.1186	0.0577	0.0891	-0.0417	-0.0072	-0.0837
<i>Poa pratensis€</i>	Gramineae	14	-0.0045	-0.0598	-0.0991	-0.0276	-0.0409	-0.0455	-0.0258	0.2466	0.0391	0.2027
<i>Poa secunda</i>	Gramineae	4	-0.0219	-0.0150	0.0621	0.0318	0.0205	-0.0237	-0.0227	-0.0207	-0.0162	-0.0172
<i>Poa sp.</i>	Gramineae	1	0.0196	0.0257	0.0239	-0.0177	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Polygonum douglasii</i>	Polygonaceae	39	-0.0015	-0.0693	-0.0321	0.1548	0.0311	0.0050	0.0855	0.1711	-0.0403	-0.0010
<i>Polypodium hesperium</i>	Polypodiaceae	6	-0.0269	-0.0470	-0.0363	-0.0402	0.0592	-0.0031	-0.0263	0.0408	0.2726	0.0228
<i>Populus fremontii</i>	Salicaceae	6	-0.0139	-0.0177	-0.0756	-0.0534	-0.0362	-0.0613	-0.0487	0.0008	0.0149	0.3936
<i>Populus tremuloides</i>	Salicaceae	2	-0.0154	-0.0290	-0.0359	-0.0100	-0.0095	-0.0199	-0.0116	0.0909	0.0426	0.0664
<i>Prunus virginiana</i>	Rosaceae	10	0.0293	-0.0582	-0.0766	-0.0404	-0.0332	0.0307	0.0676	0.2923	0.0006	0.0201
<i>Pseudotsuga menziesii</i>	Pinaceae	18	-0.0476	-0.0849	-0.0415	-0.0183	0.1328	0.0553	0.0042	0.0309	0.2559	0.0212
<i>Psoralidium lanceolatum*</i>	Leguminosae	4	0.0437	0.0173	-0.0447	-0.0232	-0.0303	-0.0449	-0.0337	-0.0255	-0.0146	0.0044
<i>Psorothamnus fremontii</i>	Leguminosae	11	0.0240	0.2031	-0.0107	-0.0679	-0.0532	-0.0747	-0.0660	-0.0428	-0.0330	-0.0289
<i>Pteridium aquilinum</i>	Polypodiaceae	3	-0.0189	-0.0327	-0.0371	0.0472	0.0421	-0.0009	-0.0232	0.0407	0.1626	0.1094
<i>Pterospora andromedea</i>	Monotropaceae	2	-0.0154	-0.0204	-0.0137	-0.0027	-0.0071	-0.0007	-0.0126	0.0241	0.0224	-0.0121
<i>Purshia mexicana</i>	Rosaceae	15	-0.0140	0.0794	0.0437	-0.0691	-0.0602	-0.0049	-0.0317	-0.0468	-0.0388	-0.0339
<i>Purshia tridentata</i>	Rosaceae	36	-0.0398	-0.0642	0.0241	0.0959	-0.0355	0.1318	0.1832	-0.0297	-0.0563	-0.0475
<i>Quercus gambelii</i>	Fagaceae	135	-0.0835	-0.2123	-0.0259	0.0354	0.0422	0.1792	0.1804	0.1418	0.0666	-0.0480
<i>Quercus turbinella</i>	Fagaceae	50	-0.0617	0.1136	0.2131	0.0456	-0.0280	-0.0980	-0.0825	-0.0805	-0.0597	-0.0524
<i>Ranunculus andersonii*</i>	Ranunculaceae	1	-0.0109	0.0337	0.0295	0.0488	0.0036	-0.0150	-0.0138	-0.0127	-0.0098	-0.0085
<i>Ranunculus sp.*</i>	Ranunculaceae	1	-0.0109	-0.0117	-0.0232	-0.0169	-0.0113	-0.0248	-0.0217	0.0263	0.0371	0.3576
<i>Ranunculus testiculatus*€</i>	Ranunculaceae	2	0.0241	-0.0166	-0.0522	-0.0340	-0.0231	-0.0352	-0.0308	-0.0179	-0.0139	0.0057
<i>Rhus aromatica</i>	Anacardiaceae	23	-0.0011	0.1709	0.1113	-0.0483	-0.0489	-0.0629	-0.0392	-0.0439	-0.0425	-0.0388
<i>Ribes velutinum*</i>	Saxifragaceae	1	-0.0109	-0.0161	0.0011	-0.0175	-0.0079	-0.0051	-0.0059	-0.0058	-0.0035	-0.0085
<i>Robinia neomexicana*</i>	Leguminosae	1	-0.0109	0.0029	0.0022	-0.0149	-0.0163	-0.0051	0.0130	0.0826	-0.0098	0.0255
<i>Rosa woodsii</i>	Rosaceae	12	0.0277	-0.0648	-0.0878	-0.0490	-0.0177	0.0722	0.0501	0.2403	0.0501	0.0438
<i>Rubia tinctoria*€</i>	Rubiaceae	1	0.1195	0.0037	-0.0215	-0.0240	-0.0163	-0.0150	-0.0138	-0.0127	0.0002	0.0114
<i>Rubus leucodermis*</i>	Rosaceae	4	-0.0219	-0.0390	-0.0585	-0.0420	0.0088	-0.0278	-0.0310	0.0373	0.2611	0.0432
<i>Rumex hymenosepalus</i>	Polygonaceae	2	0.0912	0.0270	-0.0411	-0.0340	-0.0231	-0.0352	-0.0308	-0.0179	-0.0139	-0.0121

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Salix exigua</i>	Salicaceae	3	-0.0189	-0.0139	-0.0469	-0.0335	-0.0226	-0.0431	-0.0377	0.0088	0.0332	0.4594
<i>Salix scouleriana</i>	Salicaceae	2	-0.0154	-0.0290	-0.0466	-0.0340	-0.0185	-0.0166	-0.0182	0.0239	0.0423	0.0246
<i>Salvia dorrii</i>	Labiatae	3	-0.0189	0.0757	0.0290	-0.0148	-0.0249	-0.0301	-0.0273	-0.0220	-0.0170	-0.0149
<i>Samolus floribundus*</i>	Primulaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Saxifraga rhomboidea*</i>	Saxifragaceae	1	-0.0109	-0.0159	-0.0006	0.0311	0.0898	0.0053	-0.0092	-0.0029	0.0390	-0.0085
<i>Scirpus microcarpus*</i>	Cyperaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Scirpus pungens*</i>	Cyperaceae	1	-0.0109	0.0039	-0.0287	-0.0169	-0.0113	-0.0248	-0.0217	-0.0127	0.0131	0.1908
<i>Secale cereale*</i> €	Gramineae	2	-0.0154	-0.0197	-0.0464	-0.0340	-0.0231	-0.0282	-0.0252	-0.0179	-0.0139	-0.0121
<i>Sedum debile*</i>	Crassulaceae	2	-0.0154	-0.0290	-0.0064	-0.0009	0.1569	-0.0060	-0.0005	0.0304	0.0922	-0.0025
<i>Sedum lanceolatum*</i>	Crassulaceae	3	-0.0189	-0.0245	-0.0156	0.1417	0.1751	-0.0182	-0.0178	-0.0095	0.0031	-0.0149
<i>Selaginella underwoodii*</i>	Selaginellaceae	17	-0.0462	-0.0703	-0.0063	0.0248	0.2741	0.0094	-0.0260	0.0099	0.1728	-0.0180
<i>Selaginella utahensis*</i>	Selaginellaceae	2	-0.0154	-0.0255	-0.0142	0.0623	0.1932	-0.0269	-0.0242	-0.0129	0.0264	-0.0121
<i>Selaginella watsonii**††</i>	Selaginellaceae	1	-0.0109	-0.0015	0.0336	0.0405	0.0036	0.0076	0.0042	-0.0032	-0.0043	-0.0085
<i>Senecio eremophilus</i>	Compositae	3	-0.0189	-0.0314	-0.0552	-0.0312	-0.0284	-0.0125	0.0038	0.3109	0.0063	0.0600
<i>Senecio hydrophilus*‡</i>	Compositae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Senecio multilobatus</i>	Compositae	99	-0.0802	-0.1087	0.1615	0.1919	0.1136	0.0662	0.0478	-0.0288	-0.0054	-0.0606
<i>Senecio spartioides</i>	Compositae	26	-0.0242	-0.0684	-0.0308	0.0456	0.0257	-0.0538	-0.0629	0.0085	0.1054	0.0200
<i>Shepherdia rotundifolia*</i>	Elaeagnaceae	17	-0.0160	0.1196	0.1785	-0.0524	-0.0565	-0.0523	-0.0547	-0.0453	-0.0397	-0.0271
<i>Silene antirrhina*</i>	Caryophyllaceae	3	-0.0189	0.0432	0.0061	0.0642	0.0129	-0.0307	-0.0278	-0.0178	-0.0131	0.0034
<i>Silene verecunda*</i>	Caryophyllaceae	5	-0.0245	-0.0317	0.0558	0.0450	0.0094	0.0048	-0.0040	-0.0124	-0.0171	-0.0192
<i>Sisymbrium altissimum*</i> €	Cruciferae	4	0.0640	0.0076	-0.0258	-0.0328	-0.0249	-0.0350	-0.0222	0.0272	-0.0197	0.0090
<i>Sisymbrium irio*‡‡*</i> €	Cruciferae	2	0.0271	0.0024	-0.0425	-0.0290	-0.0196	-0.0352	-0.0308	0.0097	0.0194	0.2472
<i>Smilacina racemosa</i>	Liliaceae	6	-0.0139	-0.0488	-0.0248	-0.0427	0.0917	0.0248	-0.0148	0.0463	0.2531	0.0089
<i>Smilacina stellata</i>	Liliaceae	3	-0.0189	-0.0355	-0.0640	-0.0334	-0.0215	0.0102	-0.0142	0.0284	0.0379	-0.0060
<i>Solanum sarachoides*</i> €	Solanaceae	1	-0.0109	-0.0162	-0.0368	-0.0123	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Solanum</i> sp.	Solanaceae	1	-0.0109	-0.0162	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Solidago velutina</i>	Compositae	38	0.0085	-0.1190	-0.0889	0.0046	0.1890	0.0614	0.1036	0.0889	0.1476	-0.0134
<i>Sonchus</i> sp.* €	Compositae	3	-0.0006	-0.0280	-0.0428	-0.0338	0.0013	-0.0207	-0.0332	0.0413	0.0915	0.3652
<i>Sphaeralcea ambigua</i>	Malvaceae	2	-0.0154	-0.0100	-0.0176	-0.0142	-0.0231	0.0068	0.0530	-0.0036	-0.0139	-0.0121
<i>Sphaeralcea grossulariifolia</i>	Malvaceae	14	0.2053	0.0710	-0.0075	-0.0691	-0.0565	-0.0771	-0.0643	-0.0423	-0.0347	-0.0224
<i>Sphaeralcea parvifolia*</i>	Malvaceae	1	-0.0109	-0.0043	-0.0264	-0.0115	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Sphaeromeria ruthiae</i>	Compositae	3	-0.0189	-0.0331	-0.0550	-0.0417	0.0001	-0.0374	-0.0332	0.0187	0.1957	0.0469
<i>Sporobolus contractus</i> *	Gramineae	2	0.0188	-0.0230	-0.0522	-0.0340	-0.0231	-0.0282	-0.0252	-0.0179	-0.0068	0.0665
<i>Sporobolus cryptandrus</i>	Gramineae	50	0.0570	0.0424	0.0237	0.1236	-0.0302	-0.1019	-0.0813	-0.0708	-0.0588	-0.0061
<i>Stanleya pinnata</i>	Cruciferae	2	0.0823	0.0677	-0.0303	-0.0340	-0.0142	-0.0352	-0.0308	-0.0179	-0.0139	-0.0121
<i>Stellaria jamesiana</i>	Caryophyllaceae	18	0.0048	-0.0822	-0.1064	-0.0712	-0.0308	0.0271	0.0497	0.3314	0.0534	0.0380
<i>Stephanomeria exigua</i>	Compositae	1	-0.0109	-0.0043	-0.0264	-0.0115	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Stephanomeria runcinata</i> *‡	Compositae	2	0.0315	0.0213	-0.0105	-0.0144	-0.0153	-0.0212	-0.0196	-0.0179	-0.0100	-0.0121
<i>Stephanomeria tenuifolia</i>	Compositae	51	-0.0564	-0.0310	0.0846	0.1172	0.2097	-0.0459	-0.0537	-0.0463	0.0698	-0.0371
<i>Stipa comata</i>	Gramineae	39	0.0881	0.1120	0.0299	0.0637	-0.0626	-0.0597	-0.0169	-0.0122	-0.0602	-0.0490
<i>Stipa hymenoides</i>	Gramineae	65	-0.0221	0.0813	0.0886	0.0327	-0.0549	-0.0027	-0.0408	-0.0788	-0.0721	-0.0355
<i>Stipa lettermanii</i>	Gramineae	12	-0.0077	-0.0631	-0.0723	-0.0170	-0.0363	0.0173	0.0911	0.2524	-0.0184	0.0318
<i>Stipa nelsonii</i>	Gramineae	5	0.0436	-0.0389	-0.0550	-0.0215	-0.0285	-0.0135	0.0612	0.2604	-0.0069	0.0320
<i>Stipa neomexicana</i> *†‡	Gramineae	1	0.0249	0.0040	-0.0135	-0.0240	-0.0100	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Stipa pinetorum</i> *‡	Gramineae	2	0.0411	-0.0227	-0.0200	0.0077	-0.0155	0.0865	0.0987	0.0254	-0.0139	-0.0121
<i>Stipa speciosa</i>	Gramineae	15	-0.0254	0.1855	0.0749	-0.0627	-0.0373	-0.0881	-0.0779	-0.0467	-0.0332	-0.0292
<i>Streptanthella longirostris</i> *	Cruciferae	11	-0.0200	0.1109	0.1070	0.0203	0.0009	-0.0483	-0.0412	-0.0386	-0.0243	-0.0234
<i>Streptanthus cordatus</i>	Cruciferae	38	-0.0361	0.0388	0.1502	-0.0189	-0.0797	0.0734	0.0374	-0.0617	-0.0623	-0.0533
<i>Streptopus amplexifolius</i> *‡	Liliaceae	1	-0.0109	0.0045	0.0285	-0.0112	-0.0104	-0.0023	-0.0037	-0.0127	-0.0098	-0.0085
<i>Swertia albomarginata</i> *	Gentianaceae	11	-0.0219	0.1751	0.0796	-0.0462	-0.0533	-0.0511	-0.0527	-0.0428	-0.0330	-0.0289
<i>Swertia radiata</i>	Gentianaceae	11	-0.0245	-0.0538	-0.0337	-0.0212	-0.0251	0.1911	0.1982	0.0332	-0.0213	-0.0289
<i>Swertia utahensis</i> *‡	Gentianaceae	2	-0.0154	0.0028	-0.0200	-0.0045	-0.0140	-0.0212	-0.0196	-0.0179	-0.0139	-0.0121
<i>Symphoricarpos oreophilus</i>	Caprifoliaceae	58	-0.0154	-0.1199	-0.0154	-0.0492	-0.0478	0.1381	0.1700	0.2005	-0.0018	-0.0198
<i>Talinum parviflorum</i> *	Portulacaceae	1	-0.0109	-0.0111	-0.0142	0.1325	0.0441	-0.0033	-0.0045	-0.0055	-0.0098	-0.0085
<i>Tamarix chinensis</i> €	Tamaricaceae	5	-0.0245	-0.0253	-0.0696	-0.0476	-0.0322	-0.0559	-0.0489	0.0066	0.0266	0.4269
<i>Taraxacum officinale</i> €	Compositae	11	0.0097	-0.0636	-0.0846	-0.0492	0.0331	-0.0170	-0.0108	0.2013	0.2364	0.1004
<i>Tetradymia canescens</i>	Compositae	9	-0.0062	-0.0212	0.0466	0.0890	-0.0134	-0.0331	0.0013	-0.0027	-0.0185	-0.0260
<i>Thalictrum fendleri</i>	Ranunculaceae	12	-0.0057	-0.0697	-0.0848	-0.0473	0.0290	0.0072	-0.0048	0.1990	0.2305	0.0461
<i>Thelypodium laxiflorum</i> *	Cruciferae	7	-0.0291	-0.0363	0.0363	-0.0046	0.0139	-0.0327	-0.0249	0.0083	0.1166	0.0474
<i>Thermopsis montana</i>	Leguminosae	4	-0.0219	-0.0411	-0.0546	-0.0139	-0.0028	0.0401	0.0435	0.0581	0.0169	0.0308
<i>Thysanocarpus curvipes</i> *	Cruciferae	10	-0.0252	0.0346	0.0630	0.0645	0.0103	-0.0466	-0.0401	-0.0296	-0.0253	-0.0116
<i>Townsendia incana</i> *	Compositae	1	-0.0109	0.0220	-0.0290	-0.0240	-0.0163	-0.0248	-0.0098	-0.0127	-0.0098	0.0144

Scientific Name	Family	#Occ.	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<i>Townsendia montana</i> *†‡‡	Compositae	1	-0.0109	-0.0106	0.0414	0.1042	-0.0083	-0.0248	-0.0119	0.0007	-0.0098	-0.0085
<i>Toxicodendron rydbergii</i>	Anacardiaceae	2	-0.0154	-0.0260	-0.0414	-0.0340	-0.0108	-0.0053	-0.0030	0.0161	0.1317	0.0538
<i>Tradescantia occidentalis</i>	Commelinaceae	22	-0.0477	0.0149	0.0578	0.2178	-0.0173	-0.0627	-0.0540	-0.0375	-0.0429	-0.0383
<i>Tragopogon dubius</i> €	Compositae	8	0.2444	-0.0352	-0.0837	-0.0421	-0.0323	-0.0355	0.0085	0.0961	-0.0101	0.0077
<i>Tribulus terrestris</i> €	Zygophyllaceae	2	0.1011	-0.0230	-0.0522	-0.0290	-0.0196	-0.0352	-0.0308	-0.0179	-0.0068	0.0310
<i>Trifolium longipes</i>	Leguminosae	10	-0.0014	-0.0608	-0.0552	0.0103	0.2031	-0.0044	0.0434	0.0456	0.0265	0.0045
<i>Trifolium</i> sp.	Leguminosae	2	-0.0154	-0.0161	-0.0072	-0.0228	-0.0196	-0.0282	-0.0167	0.0097	0.0194	0.2472
<i>Typha latifolia</i>	Typhaceae	1	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	0.0122	0.0117	0.1572
<i>Valeriana occidentalis</i> *†‡‡	Valerianaceae	1	-0.0109	-0.0204	-0.0217	-0.0092	0.1027	0.0191	0.0034	-0.0127	0.0353	-0.0085
<i>Verbascum thapsus</i> €	Scrophulariaceae	5	-0.0103	-0.0370	-0.0698	-0.0372	-0.0105	-0.0340	-0.0374	0.1451	0.1323	0.3743
<i>Vicia americana</i>	Leguminosae	43	-0.0117	-0.0951	-0.0621	-0.0652	-0.0499	0.0806	0.1074	0.2497	0.0347	0.0130
<i>Vicia ludoviciana</i> *	Leguminosae	1	0.0196	0.0550	-0.0107	-0.0177	-0.0163	-0.0248	-0.0217	-0.0127	-0.0098	-0.0085
<i>Viguiera multiflora</i>	Compositae	12	0.0440	-0.0363	-0.0190	-0.0051	-0.0318	0.0174	0.0916	0.0675	-0.0243	-0.0212
<i>Viola canadensis</i> *	Violaceae	5	-0.0245	-0.0441	-0.0655	-0.0388	0.0225	-0.0120	-0.0347	0.0734	0.3300	0.0892
<i>Viola nuttallii</i> *‡	Violaceae	1	-0.0109	-0.0133	-0.0368	-0.0149	-0.0163	-0.0150	-0.0040	0.0934	0.0023	0.0255
<i>Viola purpurea</i>	Violaceae	10	0.0039	-0.0474	-0.0220	-0.0334	-0.0446	0.2702	0.1282	-0.0322	-0.0314	-0.0275
<i>Vitis arizonica</i>	Vitaceae	1	-0.0109	-0.0162	-0.0290	-0.0240	-0.0163	-0.0150	-0.0138	0.0136	0.1491	0.0709
<i>Woodsia oregana</i> *	Polypodiaceae	1	-0.0109	-0.0162	0.0052	0.0023	0.0567	0.0140	-0.0138	-0.0004	0.0878	-0.0085
<i>Wyethia arizonica</i>	Compositae	5	-0.0245	-0.0350	-0.0428	0.0301	0.1696	-0.0313	-0.0292	-0.0253	-0.0192	-0.0192
<i>Xanthium strumarium</i> *	Compositae	1	-0.0109	-0.0117	-0.0232	-0.0169	-0.0113	-0.0248	-0.0217	0.0263	0.0371	0.3576
<i>Yucca angustissima</i> ⁴	Agavaceae	41	-0.0660	-0.0505	0.1162	0.1379	0.0550	-0.0337	-0.0226	-0.0629	-0.0344	-0.0521
<i>Yucca baccata</i>	Agavaceae	17	-0.0066	0.1913	0.0981	-0.0227	-0.0344	-0.0682	-0.0588	-0.0489	-0.0392	-0.0318
<i>Zauschneria latifolia</i>	Onagraceae	15	-0.0160	-0.0558	-0.0283	0.0005	0.2515	-0.0038	-0.0293	0.0217	0.1956	-0.0039
<i>Zigadenus paniculatus</i> *	Liliaceae	12	0.1141	-0.0079	-0.0120	-0.0498	-0.0432	0.0843	0.1945	-0.0016	-0.0320	-0.0302

*not recorded by the USGS-NPS vegetation survey at Zion N.P. (Cogan et al. 2004)

‡not mentioned in the Zion N.P. annotated checklist (Fertig and Alexander 2009)

†‡mentioned as unconfirmed in the Zion N.P. annotated checklist (Fertig and Alexander 2009)

‡‡ mentioned as falsely reported or questionable in the Zion N.P. annotated checklist (Fertig and Alexander 2009)

€ exotic taxa (Welsh et al. 2003, Fertig and Alexander 2009)

¹Probably equivalent to *Artemisia nova* in Cogan et al. 2004; described as *A. nova* var. *arbuscula* in Fertig and Alexander 2009

²Probably equivalent to *Elymus lanceolatus* as well as *Pascopyrum smithii* in Cogan et al. 2004

³Includes *Gutierrezia microcephala* and *G. sarothrae*

⁴Probably equivalent to *Yucca elata* var. *utahensis* in Cogan et al. 2004

APPENDIX 2.3

Modal species of vegetation types described by Harper et al. (2001) for Zion National Park, Utah (rows) and their affinities to coalition species groups of the current study (columns). Following Curtis (1959), modal species are prevalent (frequently-occurring) species assigned to the vegetation type where their percent occurrence is highest. Affinities are mean pairwise association values (in units of the phi coefficient) between a species and members of a coalition group. Species are listed in descending order of percent occurrence (percent of plots of indicated vegetation plot where species was recorded). See Table 2.3 for mean affinity values by vegetation type.

Vegetation Type	Species	% Occ	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
Abandoned Fields	<i>Bromus tectorum</i>	100	0.1156	0.2135	0.0710	-0.0590	-0.1214	-0.1832	-0.1512	-0.1083	-0.0873	0.0262
Abandoned Fields	<i>Chamaesyce albomarginata</i>	67	0.2085	0.0081	-0.0514	-0.0601	-0.0683	0.0486	0.0045	-0.0518	-0.0427	-0.0065
Abandoned Fields	<i>Sporobolus cryptandrus</i>	67	0.0570	0.0424	0.0237	0.1236	-0.0302	-0.1019	-0.0813	-0.0708	-0.0588	-0.0061
Abandoned Fields	<i>Tragopogon dubius</i>	67	0.2444	-0.0352	-0.0837	-0.0421	-0.0323	-0.0355	0.0085	0.0961	-0.0101	0.0077
Abandoned Fields	<i>Erodium cicutarium</i>	50	0.2865	0.1277	-0.0910	-0.0748	-0.0488	-0.0766	-0.0609	-0.0407	-0.0282	-0.0211
Abandoned Fields	<i>Lactuca serriola</i>	50	0.2237	-0.0045	-0.0836	-0.0662	-0.0233	-0.0521	-0.0387	0.0166	0.0243	0.0185
Abandoned Fields	<i>Astragalus nuttallianus</i>	40	0.0317	0.2167	-0.0609	-0.0709	-0.0475	-0.0755	-0.0594	-0.0385	-0.0297	-0.0260
Abandoned Fields	<i>Aristida purpurea</i>	33	0.0216	0.1291	0.0011	-0.0051	-0.0026	-0.0986	-0.0888	-0.0646	-0.0393	0.0031
Abandoned Fields	<i>Artemisia dracunculus</i>	33	0.0695	-0.0227	-0.0022	-0.0069	-0.0291	-0.0208	-0.0178	-0.0225	-0.0260	-0.0138
Abandoned Fields	<i>Atriplex canescens</i>	33	0.2722	0.0716	-0.0649	-0.0708	-0.0447	-0.0797	-0.0634	-0.0407	-0.0314	-0.0275
Abandoned Fields	<i>Lycium pallidum</i>	33	0.3355	0.0067	-0.0506	-0.0376	-0.0255	-0.0431	-0.0263	-0.0220	-0.0170	-0.0149
Abandoned Fields	<i>Sphaeralcea grossulariifolia</i>	33	0.2053	0.0710	-0.0075	-0.0691	-0.0565	-0.0771	-0.0643	-0.0423	-0.0347	-0.0224
Blackbrush	<i>Coleogyne ramosissima</i>	100	0.0145	0.2448	-0.0393	-0.0830	-0.0601	-0.0914	-0.0800	-0.0466	-0.0360	-0.0315
Blackbrush	<i>Gilia inconspicua</i>	90	-0.0137	0.2558	0.1513	0.0515	-0.1029	-0.1116	-0.0963	-0.1140	-0.0969	-0.0584
Blackbrush	<i>Bromus rubens</i>	80	0.0339	0.3680	0.0731	-0.0990	-0.0855	-0.1439	-0.1239	-0.0865	-0.0642	-0.0544
Blackbrush	<i>Gutierrezia microcephala</i> ¹	80	0.0989	0.2817	0.0670	-0.0696	-0.0984	-0.1590	-0.1114	-0.1043	-0.0962	-0.0086
Blackbrush	<i>Festuca octoflora</i>	70	0.0169	0.2480	0.0657	0.0993	-0.0069	-0.1467	-0.1123	-0.0946	-0.0819	-0.0469
Blackbrush	<i>Hilaria</i> sp.	60	0.1280	0.2507	0.0402	-0.1195	-0.0871	-0.1262	-0.1138	-0.0683	-0.0675	-0.0510

Vegetation Type	Species	% Occ	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
Blackbrush (cont.)	<i>Cryptantha</i> sp. (annual)	50	-0.0026	0.2085	0.1317	0.0211	-0.0596	-0.1192	-0.0796	-0.0746	-0.0776	-0.0639
Blackbrush	<i>Draba verna</i>	50	0.0217	0.2595	0.0867	-0.0141	-0.0704	-0.0816	-0.0745	-0.0737	-0.0615	-0.0189
Blackbrush	<i>Eriogonum palmerianum</i>	50	0.0092	0.2175	0.0316	-0.0719	-0.0476	-0.0797	-0.0779	-0.0503	-0.0369	-0.0339
Blackbrush	<i>Calochortus flexuosus</i>	40	0.0171	0.1303	-0.0365	-0.0562	-0.0395	-0.0626	-0.0550	-0.0302	-0.0261	-0.0160
Blackbrush	<i>Descurainia pinnata</i>	40	-0.0037	0.1716	0.0713	-0.0494	-0.0819	-0.0556	-0.0588	0.0116	-0.0520	-0.0214
Blackbrush	<i>Ephedra nevadensis</i>	40	0.0478	0.2637	0.0091	-0.0872	-0.0630	-0.0888	-0.0854	-0.0554	-0.0394	-0.0374
Blackbrush	<i>Stipa hymenoides</i>	40	-0.0221	0.0813	0.0886	0.0327	-0.0549	-0.0027	-0.0408	-0.0788	-0.0721	-0.0355
Blackbrush	<i>Oenothera pallida</i>	33	-0.0224	0.0081	0.0494	0.1056	-0.0116	-0.0433	-0.0436	-0.0284	-0.0092	0.1363
Blackbrush	<i>Eriastrum eremicum</i>	30	-0.0062	0.2058	0.0095	-0.0395	-0.0417	-0.0626	-0.0550	-0.0339	-0.0261	-0.0229
Blackbrush	<i>Poa bigelovii</i>	30	0.0487	0.1065	-0.0240	-0.0542	-0.0388	-0.0675	-0.0593	-0.0329	-0.0280	-0.0181
Blackbrush	<i>Psorothamnus fremontii</i>	30	0.0240	0.2031	-0.0107	-0.0679	-0.0532	-0.0747	-0.0660	-0.0428	-0.0330	-0.0289
Sagebrush	<i>Artemisia tridentata</i>	100	0.0211	0.0237	0.0140	0.0087	-0.0941	-0.0650	-0.0307	-0.0021	-0.0710	-0.0200
Sagebrush	<i>Gutierrezia sarothrae</i> ¹	71	0.0989	0.2817	0.0670	-0.0696	-0.0984	-0.1590	-0.1114	-0.1043	-0.0962	-0.0086
Sagebrush	<i>Elymus elymoides</i>	57	0.0162	0.0923	0.0099	0.0133	0.0141	-0.0536	-0.0274	-0.0206	-0.0247	-0.0314
Sagebrush	<i>Chrysothamnus nauseosus</i>	43	-0.0081	0.0136	0.0303	0.1202	-0.0282	-0.0941	-0.0726	-0.0711	-0.0435	0.0411
Sagebrush	<i>Eriastrum sparsiflorum</i>	29	0.0561	0.1752	-0.0212	-0.0568	-0.0495	-0.0914	-0.0710	-0.0466	-0.0360	-0.0250
Sagebrush	<i>Erigeron divergens</i>	29	-0.0097	-0.0041	0.0203	-0.0308	-0.0587	-0.0115	0.0173	0.0552	-0.0393	-0.0170
Sagebrush	<i>Opuntia phaeacantha</i>	29	0.1030	0.0449	0.0187	0.0060	-0.0339	-0.0589	-0.0420	-0.0291	-0.0297	-0.0200
Sagebrush	<i>Pectocarya setosa</i>	29	0.0226	0.0666	-0.0321	-0.0339	-0.0255	-0.0431	-0.0263	-0.0220	-0.0170	-0.0149
Juniper-Pinyon	<i>Juniperus osteosperma</i>	82	0.0251	0.1706	0.1868	-0.0462	-0.1486	0.0226	0.0318	-0.1258	-0.1273	-0.1076
Juniper-Pinyon	<i>Amelanchier utahensis</i>	65	-0.0900	-0.0968	0.1661	0.0340	-0.0081	0.1990	0.1598	-0.0446	-0.0817	-0.1249
Juniper-Pinyon	<i>Arabis perennans</i>	57	-0.0953	0.0295	0.2379	0.0173	0.0178	0.0472	0.0240	-0.0499	-0.0301	-0.0931
Juniper-Pinyon	<i>Pinus monophylla</i>	53	-0.0639	0.1617	0.1964	-0.0616	-0.1071	-0.1054	-0.0930	-0.0808	-0.0790	-0.0597
Juniper-Pinyon	<i>Pinus edulis</i>	34	-0.0363	-0.0799	0.0969	0.0150	-0.0519	0.2160	0.1513	-0.0540	-0.0481	-0.0640
Juniper-Pinyon	<i>Streptanthus cordatus</i>	31	-0.0361	0.0388	0.1502	-0.0189	-0.0797	0.0734	0.0374	-0.0617	-0.0623	-0.0533
Juniper-Pinyon	<i>Erigeron sionis</i>	29	-0.0219	-0.0387	-0.0358	0.0211	0.2217	0.0074	-0.0145	0.0187	0.1662	0.0426
Mountain Brush	<i>Eriogonum racemosum</i>	33	-0.0291	-0.0898	0.0990	0.1001	0.0704	-0.0038	0.0302	0.0394	0.0517	-0.0253
Mountain Brush	<i>Erysimum asperum</i>	32	-0.0775	-0.0340	0.2064	0.1601	-0.0076	0.0100	0.0369	-0.0315	-0.0287	-0.0527
Mountain Brush	<i>Machaeranthera canescens</i>	32	-0.0036	0.0284	0.0883	0.1432	-0.0142	-0.0414	-0.0210	-0.0066	-0.0534	-0.0318
Mountain Brush	<i>Phacelia heterophylla</i>	32	-0.0498	-0.0651	0.0028	0.0973	0.0148	-0.0231	-0.0256	0.1026	0.0676	0.1188

Vegetation Type	Species	% Occ	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
Rock Crevice	<i>Cercocarpus intricatus</i>	84	-0.0741	-0.0732	-0.0008	0.0948	0.3022	-0.0254	-0.0425	-0.0540	0.0241	-0.0535
Rock Crevice	<i>Arenaria fendleri</i>	64	-0.0544	-0.0513	-0.0408	0.1880	0.2407	-0.0605	-0.0522	-0.0512	-0.0203	-0.0427
Rock Crevice	<i>Chrysopsis villosa</i>	64	-0.0433	-0.0529	0.0206	0.1933	0.1966	-0.0952	-0.0770	-0.0592	0.0364	0.0487
Rock Crevice	<i>Castilleja scabrida</i>	60	-0.0392	-0.0544	-0.0185	0.0831	0.2903	-0.0401	-0.0609	-0.0449	0.0430	-0.0458
Rock Crevice	<i>Opuntia macrorhiza</i>	56	-0.0871	0.0051	0.1969	0.2100	0.0551	0.0113	0.0278	-0.0893	-0.0880	-0.1079
Rock Crevice	<i>Stephanomeria tenuifolia</i>	48	-0.0564	-0.0310	0.0846	0.1172	0.2097	-0.0459	-0.0537	-0.0463	0.0698	-0.0371
Rock Crevice	<i>Penstemon laevis</i>	40	-0.0532	-0.0162	0.0773	0.2316	0.0846	-0.0416	-0.0335	-0.0542	-0.0342	-0.0456
Rock Crevice	<i>Chaenactis douglasii</i>	36	-0.0625	-0.0719	0.1114	0.2527	0.1140	-0.0008	0.0032	-0.0622	-0.0409	-0.0586
Rock Crevice	<i>Cryptantha confertiflora</i>	36	-0.0351	0.0704	0.1252	0.0887	0.0972	-0.0691	-0.0637	-0.0683	-0.0403	-0.0469
Rock Crevice	<i>Penstemon humilis</i>	36	-0.0593	-0.0565	0.0790	0.1052	0.2021	-0.0007	-0.0125	-0.0374	0.0411	-0.0438
Rock Crevice	<i>Quercus turbinella</i>	32	-0.0617	0.1136	0.2131	0.0456	-0.0280	-0.0980	-0.0825	-0.0805	-0.0597	-0.0524
Rock Crevice	<i>Erigeron canaani</i>	28	-0.0448	-0.0719	-0.0209	0.1198	0.2338	0.0168	-0.0131	-0.0192	0.0259	-0.0316
Rock Crevice	<i>Penstemon rostriflorus</i>	28	-0.0179	-0.0415	0.0496	-0.0137	0.1297	0.0101	-0.0091	0.0460	0.1111	-0.0184
Ponderosa Pine	<i>Pinus ponderosa</i>	97	-0.0574	-0.1236	-0.0675	0.1735	0.1993	0.0611	0.0717	-0.0050	0.0285	-0.0322
Ponderosa Pine	<i>Poa fendleriana</i>	85	-0.1391	-0.0620	0.2132	0.1283	0.1186	0.0577	0.0891	-0.0417	-0.0072	-0.0837
Ponderosa Pine	<i>Arctostaphylos patula</i>	61	-0.1013	-0.0593	0.1132	0.2147	0.0767	0.0893	0.0301	-0.0959	-0.0837	-0.0949
Ponderosa Pine	<i>Senecio multilobatus</i>	52	-0.0802	-0.1087	0.1615	0.1919	0.1136	0.0662	0.0478	-0.0288	-0.0054	-0.0606
Ponderosa Pine	<i>Phlox austromontana</i>	39	-0.0767	-0.0520	0.1251	0.1886	0.0721	0.0491	0.0433	-0.0512	-0.0503	-0.0675
Ponderosa Pine	<i>Polygonum douglasii</i>	33	-0.0015	-0.0693	-0.0321	0.1548	0.0311	0.0050	0.0855	0.1711	-0.0403	-0.0010
Ponderosa Pine	<i>Purshia tridentata</i>	30	-0.0398	-0.0642	0.0241	0.0959	-0.0355	0.1318	0.1832	-0.0297	-0.0563	-0.0475
Ponderosa Pine	<i>Arenaria macradenia</i>	24	-0.0804	-0.0607	0.1271	0.0990	0.1502	-0.0052	-0.0069	-0.0401	-0.0037	-0.0588
Ponderosa Pine	<i>Comandra umbellata</i>	24	-0.0686	-0.0696	0.0777	0.1314	0.0282	0.0257	0.0821	-0.0146	-0.0470	-0.0505
Ponderosa Pine	<i>Yucca angustissima</i>	24	-0.0660	-0.0505	0.1162	0.1379	0.0550	-0.0337	-0.0226	-0.0629	-0.0344	-0.0521
Douglas Fir-White Fir	<i>Quercus gambelii</i>	86	-0.0835	-0.2123	-0.0259	0.0354	0.0422	0.1792	0.1804	0.1418	0.0666	-0.0480
Douglas Fir-White Fir	<i>Pachystima myrsinoides</i>	71	-0.0573	-0.1407	-0.0118	0.0118	0.0706	0.2096	0.1310	0.0248	0.1121	-0.0386
Douglas Fir-White Fir	<i>Carex rossii</i>	57	-0.0335	-0.1431	-0.0482	0.0505	0.1222	0.2114	0.1553	0.1027	0.0042	-0.0554
Douglas Fir-White Fir	<i>Abies concolor</i>	52	-0.0226	-0.0893	-0.0770	-0.0559	0.0865	0.0603	0.0229	0.1859	0.2236	0.0410
Douglas Fir-White Fir	<i>Symphoricarpos oreophilus</i>	52	-0.0154	-0.1199	-0.0154	-0.0492	-0.0478	0.1381	0.1700	0.2005	-0.0018	-0.0198
Douglas Fir-White Fir	<i>Pseudotsuga menziesii</i>	48	-0.0476	-0.0849	-0.0415	-0.0183	0.1328	0.0553	0.0042	0.0309	0.2559	0.0212
Douglas Fir-White Fir	<i>Vicia americana</i>	48	-0.0117	-0.0951	-0.0621	-0.0652	-0.0499	0.0806	0.1074	0.2497	0.0347	0.0130

Vegetation Type	Species	% Occ	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
Douglas Fir-White Fir (cont.)	<i>Acer grandidentatum</i>	43	-0.0556	-0.0988	-0.0451	-0.0489	0.0587	0.0312	0.0003	0.1139	0.2614	0.0429
Douglas Fir-White Fir	<i>Draba asprella</i>	38	-0.0516	-0.0823	0.0124	0.0971	0.2068	0.0802	0.0067	-0.0115	0.1409	-0.0142
Douglas Fir-White Fir	<i>Haplopappus scopolorum</i>	38	-0.0620	-0.0438	0.1804	0.0711	0.0046	0.0255	0.0324	-0.0509	-0.0055	-0.0515
Douglas Fir-White Fir	<i>Mahonia repens</i>	38	-0.0231	-0.0947	-0.0511	-0.0508	-0.0046	0.1614	0.0969	0.0752	0.1310	0.0000
Douglas Fir-White Fir	<i>Holodiscus dumosus</i>	33	-0.0356	-0.0921	-0.0438	0.0077	0.3078	-0.0046	-0.0308	-0.0027	0.1947	-0.0222
Douglas Fir-White Fir	<i>Fraxinus anomala</i>	29	-0.0588	0.0431	0.2408	-0.0204	0.0183	0.0824	0.0020	-0.0571	-0.0092	-0.0520
Douglas Fir-White Fir	<i>Heuchera rubescens</i>	29	-0.0292	-0.0709	-0.0461	-0.0292	0.1744	0.0382	-0.0019	0.0476	0.2533	-0.0007
Douglas Fir-White Fir	<i>Stellaria jamesiana</i>	29	0.0048	-0.0822	-0.1064	-0.0712	-0.0308	0.0271	0.0497	0.3314	0.0534	0.0380
Douglas Fir-White Fir	<i>Juniperus scopulorum</i>	24	-0.0178	-0.0775	-0.0461	-0.0170	-0.0283	0.0539	0.0409	0.1754	0.0134	0.0109
Hanging Gardens	<i>Adiantum capillus-veneris</i>	89	-0.0154	-0.0290	-0.0466	-0.0340	-0.0028	-0.0352	-0.0308	0.0132	0.1540	-0.0025
Hanging Gardens	<i>Calamagrostis scopulorum</i>	79	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	0.0179	-0.0085
Hanging Gardens	<i>Aster welshii</i>	72	-0.0269	-0.0449	-0.0609	-0.0507	-0.0144	-0.0372	-0.0375	0.0224	0.1633	0.1134
Hanging Gardens	<i>Lobelia cardinalis</i> ²	56	--	--	--	--	--	--	--	--	--	--
Hanging Gardens	<i>Aquilegia chrysanthia</i>	50	-0.0109	-0.0204	-0.0368	-0.0240	-0.0163	-0.0248	-0.0217	-0.0127	0.0179	-0.0085
Hanging Gardens	<i>Aquilegia formosa</i>	50	-0.0189	-0.0355	-0.0509	-0.0269	0.0919	-0.0250	-0.0232	-0.0163	0.0585	-0.0070
Hanging Gardens	<i>Dodecatheon pulchellum</i>	44	-0.0154	-0.0290	-0.0469	-0.0340	-0.0108	-0.0123	-0.0086	-0.0025	0.0387	-0.0025
Hanging Gardens	<i>Apocynum androsaemifolium</i>	40	-0.0154	-0.0290	-0.0414	-0.0189	0.0270	0.0152	-0.0030	0.0476	0.2110	0.0817
Hanging Gardens	<i>Aster glaucodes</i>	40	-0.0269	-0.0144	0.0115	-0.0389	-0.0091	-0.0054	-0.0173	0.0135	0.1160	0.1156
Hanging Gardens	<i>Eupatorium herbaceum</i>	40	-0.0081	-0.0709	-0.0670	-0.0391	0.1492	0.0054	-0.0365	0.0630	0.3612	0.0462
Hanging Gardens	<i>Solidago velutina</i>	40	0.0085	-0.1190	-0.0889	0.0046	0.1890	0.0614	0.1036	0.0889	0.1476	-0.0134
Hanging Gardens	<i>Erigeron utahensis</i>	39	-0.0341	0.1342	0.2246	0.0464	-0.0274	-0.0700	-0.0681	-0.0713	-0.0588	-0.0560
Hanging Gardens	<i>Petrosymum caespitosum</i>	39	-0.0385	-0.0612	-0.0582	0.0268	0.3143	0.0020	-0.0243	-0.0130	0.1274	0.0088
Hanging Gardens	<i>Muhlenbergia thurberi</i>	33	-0.0174	-0.0610	-0.0526	0.0809	0.2838	-0.0207	-0.0274	-0.0274	0.0811	-0.0182
Hanging Gardens	<i>Smilacina stellata</i>	33	-0.0189	-0.0355	-0.0640	-0.0334	-0.0215	0.0102	-0.0142	0.0284	0.0379	-0.0060
Hanging Gardens	<i>Agrostis stolonifera</i>	28	-0.0109	-0.0183	-0.0329	-0.0240	-0.0163	-0.0248	-0.0217	0.0068	0.0093	0.2268
Hanging Gardens	<i>Carex curvula</i> ²	28	--	--	--	--	--	--	--	--	--	--
Hanging Gardens	<i>Cirsium arizonicum</i>	26	0.0236	-0.0538	0.0075	0.0434	0.1052	0.0028	-0.0078	0.0262	0.0954	0.0911
Hanging Gardens	<i>Antennaria rosulata</i>	20	-0.0189	-0.0276	-0.0021	-0.0057	0.0238	0.0195	0.0148	0.0635	0.1447	0.0540

Vegetation Type	Species	% Occ	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
Riparian	<i>Mimulus cardinalis</i>	72	-0.0154	-0.0260	-0.0466	-0.0340	-0.0231	-0.0282	-0.0252	0.0007	0.1183	0.0442
Riparian	<i>Acer negundo</i>	63	-0.0226	-0.0711	-0.0929	-0.0678	0.0736	-0.0447	-0.0579	0.0499	0.3163	0.2513
Riparian	<i>Artemisia ludoviciana</i>	42	-0.0089	0.0007	0.1161	0.0261	0.0346	-0.0376	-0.0201	0.0195	0.0962	0.1026
Riparian	<i>Bromus diandrus</i>	42	0.0312	-0.0336	-0.0857	-0.0613	-0.0407	-0.0734	-0.0647	-0.0126	0.0071	0.3353
Riparian	<i>Bromus ciliatus</i>	37	0.0293	-0.0670	-0.0665	-0.0486	0.0757	-0.0176	-0.0460	0.0473	0.2602	0.0995
Riparian	<i>Senecio spartoides</i>	37	-0.0242	-0.0684	-0.0308	0.0456	0.0257	-0.0538	-0.0629	0.0085	0.1054	0.0200
Riparian	<i>Fraxinus velutina</i>	32	-0.0169	-0.0349	-0.0638	-0.0478	-0.0313	-0.0546	-0.0496	-0.0130	0.0263	0.1862
Riparian	<i>Populus fremontii</i>	32	-0.0139	-0.0177	-0.0756	-0.0534	-0.0362	-0.0613	-0.0487	0.0008	0.0149	0.3936
Riparian	<i>Equisetum hyemale</i>	26	0.0242	-0.0422	-0.0812	-0.0464	-0.0190	-0.0501	-0.0537	0.0372	0.1603	0.3744
Riparian	<i>Zauschneria latifolia</i>	26	-0.0160	-0.0558	-0.0283	0.0005	0.2515	-0.0038	-0.0293	0.0217	0.1956	-0.0039
Riparian	<i>Cystopteris fragilis</i>	21	0.0155	-0.0505	-0.0547	-0.0311	0.0833	-0.0160	-0.0366	0.0764	0.3184	0.0691
Riparian	<i>Poa pratensis</i>	21	-0.0045	-0.0598	-0.0991	-0.0276	-0.0409	-0.0455	-0.0258	0.2466	0.0391	0.2027
Riparian	<i>Tamarix chinensis</i>	21	-0.0245	-0.0253	-0.0696	-0.0476	-0.0322	-0.0559	-0.0489	0.0066	0.0266	0.4269
Riparian	<i>Thelypodium laxiflorum</i>	21	-0.0291	-0.0363	0.0363	-0.0046	0.0139	-0.0327	-0.0249	0.0083	0.1166	0.0474

¹The species of *Gutierrezia* listed in Harper et al. (2001) were lumped in the current analysis due to identification uncertainties

²*Carex curvata* and *Lobelia cardinalis* were listed in Harper et al. (2001) due to their inclusion of supplemental hanging garden data not used in the current analysis

APPENDIX 2.4

U.S. National Vegetation Classification System (NVCS) associations of Zion National Park, Utah as recognized by Cogan et al. (2004) (rows) with their compositional affinities to coalition species groups of the current study (columns). Affinity values are in units of the phi coefficient, calculated for each species in relation to each coalition group (see Appendix 2.2) and then averaged across associations in proportion to their species frequencies. Low values (green shading) indicate low correspondence between associations and coalition groups; high values (red shading) indicate high correspondence (shading is scaled separately for each column). Associations are grouped by physiognomic class and alliance; see also Table 2.4. Note that association names are truncated omitting physiognomic descriptors.

	Cluster ¹	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
I. FOREST											
White Fir Forest Alliance											
White Fir / Bigtooth Maple	CrvcC/HPltu	-0.0367	-0.0767	-0.0156	-0.0019	0.0521	0.0524	0.0347	0.0785	0.1288	0.0213
White Fir / Gambel Oak	CrvcC/HPltu	-0.0408	-0.0982	-0.0198	-0.0006	0.0440	0.0957	0.0788	0.1057	0.0897	-0.0068
White Fir / Greenleaf Manzanita	MsaTp	-0.0442	-0.0862	0.0015	0.0176	0.0408	0.1158	0.0961	0.0639	0.0486	-0.0254
White Fir / Mountain Snowberry	HPltu	-0.0206	-0.0714	-0.0223	-0.0081	0.0001	0.0594	0.0731	0.1542	0.0412	0.0061
Douglas-fir Forest Alliance											
Douglas-fir / Bigtooth Maple	CrvcC	-0.0368	-0.0773	-0.0487	-0.0182	0.0594	0.0435	0.0067	0.0481	0.1910	0.0634
Douglas-fir / Gambel Oak	CrvcC/HPltu	-0.0461	-0.0697	0.0161	0.0117	0.0477	0.0662	0.0436	0.0487	0.0855	-0.0095
Douglas-fir / Mountain Snowberry	CrvcC/HPltu	-0.0345	-0.0557	0.0114	0.0082	0.0298	0.0635	0.0536	0.0520	0.0551	0.0000
Bigtooth Maple Montane Forest Alliance											
Bigtooth Maple / Gambel Oak	CrvcC/HPltu	-0.0266	-0.0669	-0.0086	0.0090	0.0228	0.0316	0.0282	0.0782	0.0703	0.0274
Quaking Aspen Forest Alliance											
Quaking Aspen / Mountain Snowberry / Tall Forbs	HPltu	-0.0067	-0.0594	-0.0505	-0.0256	-0.0049	0.0256	0.0391	0.1860	0.0609	0.0457
Quaking Aspen Temporarily Flooded Forest Alliance											
Quaking Aspen / Gambel Oak / Mountain Snowberry	HPltu	0.0108	-0.0644	-0.0556	-0.0237	0.0001	0.0270	0.0497	0.1635	0.0604	0.0600
Fremont Cottonwood Temporarily Flooded Forest Alliance											
Fremont Cottonwood / Coyote Willow	Strmb	0.0077	0.0214	-0.0014	-0.0056	-0.0186	-0.0471	-0.0381	-0.0114	0.0069	0.1520
White Fir - Quaking Aspen Forest Alliance											
Quaking Aspen - White Fir / Kentucky Bluegrass Semi-natural	HPltu	0.0062	-0.0447	-0.0404	-0.0046	0.0024	0.0100	0.0221	0.1285	0.0457	0.0400
Quaking Aspen - White Fir / Mountain Snowberry	HPltu	-0.0183	-0.0708	-0.0318	-0.0087	0.0165	0.0517	0.0589	0.1266	0.0659	0.0104

	Cluster ¹	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
II. WOODLAND											
<u>Two-needle Pinyon - (Juniper species) Woodland Alliance</u>											
Two-needle Pinyon - Juniper species / Big Sagebrush	UpSnd/RckSl	-0.0224	0.0012	0.0675	0.0806	-0.0180	0.0112	0.0186	-0.0387	-0.0479	-0.0470
Two-needle Pinyon - Juniper species / Gambel Oak	MsaTp/RckSl	-0.0449	-0.0447	0.0714	0.0486	0.0052	0.0814	0.0761	-0.0107	-0.0215	-0.0477
Two-needle Pinyon - Juniper species / Mountain-mahogany	MsaTp/RckSl	-0.0374	-0.0230	0.0898	0.0229	-0.0176	0.1039	0.0808	-0.0317	-0.0461	-0.0622
Two-needle Pinyon - Utah Juniper / Greenleaf Manzanita	MsaTp/RckSl	-0.0542	-0.0407	0.0953	0.0627	0.0079	0.0983	0.0780	-0.0313	-0.0351	-0.0663
Two-needle Pinyon - Utah Juniper / Littleleaf Mtn-mahogany	UpSnd/RckSl	-0.0395	0.0053	0.1013	0.0582	0.0180	0.0221	0.0184	-0.0448	-0.0344	-0.0436
Two-needle Pinyon - Utah Juniper / Stansbury Cliff-rose	MsaTp/RckSl	-0.0407	0.0065	0.0965	0.0272	-0.0123	0.0577	0.0410	-0.0386	-0.0391	-0.0528
Two-needle Pinyon / Curl-leaf Mountain-mahogany	MsaTp	-0.0329	-0.0587	0.0439	0.0305	-0.0002	0.0817	0.0847	0.0195	-0.0217	-0.0407
<u>Ponderosa Pine Woodland Alliance</u>											
Ponderosa Pine / Black Sagebrush	MsaTp	-0.0296	-0.0548	0.0250	0.0294	0.0106	0.0564	0.0747	0.0357	-0.0105	-0.0261
Ponderosa Pine / Gambel Oak	MsaTp	-0.0295	-0.0601	0.0136	0.0478	0.0331	0.0531	0.0639	0.0427	0.0121	-0.0214
Ponderosa Pine / Greenleaf Manzanita	MsaTp/RckSl	-0.0408	-0.0507	0.0357	0.0660	0.0309	0.0768	0.0651	-0.0049	-0.0160	-0.0431
Ponderosa Pine / Northern Bracken	CrvcC/HPltu	-0.0297	-0.0532	-0.0254	0.0398	0.0498	0.0049	0.0077	0.0707	0.0378	0.0218
Ponderosa Pine / Smooth Brome Semi-natural	CrvcC/HPltu	-0.0197	-0.0553	-0.0214	0.0371	0.0429	0.0145	0.0337	0.0901	0.0337	0.0368
<u>Utah Juniper Woodland Alliance</u>											
Utah Juniper / Big Sagebrush	ArdLw	0.0212	0.1489	0.0947	0.0031	-0.0698	-0.0558	-0.0336	-0.0754	-0.0844	-0.0563
<u>Singleleaf Pinyon - (Utah Juniper) Woodland Alliance</u>											
Singleleaf Pinyon - Utah Juniper / (Rd. Buffalob., Ut. Serviceb.)	RckSl	-0.0236	0.0722	0.1224	0.0204	-0.0195	-0.0223	-0.0156	-0.0549	-0.0480	-0.0474
Singleleaf Pinyon - Utah Juniper / Big Sagebrush	ArdLw	0.0131	0.1264	0.0825	-0.0170	-0.0564	-0.0555	-0.0398	-0.0604	-0.0604	-0.0389
Singleleaf Pinyon - Utah Juniper / Black Sagebrush	RckSl	-0.0010	0.0719	0.0775	-0.0188	-0.0448	-0.0180	0.0019	-0.0356	-0.0543	-0.0459
Singleleaf Pinyon - Utah Juniper / Blackbrush	ArdLw	0.0036	0.1322	0.0856	-0.0079	-0.0693	-0.0479	-0.0263	-0.0647	-0.0696	-0.0471
Singleleaf Pinyon - Utah Juniper / Mtn-mahogany - Gambel Oak	MsaTp/RckSl	-0.0517	-0.0051	0.1151	0.0158	-0.0391	0.0742	0.0598	-0.0393	-0.0561	-0.0758
Singleleaf Pinyon - Utah Juniper / Sparse Understory	RckSl	-0.0152	0.0704	0.1077	0.0154	-0.0413	-0.0129	-0.0002	-0.0555	-0.0598	-0.0531
Singleleaf Pinyon - Utah Juniper / Turbinella Live Oak	RckSl	-0.0164	0.0753	0.1121	0.0272	-0.0245	-0.0365	-0.0259	-0.0569	-0.0490	-0.0373
<u>Rocky Mountain Juniper Woodland Alliance</u>											
Rocky Mountain Juniper - Gambel Oak	--	-0.0309	-0.0325	0.0252	0.0206	0.0073	0.0133	0.0170	0.0282	0.0380	0.0585
<u>Box-elder Temporarily Flooded Woodland Alliance</u>											
Box-elder / Disturbed Understory	Strmb/HPltu	0.0062	-0.0197	-0.0053	0.0110	-0.0027	-0.0242	-0.0165	0.0434	0.0480	0.0505
Box-elder / Tasselflower Brickelbush	CrvcC	-0.0029	-0.0499	-0.0443	-0.0152	0.0506	-0.0146	-0.0206	0.0869	0.1435	0.0664

	Cluster ¹	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
II. WOODLAND (cont.)											
Fremont Cottonwood Temporarily Flooded Woodland Alliance											
Fremont Cottonwood - Velvet Ash	Strmb	-0.0040	-0.0014	-0.0135	-0.0234	-0.0161	-0.0414	-0.0400	-0.0128	0.0370	0.1710
Fremont Cottonwood / Emory Seepwillow	Strmb	-0.0036	0.0135	0.0023	0.0057	-0.0152	-0.0474	-0.0355	-0.0098	0.0046	0.1466
Single-leaf Ash Temporarily Flooded Woodland Alliance											
Single-leaf Ash	RckSl	-0.0310	0.0288	0.0902	0.0149	-0.0133	-0.0076	-0.0074	-0.0109	-0.0067	-0.0141
III. SHRUBLAND											
Greenleaf Manzanita Shrubland Alliance											
Greenleaf Manzanita - Gambel Oak - (Ut. Serviceberry)	MsaTp/RckSl	-0.0488	-0.0501	0.0729	0.0715	0.0263	0.0718	0.0665	-0.0080	-0.0201	-0.0521
Greenleaf Manzanita - Mountain Big Sagebrush	UpSnd	-0.0460	-0.0387	0.0579	0.1264	0.0219	0.0062	0.0230	-0.0183	-0.0314	-0.0416
Greenleaf Manzanita	UpSnd/RckSl	-0.0444	-0.0437	0.0560	0.0912	0.0425	0.0383	0.0363	-0.0125	-0.0169	-0.0399
Mexican Manzanita Shrubland Alliance											
Mexican Manzanita	MsaTp	-0.0425	-0.0717	0.0225	0.0303	0.0317	0.0921	0.0651	0.0242	0.0246	0.0039
Turbinella Live Oak Shrubland Alliance											
Turbinella Live Oak - (Utah Serviceberry) Colluvial	RckSl	-0.0223	0.0281	0.0948	0.0332	-0.0017	-0.0055	-0.0012	-0.0287	-0.0253	-0.0274
Arrow-weed Seasonally Flooded Shrubland Alliance											
Arrow-weed Seasonally Flooded	LwFlt/ArdLw	0.0752	0.0821	0.0137	-0.0194	-0.0465	-0.0718	-0.0485	-0.0096	-0.0181	0.0424
Big Sagebrush Shrubland Alliance											
Big Sagebrush - (Rubber Rabbitbrush) / Cheatgrass Semi-natural	ArdLw	0.0364	0.0781	0.0419	0.0151	-0.0469	-0.0660	-0.0445	-0.0430	-0.0477	-0.0074
Big Sagebrush / Blue Grama	UpSnd	-0.0199	-0.0280	0.0227	0.0956	-0.0022	-0.0099	0.0041	0.0164	-0.0227	-0.0190
Basin Big Sagebrush Shrubland Alliance											
Basin Big Sagebrush / Western Wheatgrass - (Str. Wild Rye)	MsaTp	0.0223	-0.0711	-0.0104	-0.0127	-0.0312	0.0902	0.0811	0.0420	-0.0170	-0.0268
Mountain Big Sagebrush Shrubland Alliance											
Mountain Big Sagebrush / Needle-and-Thread	MsaTp/RckSl	0.0025	-0.0072	0.0392	0.0313	-0.0221	0.0264	0.0523	0.0116	-0.0272	-0.0307
(Stansbury Cliff-rose, Mexican Cliff-rose) Shrubland Alliance											
Stansbury Cliff-rose - Greenleaf Manzanita	UpSnd/RckSl	-0.0464	-0.0157	0.0903	0.0913	0.0418	0.0244	0.0246	-0.0432	-0.0422	-0.0622
Rubber Rabbitbrush Shrubland Alliance											
Rubber Rabbitbrush / Cheatgrass Semi-natural	RckSl/UpSnd	0.0036	0.0489	0.0925	0.0697	-0.0269	-0.0668	-0.0495	-0.0707	-0.0537	-0.0201

	Cluster ¹	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
III. SHRUBLAND (cont.)											
<u>Sand Sagebrush Shrubland Alliance</u>											
Sand Sagebrush Colorado Plateau	ArdLw	0.0276	0.0861	0.0369	0.0103	-0.0316	-0.0690	-0.0525	-0.0511	-0.0421	-0.0004
<u>Nevada Joint-fir Shrubland Alliance</u>											
Nevada Joint-fir Basalt	LwFlt/ArdLw	0.0961	0.1486	0.0207	-0.0204	-0.0509	-0.0852	-0.0684	-0.0517	-0.0405	-0.0139
<u>Mormon-tea Shrubland Alliance</u>											
Gray Horsebrush - Mormon-tea	UpSnd	-0.0231	0.0029	0.0589	0.0993	0.0032	-0.0324	-0.0125	-0.0380	-0.0328	-0.0212
<u>Blackbrush Shrubland Alliance</u>											
Blackbrush / James' Galleta	ArdLw	0.0477	0.2037	0.0586	-0.0488	-0.0706	-0.0861	-0.0701	-0.0675	-0.0653	-0.0432
Blackbrush	ArdLw	0.0591	0.1692	0.0466	-0.0290	-0.0600	-0.0745	-0.0595	-0.0695	-0.0605	-0.0327
<u>Fourwing Saltbush Shrubland Alliance</u>											
Fourwing Saltbush - Big Sagebrush	LwFlt/ArdLw	0.0954	0.1107	0.0064	-0.0267	-0.0576	-0.0758	-0.0631	-0.0460	-0.0461	-0.0115
Fourwing Saltbush	LwFlt/ArdLw	0.0864	0.1379	0.0241	-0.0258	-0.0509	-0.0783	-0.0648	-0.0620	-0.0524	-0.0251
<u>Mountain Snowberry Shrubland Alliance</u>											
Mountain Snowberry / Kentucky Bluegrass Semi-natural	HPltu	0.0232	-0.0419	-0.0253	0.0102	-0.0092	0.0036	0.0253	0.1238	0.0096	0.0214
<u>Utah Serviceberry Shrubland Alliance</u>											
Utah Serviceberry Shrubland	HPltu	-0.0105	-0.0158	0.0184	-0.0081	-0.0245	0.0201	0.0253	0.0651	0.0168	0.0181
<u>Gambel Oak Shrubland Alliance</u>											
Gambel Oak - Mountain-mahogany / (Geyer's Sedge)	MsaTp/RckSI	-0.0360	-0.0304	0.0604	0.0140	-0.0058	0.0699	0.0632	0.0114	-0.0143	-0.0427
Gambel Oak / Big Sagebrush	UpSnd/RckSI	-0.0139	-0.0193	0.0503	0.0458	-0.0108	0.0085	0.0166	0.0083	-0.0183	-0.0196
Gambel Oak / Mountain Snowberry	HPltu	-0.0133	-0.0650	-0.0049	0.0096	-0.0003	0.0521	0.0683	0.1088	0.0207	0.0038
Gambel Oak / Muttongrass	UpSnd/RckSI	-0.0330	-0.0376	0.0558	0.0686	0.0304	0.0241	0.0486	0.0113	-0.0024	-0.0228
Gambel Oak / Utah Serviceberry	MsaTp/RckSI	-0.0297	-0.0211	0.0641	0.0343	-0.0032	0.0363	0.0460	0.0101	-0.0178	-0.0286
<u>Water Birch Temporarily Flooded Shrubland Alliance</u>											
Fremont Cottonwood / Water Birch Wooded	Strmb/HPltu	-0.0225	-0.0277	-0.0108	-0.0128	0.0009	-0.0014	0.0005	0.0448	0.0421	0.1064
<u>Strapleaf Willow Temporarily Flooded Shrubland Alliance</u>											
Strapleaf Willow / Beaked Sedge	Strmb/HPltu	-0.0110	-0.0392	-0.0578	-0.0118	-0.0030	-0.0084	-0.0126	0.1070	0.0733	0.1149
<u>(Coyote Willow, Sandbar Willow) Temporarily Flooded Shrubland Alliance</u>											
Coyote Willow / Barren	--	0.0100	0.0450	0.0273	0.0545	-0.0368	-0.0746	-0.0713	-0.0464	-0.0204	0.0710
Coyote Willow / Mesic Graminoids	Strmb	-0.0075	-0.0222	-0.0435	-0.0036	-0.0087	-0.0315	-0.0250	0.0180	0.0475	0.2544

	Cluster ¹	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
IV. DWARF-SHRUBLAND											
<u>Black Sagebrush Dwarf-shrubland Alliance</u>											
Black Sagebrush / Bottlebrush	--	0.0176	0.0195	0.0215	0.0238	0.0094	-0.0173	0.0052	-0.0082	-0.0248	-0.0219
Black Sagebrush / Muttongrass	MsaTp/RckSl	-0.0180	-0.0101	0.0468	0.0361	0.0215	0.0340	0.0580	-0.0037	-0.0318	-0.0450
Black Sagebrush / Needle-and-Thread	UpSnd/RckSl	-0.0031	0.0230	0.0704	0.0474	-0.0041	-0.0101	0.0143	-0.0194	-0.0382	-0.0356
<u>Snakeweed Dwarf-shrubland Alliance</u>											
Snakeweed - (Prickly-pea species) / James' Galleta	ArdLw	0.0557	0.1171	0.0503	-0.0068	-0.0471	-0.0661	-0.0451	-0.0542	-0.0540	-0.0204
V. HERBACEOUS VEGETATION											
<u>Intermediate Wheatgrass Semi-natural Herbaceous Alliance</u>											
Intermediate Wheatgrass Semi-natural	HPltu	-0.0087	-0.0432	-0.0174	-0.0190	-0.0147	0.0089	0.0349	0.1599	0.0408	0.0487
<u>Mountain Muhy Herbaceous Alliance</u>											
(Sandhill Muhy, Mountain Muhy) - Hairy Goldenaster	Slrck/UpSnd	-0.0305	-0.0153	0.0481	0.1097	0.0557	-0.0134	-0.0024	-0.0167	-0.0156	-0.0148
<u>Needle-and-Thread Bunch Herbaceous Alliance</u>											
Needle-and-Thread Great Basin	--	0.0212	-0.0171	0.0041	0.0385	-0.0086	0.0052	0.0350	0.0378	-0.0027	-0.0083
<u>Smooth Brome Semi-natural Herbaceous Alliance</u>											
Smooth Brome - (Western Wheatgrass) Semi-natural	--	0.0931	-0.0306	-0.0070	-0.0175	-0.0124	0.0173	0.0624	0.0717	0.0021	0.0174
<u>Sand Dropseed Herbaceous Alliance</u>											
Sand Dropseed Great Basin	LwFlt/ArdLw	0.0686	0.0819	0.0181	0.0175	-0.0301	-0.0995	-0.0820	-0.0563	-0.0374	0.0469
<u>James' Galleta Herbaceous Alliance</u>											
James' Galleta	LwFlt/ArdLw	0.0877	0.1470	0.0421	-0.0409	-0.0600	-0.1016	-0.0797	-0.0669	-0.0596	-0.0165
<u>Blue Grama Herbaceous Alliance</u>											
Blue Grama - Needle-and-Thread	UpSnd	-0.0229	-0.0197	0.0485	0.0965	0.0109	-0.0001	0.0182	-0.0117	-0.0223	-0.0308
<u>Baltic Rush Seasonally Flooded Herbaceous Alliance</u>											
Baltic Rush	Strmb/HPltu	0.0144	-0.0256	-0.0491	-0.0162	-0.0073	-0.0267	-0.0133	0.0801	0.0315	0.1540
<u>Kentucky Bluegrass Semi-natural Seasonally Flooded Herbaceous Alliance</u>											
Kentucky Bluegrass Semi-natural Seasonally Flooded	--	0.0581	-0.0225	-0.0411	-0.0045	-0.0025	-0.0113	0.0234	0.1086	0.0064	0.0341
<u>(Swollen-beak Sedge, Beaked Sedge) Seasonally Flooded Herbaceous Alliance</u>											
Beaked Sedge	Strmb/HPltu	-0.0211	-0.0347	-0.0341	0.0063	0.0144	-0.0080	-0.0145	0.0198	0.0569	0.1332

	Cluster ¹	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
V. HERBACEOUS VEGETATION (cont.)											
<u>Nebraska Sedge Seasonally Flooded Herbaceous Alliance</u>											
Nebraska Sedge	Strmb/HPltu	-0.0169	-0.0402	-0.0439	0.0096	0.0197	-0.0189	-0.0137	0.0671	0.0526	0.1606
<u>Green Rabbitbrush Shrub Herbaceous Alliance</u>											
Green Rabbitbrush / Kentucky Bluegrass Semi-natural	--	0.0508	0.0253	-0.0073	0.0057	-0.0382	-0.0465	-0.0258	0.0349	-0.0229	0.0275
<u>(Field Horsetail, Variegated Scouringrush) Semipermanently Flooded Herbaceous Alliance</u>											
(Field Horsetail, Variegated Scouringrush)	Strmb	-0.0020	-0.0023	-0.0227	0.0133	-0.0139	-0.0569	-0.0468	0.0297	0.0219	0.2354
VII. SPARSE VEGETATION											
<u>Wooded Bedrock Sparsely Vegetated Alliance</u>											
Ponderosa Pine Slickrock	UpSnd/RckSl	-0.0389	-0.0118	0.0941	0.0791	0.0381	0.0123	0.0204	-0.0354	-0.0271	-0.0409
<u>Littleleaf Mountain-mahogany Sparsely Vegetated Alliance</u>											
Littleleaf Mountain-mahogany Slickrock	Slrck/UpSnd	-0.0428	-0.0225	0.0572	0.0899	0.1010	-0.0043	-0.0008	-0.0417	-0.0082	-0.0362
<u>Mountain-mahogany Sparsely Vegetated Alliance</u>											
Mountain-mahogany Rock Pavement	MsaTp/RckSl	-0.0364	-0.0268	0.0768	0.0301	-0.0053	0.0845	0.0628	-0.0280	-0.0347	-0.0565
<u>Painted Desert Sparsely Vegetated Alliance</u>											
Crispleaf Wild Buckwheat Badlands	ArdLw	0.0203	0.1338	0.0527	-0.0180	-0.0422	-0.0599	-0.0491	-0.0555	-0.0478	-0.0289
Nevada Joint-fir / Lichen	LwFlt/ArdLw	0.1295	0.1541	-0.0038	-0.0627	-0.0608	-0.0967	-0.0827	-0.0556	-0.0430	-0.0127
Xx. HIERARCHY PLACEMENT UNDETERMINED											
<u>Emory Seepwillow Shrubland [Provisional]</u>											
Emory Seepwillow	Strmb	-0.0004	-0.0124	-0.0214	-0.0201	-0.0025	-0.0408	-0.0396	0.0225	0.0431	0.2246
<u>Rubber Rabbitbrush Sand Deposit Sparse Vegetation [Provisional]</u>											
Rubber Rabbitbrush Sand Deposit	RckSl/UpSnd	0.0115	0.0194	0.0318	0.0634	0.0060	-0.0492	-0.0413	-0.0293	-0.0184	0.0072

¹Clusters identified through hierachial clustering of NVCS associations using coalition group affinities shown here (see Fig. 2.15).

ArdLw = Arid Lowlands Woodland/Shrub/Sparse; **CrvcC** = Crevice Canyon Forest/Woodland; **CrvcC/HPltu** = Crevice Canyon/High Plateau Forest/Woodland

HPltu = High Plateau Forest/Shrub/Herb; **LwFlt/ArdLw** = Lowland Flats/Arid Lowlands Shrub/Herb/Sparse

MsaTp = Mesa Top Forest/Woodland/Shrub; **MsaTp/RckSl** = Mesa Top/Rocky Slopes Woodland/Shrub/Sparse

RckSl = Rocky Slopes Woodland/Shrub; **RckSl/UpSnd** = Rocky Slopes/Upland Sands Shrub/Sparse; **Slrck/UpSnd** = Slickrock/Upland Sands Herb/Sparse

Strmb = Streambank Forest/Woodland/ Shrub/Herb; **Strmb/HPltu** = Streambank/High Plateau Woodland/Shrub/Herb

UpSnd = Upland Sands Shrub/Herb; **UpSnd/RckSl** = Upland Sands/Rocky Slopes Woodland/Shrub/Sparse

APPENDIX 2.5

Vegetation map units of Zion National Park, Utah as mapped by Cogan et al. (2004) (rows) with their spatial correspondence to coalition species groups of the current study (columns). Regression tree modeling was used to create predictive maps of plot-averaged species group affinities (in units of the phi coefficient) across the entire park at a 10 m pixel resolution, and these affinity values were then averaged within vegetation map units to give the values shown here. Low values (green shading) indicate low correspondence between map units and coalition groups; high values (red shading) indicate high correspondence (shading is scaled separately for each column). Map units are grouped into eight higher units following USGS (2004).

	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<u>Bare - Soil/Stone Formations</u>										
Basalt Talus	-0.0091	-0.0026	0.0360	0.0121	-0.0071	0.0136	0.0242	0.0738	0.0044	-0.0186
Carmel Formation (Limestone)	-0.0367	-0.0392	0.0009	0.0200	0.0076	0.0976	0.0868	0.1121	0.0334	0.0005
Chinle Formation - Petrified Forest (Shale)	0.0098	0.1266	0.0667	-0.0098	-0.0249	-0.0518	-0.0419	-0.0386	-0.0286	-0.0290
Chinle Formation - Shinarump (Shale)	0.0078	0.1070	0.0704	-0.0014	-0.0167	-0.0289	-0.0330	-0.0361	-0.0270	-0.0334
Gullies and Eroded Lands	-0.0055	0.0579	0.0629	0.0089	-0.0119	-0.0272	-0.0230	-0.0258	-0.0128	-0.0045
Kaibab Formation (Limestone)	-0.0368	-0.0142	0.0692	0.0187	-0.0260	-0.0053	-0.0091	-0.0399	-0.0298	-0.0349
Kayenta Formation (Sandstone)	-0.0029	0.0989	0.0880	0.0061	-0.0192	-0.0394	-0.0336	-0.0328	-0.0205	-0.0290
Moenave Formation (Sandstone)	-0.0103	0.1133	0.0735	0.0006	-0.0235	-0.0321	-0.0347	-0.0340	-0.0214	-0.0310
Moenkopi Formation (Conglomerate)	0.0127	0.0829	0.0689	-0.0010	-0.0252	-0.0306	-0.0211	-0.0364	-0.0248	-0.0322
Navajo Formation (Sandstone)	-0.0363	-0.0216	0.0631	0.0789	0.0749	0.0161	0.0069	-0.0126	0.0034	-0.0273
Slides (Fans and Slumps)	-0.0060	0.0883	0.0733	0.0044	-0.0166	-0.0365	-0.0306	-0.0317	-0.0176	-0.0270
Temple Cap (Sandstone)	-0.0363	-0.0378	-0.0060	0.0220	0.0055	0.0929	0.0842	0.1023	0.0346	0.0023
Unvegetated Volcanic Cinders and Cinder Cones	-0.0095	0.1324	0.0692	-0.0118	-0.0260	-0.0598	-0.0426	-0.0383	-0.0298	-0.0349
Volcanic and Basalt Cliffs	-0.0023	0.1191	0.0691	-0.0049	-0.0228	-0.0248	-0.0369	-0.0377	-0.0283	-0.0346
<u>Grasslands</u>										
Cheatgrass Annual Disturbed Grassland	0.0189	0.0177	0.0220	0.0198	-0.0165	-0.0085	-0.0059	-0.0205	-0.0191	0.0098
Dry Meadow Mixed Herbaceous Vegetation Mosaic	-0.0140	-0.0319	0.0363	0.0260	-0.0119	0.0298	0.0414	0.0323	-0.0005	-0.0195
James' Galleta Herbaceous Vegetation	0.0470	0.0938	0.0451	-0.0108	-0.0253	-0.0593	-0.0424	-0.0399	-0.0298	-0.0104
Mountain Snowberry / Kentucky Bluegrass Semi-natural Shrb.	-0.0237	-0.0381	0.0022	0.0285	-0.0134	0.0359	0.0453	0.0879	0.0287	-0.0067
Perennial Disturbed Grassland Complex	-0.0111	-0.0297	0.0315	0.0332	-0.0059	0.0304	0.0355	0.0392	-0.0012	-0.0174
Sand Dropseed Great Basin Herbaceous Vegetation	0.0431	0.0628	0.0104	-0.0061	-0.0229	-0.0514	-0.0370	-0.0390	-0.0283	0.0474

	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<u>Higher Elevation Forest</u>										
Douglas-fir Forest Alliance	-0.0330	-0.0271	0.0436	0.0463	0.0615	0.0188	0.0106	-0.0013	0.0515	-0.0101
Ponderosa Pine / Gambel Oak Woodland Complex	-0.0295	-0.0287	0.0485	0.0433	0.0282	0.0345	0.0340	0.0062	0.0058	-0.0252
Ponderosa Pine / Greenleaf Manzanita Woodland	-0.0348	-0.0188	0.0611	0.0724	0.0592	0.0228	0.0175	-0.0121	-0.0083	-0.0315
Ponderosa Pine / Mixed Herbaceous Woodland Complex	-0.0250	-0.0332	0.0318	0.0274	-0.0104	0.0313	0.0419	0.0410	0.0054	-0.0185
Ponderosa Pine Forest (Closed Canopy)	-0.0268	-0.0392	0.0405	0.0315	0.0032	0.0626	0.0585	0.0068	-0.0043	-0.0332
Quaking Aspen Forest Complex	-0.0232	-0.0392	-0.0099	0.0187	-0.0127	0.0327	0.0474	0.1196	0.0363	0.0030
Snags	-0.0205	-0.0392	0.0271	0.0234	-0.0048	0.0441	0.0499	0.0519	0.0097	-0.0174
White Fir Forest Alliance	-0.0289	-0.0347	0.0202	0.0327	0.0223	0.0490	0.0484	0.0730	0.0274	-0.0076
<u>Higher Elevation Shrub</u>										
Bigtooth Maple / Gambel Oak Forest	-0.0226	-0.0389	0.0051	0.0257	0.0063	0.0475	0.0543	0.0868	0.0277	-0.0051
Black Sagebrush Dwarf-shrubland Complex	-0.0211	-0.0387	0.0107	0.0204	-0.0178	0.0299	0.0447	0.0977	0.0211	-0.0092
Gambel Oak Shrubland Alliance	-0.0274	-0.0278	0.0446	0.0291	0.0079	0.0471	0.0480	0.0123	-0.0003	-0.0276
Gambel Oak Woodland	-0.0214	-0.0266	0.0416	0.0318	0.0052	0.0353	0.0382	0.0156	0.0011	-0.0248
Greenleaf Manzanita - Gambel Oak - (Utah Serviceberry) Shrb.	-0.0306	-0.0350	0.0457	0.0455	0.0234	0.0496	0.0466	0.0282	0.0029	-0.0255
Greenleaf Manzanita Shrubland Complex	-0.0332	-0.0077	0.0589	0.0630	0.0440	0.0168	0.0160	-0.0113	-0.0096	-0.0306
Mixed Mountain Shrubland Complex	-0.0319	-0.0309	0.0517	0.0344	0.0140	0.0544	0.0534	0.0077	-0.0046	-0.0307
Mountain-mahogany Rock Pavement Sparse Vegetation	-0.0367	-0.0392	0.0185	0.0200	0.0072	0.0993	0.0883	0.0685	0.0136	-0.0168
Utah Serviceberry Shrubland	-0.0322	-0.0362	0.0487	0.0233	-0.0067	0.0479	0.0551	0.0154	-0.0014	-0.0289
<u>Lower Elevation Forest (Pinyon/Juniper)</u>										
Juniper / Big Sagebrush Woodland Complex	0.0065	0.1108	0.0619	-0.0034	-0.0210	-0.0454	-0.0365	-0.0387	-0.0283	-0.0240
Pinyon - Juniper / Gamble Oak Woodland Complex	-0.0243	-0.0112	0.0567	0.0292	0.0028	0.0308	0.0335	-0.0093	-0.0118	-0.0316
Pinyon - Juniper Woodland Complex	-0.0136	0.0483	0.0683	0.0196	-0.0047	-0.0128	-0.0071	-0.0292	-0.0213	-0.0296
<u>Lower Elevation Shrub</u>										
Big Sagebrush Shrubland Complex	-0.0008	0.0245	0.0425	0.0284	-0.0166	-0.0071	0.0005	-0.0202	-0.0193	-0.0235
Blackbrush Shrubland Complex	0.0418	0.1012	0.0300	-0.0088	-0.0223	-0.0564	-0.0425	-0.0397	-0.0297	0.0001
Croplands and Pastures	-0.0001	0.0024	0.0355	0.0501	-0.0254	0.0034	0.0039	-0.0267	-0.0212	-0.0336
Mixed Urban or Built-up Land	0.0625	0.0621	-0.0088	-0.0084	-0.0256	-0.0541	-0.0381	-0.0364	-0.0288	0.0501
Nevada Joint-fir Basalt Shrubland	0.0110	0.1035	0.0607	-0.0082	-0.0143	-0.0409	-0.0420	-0.0386	-0.0283	-0.0197
Other Agricultural Lands	-0.0249	-0.0392	0.0346	0.0187	-0.0114	0.0621	0.0656	0.1219	-0.0056	-0.0140

	Lowland Flats	Arid Lowlands	Rocky Slopes	Upland Sands	Slickrock	Xeric Mesa Top	Mesic Mesa Top	High Plateau	Crevice Canyon	Stream-bank
<u>Lower Elevation Shrub (cont.)</u>										
Painted Desert Sparsely Vegetated Alliance	0.0553	0.1308	0.0679	-0.0117	-0.0259	-0.0575	-0.0425	-0.0395	-0.0295	-0.0320
Rabbitbrush Shrubland Complex	0.0144	0.0303	0.0243	0.0061	-0.0116	-0.0170	-0.0082	-0.0118	-0.0116	0.0173
Sand Sagebrush Colorado Plateau Shrubland	0.0077	0.0679	0.0394	0.0048	-0.0082	-0.0429	-0.0351	-0.0382	-0.0276	0.0033
Snakeweed -(Prickly-pear) / James' Galleta Dwarf-shrubland	0.0159	0.0930	0.0503	-0.0006	-0.0137	-0.0441	-0.0343	-0.0338	-0.0245	-0.0140
Strip Mines, Quarries, and Gravel Pits	-0.0120	0.0311	0.0400	0.0187	-0.0260	-0.0275	-0.0091	-0.0399	-0.0298	-0.0126
Talus Mixed Shrubland	-0.0186	0.0387	0.0698	0.0306	0.0155	-0.0135	-0.0141	-0.0264	-0.0094	-0.0260
Transportation, Communications, and Utilities	-0.0011	0.0260	0.0403	0.0180	-0.0115	-0.0095	-0.0010	-0.0163	-0.0182	-0.0055
<u>Riparian/Flooded Woodland</u>										
Boxelder Woodland Alliance	-0.0180	0.0101	0.0477	0.0300	0.0129	-0.0136	-0.0113	-0.0212	0.0097	-0.0118
Cattail, Bulrush, Emergent Wetland Complex	-0.0133	-0.0064	0.0710	0.0187	-0.0260	0.0027	0.0199	0.0464	-0.0169	-0.0208
Emory Seepwillow Shrubland	0.0242	0.0620	-0.0026	-0.0095	-0.0220	-0.0476	-0.0413	-0.0351	-0.0198	0.1421
Fremont Cottonwood - Velvet Ash Woodland	0.0011	0.0344	0.0519	0.0103	0.0062	-0.0198	-0.0257	-0.0198	0.0034	0.0068
Fremont Cottonwood Woodland Complex	0.0297	0.0585	0.0150	-0.0055	-0.0190	-0.0452	-0.0375	-0.0331	-0.0179	0.0669
Intermittent Streams	-0.0151	0.0257	0.0491	0.0259	0.0056	-0.0158	-0.0088	-0.0150	0.0022	0.0028
Perennial Streams	-0.0011	0.0407	0.0229	0.0052	-0.0134	-0.0343	-0.0297	-0.0192	0.0105	0.0586
Reservoirs	-0.0011	-0.0392	-0.0071	0.0206	0.0056	0.0300	0.0373	0.0880	0.0664	-0.0030
Russian Olive Semi-natural Woodland	0.0499	0.0651	-0.0151	-0.0118	-0.0260	-0.0510	-0.0426	-0.0399	-0.0298	0.1060
Sand Bars and Beaches	0.0066	0.0467	0.0220	0.0084	-0.0248	-0.0347	-0.0225	-0.0364	-0.0247	0.0440
Sandbar Willow Shrubland Alliance	-0.0063	0.0429	0.0343	0.0159	-0.0119	-0.0292	-0.0187	-0.0305	-0.0151	0.0183
Sedge-Rush Wet Meadow Herbaceous Vegetation Mosaic	-0.0097	-0.0366	0.0003	0.0196	-0.0168	0.0376	0.0491	0.0880	0.0263	-0.0048
Stock Ponds	-0.0044	-0.0392	0.0177	0.0302	-0.0046	0.0304	0.0407	0.0636	0.0141	-0.0088
Strapleaf Willow / Beaked Sedge Shrubland	-0.0225	-0.0368	0.0068	0.0290	0.0061	0.0268	0.0324	0.0684	0.0428	-0.0080
Tamarisk spp. Temporarily Flooded Shrubland	0.0144	0.0823	0.0050	-0.0118	-0.0260	-0.0481	-0.0426	-0.0399	-0.0298	0.1326
Tinajas (natural water holes / tanks)	-0.0368	0.0228	0.0692	0.0562	0.0491	-0.0178	-0.0066	-0.0246	-0.0074	-0.0243
<u>Slickrock</u>										
Littleleaf Mountain-mahogany Slickrock Sparse Vegetation	-0.0364	-0.0007	0.0668	0.0787	0.0640	0.0026	0.0018	-0.0225	-0.0138	-0.0317
Ponderosa Pine Slickrock Sparse Vegetation	-0.0363	-0.0124	0.0669	0.0836	0.0716	0.0103	0.0048	-0.0180	-0.0109	-0.0317

APPENDIX 3.1

Vascular plant genera and species included in niche overlap analyses for plant communities of Zion National Park, Utah, showing correspondence between different taxonomic concepts. Genus names follow NCBI (2008), species names follow Welsh et al. (2003) and database names follow Kartesz (1999). #In=number of plot records (out of 332) in the 1999-2000 USGS-NPS Zion N.P. vegetation survey database in which the taxon was recorded within the plot (i.e., cover>0); #Out=number of additional plot records in which the taxon was recorded near but not within the plot (i.e., cover=0). All records (#In + #Out) where used in presence-based niche overlap analyses, but only within-plot records were used in cover-based analyses.

Genus	Species	Database Name	#In	#Out
<i>Abies</i>	<i>Abies concolor</i>	<i>Abies concolor</i> (Gord. & Glend.) Lindl. ex Hildebr.	33	5
<i>Abronia</i>	<i>Abronia fragrans</i>	<i>Abronia fragrans</i> Nutt. ex Hook.	4	
<i>Acer</i>	<i>Acer grandidentatum</i>	<i>Acer grandidentatum</i> Nutt.	32	3
<i>Acer</i>	<i>Acer negundo</i>	<i>Acer negundo</i> L.	23	2
<i>Achillea</i>	<i>Achillea millefolium</i>	<i>Achillea millefolium</i> L.	24	
<i>Achnatherum</i>	<i>Stipa hymenoides</i>	<i>Achnatherum contractum</i> (B.L. Johnson) Barkworth	1	
<i>Achnatherum</i>	<i>Stipa hymenoides</i>	<i>Achnatherum hymenoides</i> (Roemer & J.A. Schultes) Barkworth	54	4
<i>Achnatherum</i>	<i>Stipa lettermanii</i>	<i>Achnatherum lettermanii</i> (Vasey) Barkworth	12	
<i>Achnatherum</i>	<i>Stipa nelsonii</i>	<i>Achnatherum nelsonii</i> ssp. <i>nelsonii</i> (Scribn.) Barkworth	2	
<i>Achnatherum</i>	<i>Stipa speciosa</i>	<i>Achnatherum speciosum</i> (Trin. & Rupr.) Barkworth	5	
<i>Agastache</i>	<i>Agastache urticifolia</i>	<i>Agastache urticifolia</i> (Benth.) Kuntze	3	
<i>Agoseris</i>		<i>Agoseris</i> Raf.	2	1
<i>Agropyron</i>	<i>Agropyron cristatum</i>	<i>Agropyron cristatum</i> (L.) Gaertn.	3	
<i>Agrostis</i>	<i>Agrostis exarata</i>	<i>Agrostis exarata</i> Trin.	3	
<i>Agrostis</i>	<i>Agrostis stolonifera</i>	<i>Agrostis stolonifera</i> L.	4	
<i>Agrostis</i>		<i>Agrostis</i> L.	2	
<i>Allionia</i>	<i>Allionia incarnata</i>	<i>Allionia incarnata</i> L.	1	
<i>Allium</i>		<i>Allium</i> L.	6	1
<i>Ambrosia</i>	<i>Ambrosia acanthicarpa</i>	<i>Ambrosia acanthicarpa</i> Hook.	1	
<i>Amelanchier</i>	<i>Amelanchier alnifolia</i>	<i>Amelanchier alnifolia</i> (Nutt.) Nutt. ex M. Roemer	17	
<i>Amelanchier</i>	<i>Amelanchier utahensis</i>	<i>Amelanchier utahensis</i> Koehne	136	2
<i>Andropogon</i>	<i>Andropogon gerardii</i>	<i>Andropogon gerardii</i> Vitman	1	
<i>Antennaria</i>	<i>Antennaria dimorpha</i>	<i>Antennaria dimorpha</i> (Nutt.) Torr. & Gray	4	
<i>Antennaria</i>		<i>Antennaria</i> Gaertn.	7	
<i>Apocynum</i>	<i>Apocynum androsaemifolium</i>	<i>Apocynum androsaemifolium</i> L.	1	
<i>Aquilegia</i>		<i>Aquilegia</i> L.	1	1
<i>Arabis</i>	<i>Arabis holboellii</i>	<i>Arabis holboellii</i> Hornem.	8	
<i>Arabis</i>	<i>Arabis perennans</i>	<i>Arabis perennans</i> S. Wats.	1	
<i>Arabis</i>		<i>Arabis</i> L.	19	
<i>Arctostaphylos</i>	<i>Arctostaphylos patula</i>	<i>Arctostaphylos patula</i> Greene	89	2
<i>Arctostaphylos</i>	<i>Arctostaphylos pungens</i>	<i>Arctostaphylos pungens</i> Kunth	11	
<i>Arenaria</i>	<i>Arenaria fendleri</i>	<i>Arenaria fendleri</i> Gray	25	
<i>Arenaria</i>	<i>Arenaria macradenia</i>	<i>Arenaria macradenia</i> S. Wats.	14	
<i>Arenaria</i>		<i>Arenaria</i> L.	3	

Genus	Species	Database Name	#In	#Out
Aristida	<i>Aristida purpurascens</i>	<i>Aristida purpurascens</i> Poir.	2	
Aristida	<i>Aristida purpurea</i>	<i>Aristida purpurea</i> Nutt.	18	
Arnica		<i>Arnica</i> L.	1	
Artemisia	<i>Artemisia campestris</i>	<i>Artemisia campestris</i> L.	13	
Artemisia	<i>Artemisia dracunculus</i>	<i>Artemisia dracunculus</i> L.	10	
Artemisia	<i>Artemisia dracunculus</i>	<i>Artemisia dracunculus</i> ssp. <i>dracunculus</i> L.	1	
Artemisia	<i>Artemisia filifolia</i>	<i>Artemisia filifolia</i> Torr.	10	
Artemisia	<i>Artemisia ludoviciana</i>	<i>Artemisia ludoviciana</i> Nutt.	47	1
Artemisia	<i>Artemisia nova</i>	<i>Artemisia nova</i> A. Nels.	22	1
Artemisia	<i>Artemisia tridentata</i>	<i>Artemisia tridentata</i> Nutt.	63	
Artemisia	<i>Artemisia tridentata</i>	<i>Artemisia tridentata</i> ssp. <i>tridentata</i> Nutt.	2	
Artemisia	<i>Artemisia tridentata</i>	<i>Artemisia tridentata</i> ssp. <i>vaseyana</i> (Rydb.) Beetle	9	
Asclepias	<i>Asclepias subverticillata</i>	<i>Asclepias subverticillata</i> (Gray) Vail	2	
Asclepias		unknown milkweed	1	
Asplenium		<i>Asplenium</i> L.	1	
Asteraceae spp.		Asteraceae	1	
Asteraceae spp.		unknown aster	1	
Asteraceae spp.		unknown aster 2	1	
Asteraceae spp.		unknown composite	4	
Asteraceae spp.		unknown forb composite	1	
Asterinae spp.		<i>Aster</i> L.	8	
Astragalus	<i>Astragalus flavus</i>	<i>Astragalus flavus</i> Nutt.	2	
Astragalus	<i>Astragalus subcinereus</i>	<i>Astragalus subcinereus</i> Gray	2	
Astragalus		<i>Astragalus</i> L.	30	1
Atriplex	<i>Atriplex canescens</i>	<i>Atriplex canescens</i> (Pursh) Nutt.	12	
Atriplex		<i>Atriplex confertifolia</i> (Torr. & Frem.) S. Wats.	2	
Atriplex		<i>Atriplex</i> L.	1	
Baccharis	<i>Baccharis salicina</i>	<i>Baccharis emoryi</i> Gray	9	
Baccharis	<i>Baccharis salicina</i>	<i>Baccharis salicifolia</i> (Ruiz & Pavon) Pers.	4	
Baccharis		<i>Baccharis</i> L.	1	
Baileya	<i>Baileya multiradiata</i>	<i>Baileya multiradiata</i> Harvey & Gray ex Gray	1	
Balsamorhiza	<i>Balsamorhiza sagittata</i>	<i>Balsamorhiza sagittata</i> (Pursh) Nutt.	9	1
Betula	<i>Betula occidentalis</i>	<i>Betula occidentalis</i> Hook.	6	1
Boraginaceae spp.		unknown borage	1	
Boraginaceae spp.		unknown forb - borage	1	
Bouteloua	<i>Bouteloua barbata</i>	<i>Bouteloua barbata</i> Lag.	1	
Bouteloua	<i>Bouteloua gracilis</i>	<i>Bouteloua gracilis</i> (Willd. ex Kunth) Lag. ex Griffiths	31	1
Brassicaceae spp.		<i>Brassica</i> L.	1	
Brassicaceae spp.		brassicaceae unknown	1	
Brassicaceae spp.		mustard	4	
Brassicaceae spp.		Mustard - small	1	
Brassicaceae spp.		mustard species	5	
Brassicaceae spp.		unknown mustard	2	
Brickellia	<i>Brickellia atractyloides</i>	<i>Brickellia atractyloides</i> Gray	3	
Brickellia	<i>Brickellia californica</i>	<i>Brickellia californica</i> (Torr. & Gray) Gray	3	
Brickellia	<i>Brickellia grandiflora</i>	<i>Brickellia grandiflora</i> (Hook.) Nutt.	5	
Brickellia	<i>Brickellia longifolia</i>	<i>Brickellia longifolia</i> S. Wats.	6	

Genus	Species	Database Name	#In	#Out
<i>Brickellia</i>	<i>Brickellia microphylla</i>	<i>Brickellia microphylla</i> (Nutt.) Gray	1	1
<i>Brickellia</i>		<i>Brickellia</i> Ell.	7	1
<i>Bromus</i>	<i>Bromus anomalus</i>	<i>Bromus anomalus</i> Rupr. ex Fourn.	7	
<i>Bromus</i>	<i>Bromus diandrus</i>	<i>Bromus diandrus</i> Roth	15	
<i>Bromus</i>	<i>Bromus inermis</i>	<i>Bromus inermis</i> Leyss.	12	1
<i>Bromus</i>	<i>Bromus rubens</i>	<i>Bromus rubens</i> L.	6	
<i>Bromus</i>	<i>Bromus tectorum</i>	<i>Bromus tectorum</i> L.	93	1
<i>Bromus</i>		<i>Bromus</i> L.	3	
<i>Calamagrostis</i>		<i>Calamagrostis</i> Adans.	1	
<i>Calochortus</i>	<i>Calochortus nuttallii</i>	<i>Calochortus nuttallii</i> Torr. & Gray	6	1
<i>Calochortus</i>		<i>Calochortus</i> Pursh	3	
<i>Carex</i>	<i>Carex geyeri</i>	<i>Carex geyeri</i> Boott	2	
<i>Carex</i>	<i>Carex microptera</i>	<i>Carex microptera</i> Mackenzie	5	
<i>Carex</i>	<i>Carex nebrascensis</i>	<i>Carex nebrascensis</i> Dewey	2	
<i>Carex</i>	<i>Carex occidentalis</i>	<i>Carex occidentalis</i> Bailey	8	
<i>Carex</i>	<i>Carex rossii</i>	<i>Carex rossii</i> Boott	10	
<i>Carex</i>	<i>Carex utriculata</i>	<i>Carex utriculata</i> Boott	2	
<i>Carex</i>		<i>Carex</i> L.	18	1
<i>Carex</i>		<i>Carex</i> L. (<i>occidentalis</i> ?)	1	
<i>Carex</i>		<i>Carex</i> L. (<i>rossii</i> ?)	1	
<i>Caryophyllaceae</i> spp.		unknown caryophyllaceae	2	
<i>Castilleja</i>	<i>Castilleja chromosa</i>	<i>Castilleja applegatei</i> ssp. <i>martinii</i> (Abrams) Chuang & Heckard	3	
<i>Castilleja</i>	<i>Castilleja flava</i>	<i>Castilleja flava</i> var. <i>flava</i> S. Wats.	1	
<i>Castilleja</i>	<i>Castilleja linariifolia</i>	<i>Castilleja linariifolia</i> Benth.	7	
<i>Castilleja</i>		<i>Castilleja</i> Mutis ex L. f.	6	
<i>Ceanothus</i>	<i>Ceanothus fendleri</i>	<i>Ceanothus fendleri</i> Gray	4	1
<i>Ceanothus</i>	<i>Ceanothus martinii</i>	<i>Ceanothus martinii</i> M.E. Jones	2	
<i>Ceanothus</i>		<i>Ceanothus</i> L.	1	
<i>Celtis</i>	<i>Celtis reticulata</i>	<i>Celtis laevigata</i> var. <i>reticulata</i> (Torr.) L. Benson	4	1
<i>Cenchrus</i>	<i>Cenchrus longispinus</i>	<i>Cenchrus longispinus</i> (Hack.) Fern.	1	
<i>Cercocarpus</i>	<i>Cercocarpus intricatus</i>	<i>Cercocarpus intricatus</i> S. Wats.	18	3
<i>Cercocarpus</i>	<i>Cercocarpus ledifolius</i>	<i>Cercocarpus ledifolius</i> Nutt.	2	
<i>Cercocarpus</i>	<i>Cercocarpus montanus</i>	<i>Cercocarpus montanus</i> Raf.	44	1
<i>Chaenactis</i>	<i>Chaenactis douglasii</i>	<i>Chaenactis douglasii</i> (Hook.) Hook. & Arn.	9	
<i>Chaenactis</i>	<i>Chaenactis stevioides</i>	<i>Chaenactis stevioides</i> Hook. & Arn.	1	
<i>Chaenactis</i>		<i>Chaenactis</i> DC.	2	
<i>Chaenactis</i>		<i>Chaenactis</i> DC. (<i>douglasii</i> or <i>stevioides</i> ??)	1	
<i>Chaetopappa</i>	<i>Chaetopappa ericoides</i>	<i>Chaetopappa ericoides</i> (Torr.) Nesom	1	
<i>Chamaesyce</i>	<i>Chamaesyce albomarginata</i>	<i>Chamaesyce albomarginata</i> (Torr. & Gray) Small	1	
<i>Chamaesyce</i>	<i>Chamaesyce fendleri</i>	<i>Chamaesyce fendleri</i> (Torr. & Gray) Small	7	
<i>Chamaesyce</i>	<i>Chamaesyce glyptosperma</i>	<i>Chamaesyce glyptosperma</i> (Engelm.) Small	4	
<i>Chamaesyce</i>	<i>Chamaesyce parryi</i>	<i>Chamaesyce parryi</i> (Engelm.) Rydb.	1	
<i>Chenopodium</i>	<i>Chenopodium album</i>	<i>Chenopodium album</i> L.	2	
<i>Chenopodium</i>	<i>Chenopodium fremontii</i>	<i>Chenopodium fremontii</i> S. Wats.	4	
<i>Chenopodium</i>		<i>Chenopodium</i> L.	1	
<i>Chimaphila</i>	<i>Chimaphila umbellata</i>	<i>Chimaphila umbellata</i> (L.) W. Bart.	1	

Genus	Species	Database Name	#In	#Out
<i>Chrysopsis</i>	<i>Chrysopsis villosa</i>	<i>Heterotheca villosa</i> (Pursh) Shinners	59	
<i>Chrysopsis</i>	<i>Chrysopsis villosa</i>	<i>Heterotheca viscosa</i> (Gray) Harms	1	
<i>Chrysothamnus</i>	<i>Chrysothamnus depressus</i>	<i>Chrysothamnus depressus</i> Nutt.	3	
<i>Chrysothamnus</i>	<i>Chrysothamnus viscidiflorus</i>	<i>Chrysothamnus viscidiflorus</i> (Hook.) Nutt.	12	
<i>Chrysothamnus</i>	<i>Chrysothamnus viscidiflorus</i>	<i>Chrysothamnus viscidiflorus</i> ssp. <i>axillaris</i> (Keck) L.C. Anders.	1	
<i>Chrysothamnus</i>		<i>Chrysothamnus</i> Nutt.	1	
<i>Cirsium</i>	<i>Cirsium arizonicum</i>	<i>Cirsium arizonicum</i> (Gray) Petrak	1	
<i>Cirsium</i>	<i>Cirsium calcareum</i>	<i>Cirsium calcareum</i> (M.E. Jones) Woot. & Standl.	1	
<i>Cirsium</i>	<i>Cirsium vulgare</i>	<i>Cirsium vulgare</i> (Savi) Ten.	1	
<i>Cirsium</i>	<i>Cirsium wheeleri</i>	<i>Cirsium wheeleri</i> (Gray) Petrak	6	
<i>Cirsium</i>		<i>Cirsium</i> P. Mill.	16	
<i>Claytonia</i>	<i>Montia perfoliata</i>	<i>Claytonia perfoliata</i> ssp. <i>perfoliata</i> var. <i>nubigena</i> (Greene) Poelln.	4	
<i>Clematis</i>	<i>Clematis columbiana</i>	<i>Clematis columbiana</i> (Nutt.) Torr. & Gray	2	
<i>Clematis</i>	<i>Clematis ligusticifolia</i>	<i>Clematis ligusticifolia</i> Nutt.	9	
<i>Clematis</i>		<i>Clematis</i> L.	1	
<i>Coleogyne</i>	<i>Coleogyne ramosissima</i>	<i>Coleogyne ramosissima</i> Torr.	12	
<i>Collinsia</i>	<i>Collinsia parviflora</i>	<i>Collinsia parviflora</i> Lindl.	1	
<i>Comandra</i>	<i>Comandra umbellata</i>	<i>Comandra umbellata</i> (L.) Nutt.	37	1
<i>Comandra</i>	<i>Comandra umbellata</i>	<i>Comandra umbellata</i> ssp. <i>pallida</i> (A. DC.) Piehl	2	
<i>Convolvulus</i>	<i>Convolvulus arvensis</i>	<i>Convolvulus arvensis</i> L.	2	
<i>Cordylanthus</i>	<i>Cordylanthus kingii</i>	<i>Cordylanthus kingii</i> S. Wats.	2	
<i>Cordylanthus</i>	<i>Cordylanthus parviflorus</i>	<i>Cordylanthus parviflorus</i> (Ferris) Wiggins	4	
<i>Crepis</i>	<i>Crepis intermedia</i>	<i>Crepis intermedia</i> Gray	1	
<i>Cryptantha</i>	<i>Cryptantha humilis</i>	<i>Cryptantha humilis</i> (Gray) Payson	2	
<i>Cryptantha</i>		<i>Cryptantha</i> Lehm. ex G. Don	14	
<i>Cupressus</i>	<i>Cupressus arizonica</i>	<i>Cupressus arizonica</i> Greene	1	
<i>Dactylis</i>	<i>Dactylis glomerata</i>	<i>Dactylis glomerata</i> L.	3	
<i>Dalea</i>	<i>Dalea searlsiae</i>	<i>Dalea searlsiae</i> (Gray) Barneby	3	
<i>Datura</i>	<i>Datura wrightii</i>	<i>Datura wrightii</i> Regel	5	
<i>Datura</i>		<i>Datura</i> L.	1	
<i>Descurainia</i>	<i>Descurainia pinnata</i>	<i>Descurainia pinnata</i> (Walt.) Britt.	1	
<i>Descurainia</i>		<i>Descurainia</i> Webb & Berth.	1	
<i>Distichlis</i>	<i>Distichlis spicata</i>	<i>Distichlis spicata</i> (L.) Greene	1	
<i>Draba</i>		<i>Draba</i> L.	4	
<i>Dracocephalum</i>	<i>Dracocephalum parviflorum</i>	<i>Dracocephalum parviflorum</i> Nutt.	1	
<i>Echinocereus</i>	<i>Echinocereus engelmannii</i>	<i>Echinocereus engelmannii</i> (Parry ex Engelm.) Lem.	8	1
<i>Echinocereus</i>	<i>Echinocereus triglochidiatus</i>	<i>Echinocereus triglochidiatus</i> Engelm.	10	1
<i>Echinocereus</i>		<i>Echinocereus</i> Engelm.	3	
<i>Elaeagnus</i>	<i>Elaeagnus angustifolia</i>	<i>Elaeagnus angustifolia</i> L.	2	2
<i>Eleocharis</i>		<i>Eleocharis</i> R. Br.	1	
<i>Elymus</i>	<i>Elymus canadensis</i>	<i>Elymus canadensis</i> L.	2	
<i>Elymus</i>	<i>Elymus elymoides</i>	<i>Elymus elymoides</i> (Raf.) Swezey	57	
<i>Elymus</i>	<i>Elymus elymoides</i>	<i>Elymus elymoides</i> ssp. <i>elymoides</i> (Raf.) Swezey	1	
<i>Elymus</i>	<i>Elymus glaucus</i>	<i>Elymus glaucus</i> Buckl.	1	
<i>Elymus</i>	<i>Elymus hispidus</i>	<i>Elytrigia intermedia</i> (Host) Nevski	2	
<i>Elymus</i>	<i>Elymus hispidus</i>	<i>Elytrigia intermedia</i> ssp. <i>intermedia</i> (Host) Nevski	1	
<i>Elymus</i>	<i>Elymus lanceolatus</i>	<i>Elymus lanceolatus</i> (Scribn. & J.G. Sm.) Gould	11	

Genus	Species	Database Name	#In	#Out
<i>Elymus</i>	<i>Elymus repens</i>	<i>Elytrigia repens</i> (L.) Desv. ex B.D. Jackson	6	
<i>Elymus</i>	<i>Elymus smithii</i>	<i>Pascopyrum smithii</i> (Rydb.) A. Love	9	
<i>Elymus</i>	<i>Elymus spicatus</i>	<i>Pseudoroegneria spicata</i> ssp. <i>spicata</i> (Pursh) A. Love	1	
<i>Elymus</i>		<i>Elymus</i> L.	10	
<i>Ephedra</i>	<i>Ephedra nevadensis</i>	<i>Ephedra nevadensis</i>	1	
<i>Ephedra</i>	<i>Ephedra nevadensis</i>	<i>Ephedra nevadensis</i> S. Wats.	16	2
<i>Ephedra</i>	<i>Ephedra viridis</i>	<i>Ephedra viridis</i> Coville	20	5
<i>Epilobium</i>	<i>Epilobium brachycarpum</i>	<i>Epilobium brachycarpum</i> K. Presl	7	
<i>Epilobium</i>	<i>Zauschneria latifolia</i>	<i>Epilobium canum</i> ssp. <i>garrettii</i> (A. Nels.) Raven	1	
<i>Epilobium</i>	<i>Zauschneria latifolia</i>	<i>Epilobium canum</i> ssp. <i>latifolium</i> (Hook.) Raven	3	
<i>Equisetum</i>	<i>Equisetum arvense</i>	<i>Equisetum arvense</i> L.	5	
<i>Equisetum</i>	<i>Equisetum hyemale</i>	<i>Equisetum hyemale</i> L.	3	
<i>Equisetum</i>	<i>Equisetum laevigatum</i>	<i>Equisetum laevigatum</i> A. Braun	6	
<i>Equisetum</i>	<i>Equisetum variegatum</i>	<i>Equisetum variegatum</i> Schleich. ex F. Weber & D.M.H. Mohr	4	
<i>Equisetum</i>		<i>Equisetum</i> L.	2	1
<i>Ericameria</i>	<i>Chrysanthus nauseosus</i>	<i>Ericameria nauseosa</i> ssp. <i>nauseosa</i> var. <i>nauseosa</i>	37	2
<i>Ericameria</i>	<i>Chrysanthus parryi</i>	<i>Ericameria parryi</i> var. <i>parryi</i> (Gray) Nesom	1	
<i>Ericameria</i>	<i>Haplopappus linearifolius</i>	<i>Ericameria linearifolia</i> (DC.) Urbatsch & Wussow	4	
<i>Erigeron</i>	<i>Erigeron argentatus</i>	<i>Erigeron argentatus</i> Gray	1	
<i>Erigeron</i>	<i>Erigeron divergens</i>	<i>Erigeron divergens</i> Torr. & Gray	1	
<i>Erigeron</i>	<i>Erigeron pumilus</i>	<i>Erigeron concinnus</i> var. <i>concinnus</i> (Hook. & Arn.) Torr. & Gray	1	
<i>Erigeron</i>	<i>Erigeron pumilus</i>	<i>Erigeron pumilus</i> Nutt.	1	
<i>Erigeron</i>	<i>Erigeron speciosus</i>	<i>Erigeron speciosus</i> (Lindl.) DC.	2	
<i>Erigeron</i>	<i>Erigeron speciosus</i>	<i>Erigeron speciosus</i> var. <i>macranthus</i> (Nutt.) Cronq.	1	
<i>Erigeron</i>	<i>Erigeron utahensis</i>	<i>Erigeron utahensis</i> Gray	6	
<i>Erigeron</i>		<i>Erigeron</i> L.	34	1
<i>Eriogonum</i>	<i>Eriogonum corymbosum</i>	<i>Eriogonum corymbosum</i> Benth.	1	
<i>Eriogonum</i>	<i>Eriogonum fasciculatum</i>	<i>Eriogonum fasciculatum</i> Benth.	1	
<i>Eriogonum</i>	<i>Eriogonum flavum</i>	<i>Eriogonum flavum</i> Nutt.	1	
<i>Eriogonum</i>	<i>Eriogonum inflatum</i>	<i>Eriogonum inflatum</i> Torr. & Frem.	3	
<i>Eriogonum</i>	<i>Eriogonum microthecum</i>	<i>Eriogonum microthecum</i> Nutt.	4	1
<i>Eriogonum</i>	<i>Eriogonum ovalifolium</i>	<i>Eriogonum ovalifolium</i> Nutt.	1	
<i>Eriogonum</i>	<i>Eriogonum panguicense</i>	<i>Eriogonum panguicense</i> (M.E. Jones) Reveal	2	
<i>Eriogonum</i>	<i>Eriogonum racemosum</i>	<i>Eriogonum racemosum</i> Nutt.	20	
<i>Eriogonum</i>	<i>Eriogonum umbellatum</i>	<i>Eriogonum umbellatum</i> Torr.	24	
<i>Eriogonum</i>		<i>Eriogonum</i> Michx.	21	
<i>Eriogonum</i>		<i>Eriogonum2</i> Michx.	2	
<i>Erioneuron</i>	<i>Erioneuron pulchellum</i>	<i>Erioneuron pulchellum</i> (Kunth) Tateoka	1	
<i>Erodium</i>	<i>Erodium cicutarium</i>	<i>Erodium cicutarium</i> (L.) L'Her. ex Ait.	6	
<i>Erysimum</i>	<i>Erysimum asperum</i>	<i>Erysimum capitatum</i> var. <i>argillosum</i> (Greene) R.J. Davis	7	
<i>Erysimum</i>	<i>Erysimum asperum</i>	<i>Erysimum capitatum</i> var. <i>capitatum</i> (Dougl. ex Hook.) Greene	1	
<i>Euphorbia</i>	<i>Euphorbia brachycera</i>	<i>Euphorbia brachycera</i> Engelm.	1	
<i>Euphorbia</i>		<i>Euphorbia</i> L.	14	
<i>Eurybia</i>	<i>Aster glaucodes</i>	<i>Aster glaucodes</i> Blake	1	
<i>Fabaceae</i> spp.		<i>Fabaceae</i> sp.	1	
<i>Fabaceae</i> spp.		legume species	1	
<i>Fendlerella</i>	<i>Fendlerella utahensis</i>	<i>Fendlerella utahensis</i> (S. Wats.) Heller	2	

Genus	Species	Database Name	#In	#Out
<i>Festuca</i>	<i>Festuca occidentalis</i>	<i>Festuca occidentalis</i> Hook.	1	
<i>Festuca</i>	<i>Festuca octoflora</i>	<i>Vulpia octoflora</i> var. <i>octoflora</i> (Walt.) Rydb.	4	
<i>Festuca</i>		<i>Festuca</i> L.	2	
<i>Fraxinus</i>	<i>Fraxinus anomala</i>	<i>Fraxinus anomala</i> Torr. ex S. Wats.	28	4
<i>Fraxinus</i>	<i>Fraxinus velutina</i>	<i>Fraxinus velutina</i> Torr.	6	
<i>Gaillardia</i>		<i>Gaillardia</i> Foug.	1	
<i>Galium</i>	<i>Galium aparine</i>	<i>Galium aparine</i> L.	1	
<i>Galium</i>		<i>Galium</i> L.	6	
<i>Gayophytum</i>		<i>Gayophytum</i> A. Juss.	1	
<i>Geraea</i>	<i>Geraea canescens</i>	<i>Geraea canescens</i> Torr. & Gray	1	
<i>Geranium</i>	<i>Geranium caespitosum</i>	<i>Geranium caespitosum</i> James	1	
<i>Geranium</i>		<i>Geranium</i> L.	1	
<i>Gilia</i>		<i>Gilia</i> Ruiz & Pavon	1	
<i>Grayia</i>	<i>Grayia spinosa</i>	<i>Grayia spinosa</i> (Hook.) Moq.	2	
<i>Gutierrezia</i>	<i>Gutierrezia microcephala</i>	<i>Gutierrezia microcephala</i> (DC.) Gray	19	1
<i>Gutierrezia</i>	<i>Gutierrezia sarothrae</i>	<i>Gutierrezia sarothrae</i> (Pursh) Britt. & Rusby	77	2
<i>Helianthella</i>	<i>Helianthella uniflora</i>	<i>Helianthella uniflora</i> (Nutt.) Torr. & Gray	5	
<i>Helianthus</i>	<i>Helianthus petiolaris</i>	<i>Helianthus petiolaris</i> Nutt.	1	
<i>Hesperodoria</i>	<i>Haplopappus scopolorum</i>	<i>Hesperodoria scopularum</i> (M.E. Jones) Greene	3	
<i>Hesperostipa</i>	<i>Stipa comata</i>	<i>Hesperostipa comata</i> ssp. <i>comata</i> (Trin. & Rupr.) Barkworth	30	2
<i>Heuchera</i>	<i>Heuchera rubescens</i>	<i>Heuchera rubescens</i> Torr.	1	
<i>Heuchera</i>	<i>Heuchera rubescens</i>	<i>Heuchera rubescens</i> var. <i>versicolor</i> (Greene) M.G. Stewart	2	
<i>Hilaria</i>		<i>Pleuraphis jamesii</i> Torr.	26	
<i>Holodiscus</i>	<i>Holodiscus dumosus</i>	<i>Holodiscus dumosus</i> (Nutt. ex Hook.) Heller	2	
<i>Hordeum</i>	<i>Hordeum brachyantherum</i>	<i>Hordeum brachyantherum</i> Nevski	1	
<i>Hydrophyllum</i>	<i>Hydrophyllum occidentale</i>	<i>Hydrophyllum occidentale</i> (S. Wats.) Gray	3	
<i>Hymenopappus</i>	<i>Hymenopappus filifolius</i>	<i>Hymenopappus filifolius</i> Hook.	20	
<i>Hymenopappus</i>	<i>Hymenopappus filifolius</i>	<i>Hymenopappus</i> L'Her.	1	
<i>Hypochaeris</i>	<i>Hypochaeris radicata</i>	<i>Hypochaeris radicata</i> L.	1	
<i>Ipomoea</i>	<i>Ipomoea purpurea</i>	<i>Ipomoea purpurea</i> (L.) Roth	1	
<i>Ipomopsis</i>	<i>Ipomopsis aggregata</i>	<i>Ipomopsis aggregata</i> (Pursh) V. Grant	3	
<i>Ipomopsis</i>	<i>Ipomopsis aggregata</i>	<i>Ipomopsis arizonica</i> (Greene) Wherry	1	
<i>Ipomopsis</i>	<i>Ipomopsis congesta</i>	<i>Ipomopsis congesta</i> ssp. <i>congesta</i> (Hook.) V. Grant	5	
<i>Ipomopsis</i>	<i>Ipomopsis congesta</i>	<i>Ipomopsis congesta</i> ssp. <i>frutescens</i> (Rydb.) Day	1	
<i>Ivesia</i>	<i>Ivesia sabulosa</i>	<i>Ivesia sabulosa</i> (M.E. Jones) Keck	3	
<i>Juncus</i>	<i>Juncus arcticus</i>	<i>Juncus balticus</i> Willd.	4	1
<i>Juncus</i>	<i>Juncus ensifolius</i>	<i>Juncus ensifolius</i> Wikstr.	2	
<i>Juncus</i>	<i>Juncus longistylis</i>	<i>Juncus longistylis</i> Torr.	3	
<i>Juncus</i>	<i>Juncus tenuis</i>	<i>Juncus tenuis</i> Willd.	3	
<i>Juncus</i>	<i>Juncus torreyi</i>	<i>Juncus torreyi</i> Coville	1	
<i>Juncus</i>		<i>Juncus</i> L.	2	1
<i>Juniperus</i>	<i>Juniperus osteosperma</i>	<i>Juniperus osteosperma</i> (Torr.) Little	104	12
<i>Juniperus</i>	<i>Juniperus scopulorum</i>	<i>Juniperus scopulorum</i> Sarg.	31	16
<i>Juniperus</i>		<i>Juniperus</i> L.	1	
<i>Kelloggia</i>	<i>Kelloggia galionoides</i>	<i>Kelloggia galionoides</i> Torr.	4	
<i>Koeleria</i>	<i>Koeleria macrantha</i>	<i>Koeleria macrantha</i> (Ledeb.) J.A. Schultes	6	
<i>Krameria</i>		<i>Krameria</i> L.	1	

Genus	Species	Database Name	#In	#Out
<i>Lactuca</i>	<i>Lactuca serriola</i>	<i>Lactuca serriola</i> L.	2	
<i>Lactuca</i>		<i>Lactuca</i> L.	3	
<i>Lathyrus</i>	<i>Lathyrus brachycalyx</i>	<i>Lathyrus brachycalyx</i> Rydb.	1	
<i>Lathyrus</i>		<i>Lathyrus</i> L.	3	
<i>Lathyrus</i>		<i>Lathyrus</i> L. ? <i>Vicia</i> ?	1	
<i>Lepidium</i>		<i>Lepidium</i> L.	3	
<i>Lepidium</i>		<i>Lepidium</i> species	1	
<i>Leptosiphon</i>	<i>Linanthastrum nuttallii</i>	<i>Linanthus nuttallii</i> ssp. <i>nuttallii</i> (Gray) Greene ex Milliken	1	
<i>Linanthus</i>	<i>Leptodactylon watsonii</i>	<i>Leptodactylon watsonii</i> (Gray) Rydb.	1	
<i>Linaria</i>	<i>Linaria dalmatica</i>	<i>Linaria dalmatica</i> (L.) P. Mill.	2	
<i>Linum</i>	<i>Linum perenne</i>	<i>Linum perenne</i> L.	4	
<i>Linum</i>		<i>Linum</i> L.	4	
<i>Lomatium</i>	<i>Lomatium graveolens</i>	<i>Lomatium graveolens</i> (S. Wats.) Dorn & Hartman	1	
<i>Lomatium</i>		<i>Lomatium</i> Raf.	3	
<i>Lotus</i>	<i>Lotus rigidus</i>	<i>Lotus rigidus</i> (Benth.) Greene	2	
<i>Lotus</i>	<i>Lotus utahensis</i>	<i>Lotus utahensis</i> Ottley	23	
<i>Lotus</i>		<i>Lotus</i> L.	5	
<i>Lupinus</i>	<i>Lupinus argenteus</i>	<i>Lupinus argenteus</i> Pursh	15	
<i>Lupinus</i>	<i>Lupinus concinnus</i>	<i>Lupinus concinnus</i> J.G. Agardh	1	
<i>Lupinus</i>	<i>Lupinus sericeus</i>	<i>Lupinus sericeus</i> Pursh	2	
<i>Lupinus</i>		<i>Lupinus</i> L.	18	
<i>Luzula</i>	<i>Luzula campestris</i>	<i>Luzula campestris</i> (L.) DC.	1	
<i>Luzula</i>	<i>Luzula parviflora</i>	<i>Luzula parviflora</i> (Ehrh.) Desv.	4	
<i>Lycium</i>	<i>Lycium pallidum</i>	<i>Lycium pallidum</i> Miers	4	
<i>Lycium</i>		<i>Lycium</i> L.	0	1
<i>Lygodesmia</i>	<i>Lygodesmia juncea</i>	<i>Lygodesmia juncea</i> (Pursh) D. Don ex Hook.	1	
<i>Lygodesmia</i>		skeletonweed	1	
<i>Machaeranthera</i>	<i>Haplopappus gracilis</i>	<i>Machaeranthera gracilis</i> (Nutt.) Shinners	2	
<i>Machaeranthera</i>	<i>Machaeranthera canescens</i>	<i>Machaeranthera canescens</i> (Pursh) Gray	18	
<i>Machaeranthera</i>		<i>Machaeranthera</i> Nees	5	
<i>Mahonia</i>	<i>Mahonia repens</i>	<i>Mahonia repens</i> (Lindl.) G. Don	39	
<i>Maianthemum</i>	<i>Smilacina racemosa</i>	<i>Maianthemum racemosum</i> ssp. <i>racemosum</i> (L.) Link	6	
<i>Maianthemum</i>	<i>Smilacina stellata</i>	<i>Maianthemum stellatum</i> (L.) Link	9	
<i>Medicago</i>	<i>Medicago lupulina</i>	<i>Medicago lupulina</i> L.	2	
<i>Melilotus</i>	<i>Melilotus officinalis</i>	<i>Melilotus officinalis</i> (L.) Lam.	14	
<i>Mentha</i>	<i>Mentha arvensis</i>	<i>Mentha arvensis</i> L.	3	
<i>Mentha</i>	<i>Mentha piperita</i>	<i>Mentha X piperita</i> L. (pro sp.)	1	
<i>Mertensia</i>	<i>Mertensia arizonica</i>	<i>Mertensia arizonica</i> Greene	10	
<i>Mertensia</i>		<i>Mertensia</i> Roth	3	
<i>Mirabilis</i>	<i>Mirabilis multiflora</i>	<i>Mirabilis multiflora</i> (Torr.) Gray	3	
<i>Monardella</i>	<i>Monardella odoratissima</i>	<i>Monardella odoratissima</i> Benth.	1	
<i>Muhlenbergia</i>	<i>Muhlenbergia asperifolia</i>	<i>Muhlenbergia asperifolia</i> (Nees & Meyen ex Trin.) Parodi	3	
<i>Muhlenbergia</i>	<i>Muhlenbergia montana</i>	<i>Muhlenbergia montana</i> (Nutt.) A.S. Hitchc.	8	
<i>Muhlenbergia</i>	<i>Muhlenbergia pauciflora</i>	<i>Muhlenbergia pauciflora</i> Buckl.	1	
<i>Muhlenbergia</i>	<i>Muhlenbergia porteri</i>	<i>Muhlenbergia porteri</i> Scribn. ex Beal	3	
<i>Muhlenbergia</i>	<i>Muhlenbergia racemosa</i>	<i>Muhlenbergia racemosa</i> (Michx.) B.S.P.	2	
<i>Muhlenbergia</i>	<i>Muhlenbergia thurberi</i>	<i>Muhlenbergia thurberi</i> Rydb.	1	

Genus	Species	Database Name	#In	#Out
<i>Muhlenbergia</i>	<i>Muhlenbergia wrightii</i>	<i>Muhlenbergia wrightii</i> Vasey ex Coult.	1	
<i>Muhlenbergia</i>		<i>Muhlenbergia</i> Schreb.	5	
<i>Nassella</i>	<i>Stipa viridula</i>	<i>Nassella viridula</i> (Trin.) Barkworth	2	
<i>Nasturtium</i>	<i>Nasturtium officinale</i>	<i>Rorippa nasturtium-aquaticum</i> (L.) Hayek	1	
<i>Nicotiana</i>	<i>Nicotiana attenuata</i>	<i>Nicotiana attenuata</i> Torr. ex S. Wats.	1	
<i>Oenothera</i>	<i>Oenothera caespitosa</i>	<i>Oenothera cespitosa</i> Nutt.	1	
<i>Oenothera</i>	<i>Oenothera flava</i>	<i>Oenothera flava</i> (A. Nels.) Garrett	1	
<i>Oenothera</i>	<i>Oenothera howardii</i>	<i>Oenothera howardii</i> (A. Nels.) W.L. Wagner	1	
<i>Oenothera</i>	<i>Oenothera pallida</i>	<i>Oenothera pallida</i> Lindl.	5	
<i>Oenothera</i>		<i>Oenothera</i> L.	7	
<i>Opuntia</i>	<i>Opuntia basilaris</i>	<i>Opuntia basilaris</i> Engelm. & Bigelow	8	1
<i>Opuntia</i>	<i>Opuntia chlorotica</i>	<i>Opuntia chlorotica</i> Engelm. & Bigelow	3	
<i>Opuntia</i>	<i>Opuntia echinocarpa</i>	<i>Opuntia echinocarpa</i> Engelm. & Bigelow	2	
<i>Opuntia</i>	<i>Opuntia erinacea</i>	<i>Opuntia erinacea</i> Engelm. & Bigelow ex Engelm.	5	
<i>Opuntia</i>	<i>Opuntia macrorhiza</i>	<i>Opuntia macrorhiza</i> Engelm.	93	2
<i>Opuntia</i>	<i>Opuntia phaeacantha</i>	<i>Opuntia engelmannii</i> Salm-Dyck	1	
<i>Opuntia</i>	<i>Opuntia phaeacantha</i>	<i>Opuntia phaeacantha</i> Engelm.	35	1
<i>Opuntia</i>	<i>Opuntia polyacantha</i>	<i>Opuntia polyacantha</i> Haw.	4	
<i>Opuntia</i>	<i>Opuntia whipplei</i>	<i>Opuntia whipplei</i> Engelm. & Bigelow	4	2
<i>Opuntia</i>		<i>Opuntia</i> P. Mill.	3	
<i>Osmorrhiza</i>	<i>Osmorrhiza depauperata</i>	<i>Osmorrhiza depauperata</i> Phil.	2	
<i>Osmorrhiza</i>	<i>Osmorrhiza occidentalis</i>	<i>Osmorrhiza occidentalis</i> (Nutt. ex Torr. & Gray) Torr.	6	
<i>Osmorrhiza</i>		<i>Osmorrhiza</i> Raf.	1	
<i>Oxytropis</i>		<i>Oxytropis</i> DC.	1	
<i>Pachystima</i>	<i>Pachystima myrsinoides</i>	<i>Paxistima myrsinoides</i> (Pursh) Raf.	38	
<i>Penstemon</i>	<i>Penstemon caespitosus</i>	<i>Penstemon caespitosus</i> Nutt. ex Gray	9	
<i>Penstemon</i>	<i>Penstemon caespitosus</i>	<i>Penstemon tusharensis</i> N. Holmgren	1	
<i>Penstemon</i>	<i>Penstemon eatonii</i>	<i>Penstemon eatonii</i> Gray	13	
<i>Penstemon</i>	<i>Penstemon higginsii</i>	<i>Penstemon leonardii</i> Rydb.	6	
<i>Penstemon</i>	<i>Penstemon higginsii</i>	<i>Penstemon leonardii</i> var. <i>higginsii</i> Neese	1	
<i>Penstemon</i>	<i>Penstemon humilis</i>	<i>Penstemon humilis</i> Nutt. ex Gray	3	
<i>Penstemon</i>	<i>Penstemon lentus</i>	<i>Penstemon lentus</i> Pennell	1	
<i>Penstemon</i>	<i>Penstemon linarioides</i>	<i>Penstemon linarioides</i> Gray	9	
<i>Penstemon</i>	<i>Penstemon pachyphyllus</i>	<i>Penstemon pachyphyllus</i> Gray ex Rydb.	2	
<i>Penstemon</i>	<i>Penstemon palmeri</i>	<i>Penstemon palmeri</i> Gray	13	2
<i>Penstemon</i>	<i>Penstemon rostriflorus</i>	<i>Penstemon rostriflorus</i> Kellogg	2	
<i>Penstemon</i>		<i>Penstemon</i> Schmidel	35	
<i>Peraphyllum</i>	<i>Peraphyllum ramosissimum</i>	<i>Peraphyllum ramosissimum</i> Nutt.	8	
<i>Petradoria</i>	<i>Petradoria pumila</i>	<i>Petradoria pumila</i> (Nutt.) Greene	7	
<i>Petrophytum</i>	<i>Petrophytum caespitosum</i>	<i>Petrophytum caespitosum</i> (Nutt.) Rydb.	3	
<i>Phacelia</i>	<i>Phacelia heterophylla</i>	<i>Phacelia heterophylla</i> Pursh	21	
<i>Phacelia</i>		<i>Phacelia</i> Juss.	4	
<i>Phleum</i>	<i>Phleum pratense</i>	<i>Phleum pratense</i> L.	1	
<i>Phlox</i>	<i>Phlox austromontana</i>	<i>Phlox austromontana</i> Coville	30	
<i>Phlox</i>	<i>Phlox hoodii</i>	<i>Phlox hoodii</i> Richards.	4	
<i>Phlox</i>	<i>Phlox longifolia</i>	<i>Phlox longifolia</i> Nutt.	1	

Genus	Species	Database Name	#In	#Out
<i>Phlox</i>	<i>Phlox pulvinata</i>	<i>Phlox caespitosa</i> Nutt.	1	
<i>Phlox</i>		<i>Phlox</i> L.	7	
<i>Phoradendron</i>	<i>Phoradendron juniperinum</i>	<i>Phoradendron juniperinum</i> Engelm.	2	
<i>Phragmites</i>	<i>Phragmites australis</i>	<i>Phragmites australis</i> (Cav.) Trin. ex Steud.	1	
<i>Physalis</i>	<i>Physalis hederifolia</i>	<i>Physalis hederifolia</i> Gray	1	
<i>Physalis</i>	<i>Physalis heterophylla</i>	<i>Physalis heterophylla</i> Nees	1	
<i>Physaria</i>	<i>Physaria chambersii</i>	<i>Physaria chambersii</i> Rollins	6	
<i>Physaria</i>	<i>Physaria newberryi</i>	<i>Physaria newberryi</i> Gray	1	
<i>Physaria</i>		<i>Physaria</i> (Nutt. ex Torr. & Gray) Gray	2	
<i>Pinus</i>	<i>Pinus edulis</i>	<i>Pinus edulis</i> Engelm.	43	3
<i>Pinus</i>	<i>Pinus edulis</i>	<i>Pinus edulis x monophylla</i> Engelm.	4	
<i>Pinus</i>	<i>Pinus monophylla</i>	<i>Pinus monophylla</i> Torr. & Frem.	52	5
<i>Pinus</i>	<i>Pinus ponderosa</i>	<i>Pinus ponderosa</i> P. & C. Lawson	52	14
<i>Plagiobothrys</i>	<i>Plagiobothrys tenellus</i>	<i>Plagiobothrys tenellus</i> (Nutt. ex Hook.) Gray	1	
<i>Plantago</i>	<i>Plantago patagonica</i>	<i>Plantago patagonica</i> Jacq.	3	
<i>Pluchea</i>	<i>Pluchea sericea</i>	<i>Pluchea sericea</i> (Nutt.) Coville	2	
<i>Poa</i>	<i>Poa fendleriana</i>	<i>Poa fendleriana</i> (Steud.) Vasey	147	1
<i>Poa</i>	<i>Poa pratensis</i>	<i>Poa pratensis</i> L.	43	
<i>Poa</i>	<i>Poa secunda</i>	<i>Poa secunda</i> J. Presl	12	
<i>Poa</i>		<i>Poa</i> L.	8	
<i>Poa</i>		<i>Poa</i> 2 L.	1	
<i>Poaceae</i> spp.		unknown grass	3	
<i>Polygonum</i>	<i>Polygonum douglasii</i>	<i>Polygonum douglasii</i> Greene	7	
<i>Polygonum</i>		<i>Polygonum</i> L.	3	
<i>Polypodium</i>	<i>Polypodium hesperium</i>	<i>Polypodium hesperium</i> Maxon	1	
<i>Polypogon</i>		<i>Polypogon</i> Desf.	1	
<i>Populus</i>	<i>Populus angustifolia</i>	<i>Populus angustifolia</i> James	1	
<i>Populus</i>	<i>Populus fremontii</i>	<i>Populus fremontii</i> S. Wats.	18	2
<i>Populus</i>	<i>Populus tremuloides</i>	<i>Populus tremuloides</i> Michx.	10	
<i>Potentilla</i>	<i>Potentilla glandulosa</i>	<i>Potentilla glandulosa</i> Lindl.	1	
<i>Prosopis</i>	<i>Prosopis glandulosa</i>	<i>Prosopis glandulosa</i> Torr.	1	
<i>Prunus</i>	<i>Prunus virginiana</i>	<i>Prunus virginiana</i> L.	14	
<i>Prunus</i>		<i>Prunus</i> L.	1	
<i>Pseudotsuga</i>	<i>Pseudotsuga menziesii</i>	<i>Pseudotsuga menziesii</i> (Mirbel) Franco	18	1
<i>Psoralidium</i>		<i>Psoralidium</i> Rydb.	1	
<i>Psorothamnus</i>	<i>Psorothamnus fremontii</i>	<i>Psorothamnus fremontii</i> (Torr. ex Gray) Barneby	6	2
<i>Psorothamnus</i>	<i>Psorothamnus fremontii</i>	<i>Psorothamnus fremontii</i> var. <i>fremontii</i> (Torr. ex Gray) Barneby	1	
<i>Pteridium</i>	<i>Pteridium aquilinum</i>	<i>Pteridium aquilinum</i> (L.) Kuhn	4	
<i>Pterospora</i>	<i>Pterospora andromedea</i>	<i>Pterospora andromedea</i> Nutt.	1	
<i>Puccinellia</i>	<i>Puccinellia distans</i>	<i>Puccinellia distans</i> (Jacq.) Parl.	1	
<i>Purshia</i>	<i>Purshia mexicana</i>	<i>Purshia mexicana</i> (D. Don) Henrickson	10	1
<i>Purshia</i>	<i>Purshia mexicana</i>	<i>Purshia stansburiana</i> (Torr.) Henrickson	6	
<i>Purshia</i>	<i>Purshia tridentata</i>	<i>Purshia tridentata</i> (Pursh) DC.	46	1
<i>Purshia</i>		<i>Purshia</i> DC. ex Poir.	1	
<i>Quercus</i>	<i>Quercus gambelii</i>	<i>Quercus gambelii</i> Nutt.	160	3
<i>Quercus</i>	<i>Quercus turbinella</i>	<i>Quercus turbinella</i> Greene	38	1

Genus	Species	Database Name	#In	#Out
<i>Rhus</i>	<i>Rhus aromatica</i>	<i>Rhus aromatica</i> Ait.	6	3
<i>Rhus</i>	<i>Rhus aromatica</i>	<i>Rhus trilobata</i> Nutt.	13	2
<i>Rhus</i>	<i>Rhus aromatica</i>	<i>Rhus trilobata</i> var. <i>trilobata</i> Nutt.	7	2
<i>Rosa</i>	<i>Rosa woodsii</i>	<i>Rosa woodsii</i> Lindl.	16	
<i>Rumex</i>	<i>Rumex acetosella</i>	<i>Rumex acetosella</i> L.	2	
<i>Rumex</i>	<i>Rumex hymenosepalus</i>	<i>Rumex hymenosepalus</i> Torr.	2	
<i>Salix</i>	<i>Salix eriocephala</i>	<i>Salix ligulifolia</i> (Ball) Ball ex Schneid.	2	
<i>Salix</i>	<i>Salix exigua</i>	<i>Salix exigua</i> Nutt.	11	1
<i>Salix</i>	<i>Salix gooddingii</i>	<i>Salix gooddingii</i> Ball	1	1
<i>Salix</i>	<i>Salix lasiolepis</i>	<i>Salix lasiolepis</i> Benth.		1
<i>Salix</i>	<i>Salix lucida</i>	<i>Salix lucida</i> Muhl.		1
<i>Salix</i>	<i>Salix lucida</i>	<i>Salix lucida</i> ssp. <i>lasiandra</i> (Benth.) E. Murr.		1
<i>Salix</i>	<i>Salix scouleriana</i>	<i>Salix scouleriana</i> Barratt ex Hook.	0	1
<i>Salix</i>		<i>Salix</i> L.	3	1
<i>Salsola</i>	<i>Salsola tragus</i>	<i>Salsola kali</i> ssp. <i>tragus</i> (L.) Celak.	1	
<i>Salvia</i>	<i>Salvia dorrii</i>	<i>Salvia dorrii</i> (Kellogg) Abrams	4	1
<i>Sambucus</i>		<i>Sambucus</i> L.		1
<i>Schizachyrium</i>	<i>Schizachyrium scoparium</i>	<i>Schizachyrium scoparium</i> (Michx.) Nash	1	
<i>Scirpus</i>	<i>Scirpus americanus</i>	<i>Schoenoplectus americanus</i> (Pers.) Volk. ex Schinz & R. Keller	1	
<i>Scirpus</i>	<i>Scirpus validus</i>	<i>Schoenoplectus tabernaemontani</i> (K.C. Gmel.) Palla	1	
<i>Scirpus</i>		<i>Scirpus</i> L.		1
<i>Senecio</i>	<i>Senecio eremophilus</i>	<i>Senecio eremophilus</i> Richards.	3	
<i>Senecio</i>	<i>Senecio integerrimus</i>	<i>Senecio integerrimus</i> Nutt.	1	
<i>Senecio</i>	<i>Senecio multilobatus</i>	<i>Senecio multilobatus</i> Torr. & Gray ex Gray	30	
<i>Senecio</i>	<i>Senecio spartioides</i>	<i>Senecio spartioides</i> Torr. & Gray	16	
<i>Senecio</i>		<i>Senecio</i> L.		13
<i>Shepherdia</i>	<i>Shepherdia rotundifolia</i>	<i>Shepherdia rotundifolia</i> Parry	11	
<i>Sisymbrium</i>	<i>Sisymbrium altissimum</i>	<i>Sisymbrium altissimum</i> L.	3	
<i>Sisymbrium</i>		<i>Sisymbrium</i> L.		1
<i>Sisyrinchium</i>	<i>Sisyrinchium demissum</i>	<i>Sisyrinchium demissum</i> Greene	2	
<i>Solanum</i>	<i>Solanum elaeagnifolium</i>	<i>Solanum elaeagnifolium</i> Cav.	1	
<i>Solidago</i>	<i>Solidago velutina</i>	<i>Solidago velutina</i> DC.	8	
<i>Solidago</i>		<i>Solidago</i> L.		1
<i>Sorghastrum</i>	<i>Sorghastrum nutans</i>	<i>Sorghastrum nutans</i> (L.) Nash	2	
<i>Sphaeralcea</i>	<i>Sphaeralcea ambigua</i>	<i>Sphaeralcea ambigua</i> Gray	2	
<i>Sphaeralcea</i>	<i>Sphaeralcea coccinea</i>	<i>Sphaeralcea coccinea</i> (Nutt.) Rydb.	10	2
<i>Sphaeralcea</i>	<i>Sphaeralcea grossulariifolia</i>	<i>Sphaeralcea grossulariifolia</i> (Hook. & Arn.) Rydb.	3	
<i>Sphaeralcea</i>		<i>Sphaeralcea</i> St.-Hil.		5
<i>Sphaeromeria</i>	<i>Sphaeromeria ruthiae</i>	<i>Sphaeromeria ruthiae</i> Holmgren, Shultz & Lowrey	1	1
<i>Sporobolus</i>	<i>Sporobolus cryptandrus</i>	<i>Sporobolus cryptandrus</i> (Torr.) Gray	56	
<i>Sporobolus</i>		<i>Sporobolus</i> R. Br.		1
<i>Stanleya</i>	<i>Stanleya pinnata</i>	<i>Stanleya pinnata</i> (Pursh) Britt.	3	
<i>Stellaria</i>	<i>Stellaria jamesiana</i>	<i>Pseudostellaria jamesiana</i> (Torr.) W.A. Weber & R.L. Hartman	3	
<i>Stellaria</i>	<i>Stellaria media</i>	<i>Stellaria media</i> (L.) Vill.	1	
<i>Stellaria</i>		<i>Stellaria</i> L.		2

Genus	Species	Database Name	#In	#Out
<i>Stephanomeria</i>	<i>Stephanomeria exigua</i>	<i>Stephanomeria exigua</i> Nutt.	1	
<i>Stephanomeria</i>	<i>Stephanomeria tenuifolia</i>	<i>Stephanomeria tenuifolia</i> (Raf.) Hall	1	
<i>Stephanomeria</i>		<i>Stephanomeria</i> Nutt.	2	
<i>Stipinae</i> spp.		<i>Stipa</i> L.	6	
<i>Stipinae</i> spp.		<i>Stipa</i> L. (comata or lettermanii)	1	
<i>Streptanthus</i>	<i>Streptanthus cordatus</i>	<i>Streptanthus cordatus</i> Nutt.	3	
<i>Swertia</i>	<i>Swertia radiata</i>	<i>Frasera speciosa</i> Dougl. ex Griseb.	11	
<i>Symphoricarpos</i>	<i>Symphoricarpos longiflorus</i>	<i>Symphoricarpos longiflorus</i> Gray	2	
<i>Symphoricarpos</i>	<i>Symphoricarpos oreophilus</i>	<i>Symphoricarpos oreophilus</i> Gray	78	
<i>Symphytum</i>	<i>Aster ascendens</i>	<i>Aster ascendens</i> Lindl.	1	
<i>Tamarix</i>	<i>Tamarix chinensis</i>	<i>Tamarix ramosissima</i> Ledeb.	8	2
<i>Taraxacum</i>	<i>Taraxacum officinale</i>	<i>Taraxacum officinale</i> G.H. Weber ex Wiggers	6	
<i>Tetradymia</i>	<i>Tetradymia axillaris</i>	<i>Tetradymia axillaris</i> A. Nels.	1	
<i>Tetradymia</i>	<i>Tetradymia canescens</i>	<i>Tetradymia canescens</i> DC.	9	1
<i>Thalictrum</i>	<i>Thalictrum fendleri</i>	<i>Thalictrum fendleri</i> Engelm. ex Gray	15	
<i>Thalictrum</i>		<i>Thalictrum</i> L.		1
<i>Thermopsis</i>	<i>Thermopsis montana</i>	<i>Thermopsis montana</i> Nutt.	2	
<i>Townsendia</i>		<i>Townsendia</i> Hook.	2	
<i>Toxicodendron</i>	<i>Toxicodendron rydbergii</i>	<i>Toxicodendron rydbergii</i> (Small ex Rydb.) Greene	2	
<i>Tradescantia</i>	<i>Tradescantia occidentalis</i>	<i>Tradescantia occidentalis</i> (Britt.) Smyth	13	
<i>Tragia</i>	<i>Tragia ramosa</i>	<i>Tragia ramosa</i> Torr.	1	
<i>Tragopogon</i>	<i>Tragopogon dubius</i>	<i>Tragopogon dubius</i> Scop.	21	
<i>Tribulus</i>	<i>Tribulus terrestris</i>	<i>Tribulus terrestris</i> L.	2	
<i>Trifolium</i>	<i>Trifolium gymnocarpon</i>	<i>Trifolium gymnocarpon</i> Nutt.	1	
<i>Trifolium</i>	<i>Trifolium longipes</i>	<i>Trifolium longipes</i> Nutt.	3	
<i>Trifolium</i>		<i>Trifolium</i> L.	3	
<i>Typha</i>	<i>Typha domingensis</i>	<i>Typha angustifolia</i> L.	2	
<i>Typha</i>	<i>Typha domingensis</i>	<i>Typha domingensis</i> Pers.	1	
<i>Verbascum</i>	<i>Verbascum thapsus</i>	<i>Verbascum thapsus</i> L.	12	
<i>Verbena</i>	<i>Verbena bracteata</i>	<i>Verbena bracteata</i> Lag. & Rodr.	2	
<i>Vicia</i>	<i>Vicia americana</i>	<i>Vicia americana</i> Muhl. ex Willd.	50	
<i>Vicia</i>		<i>Vicia</i> L.	5	
<i>Viguiera</i>	<i>Viguiera multiflora</i>	<i>Heliomeris multiflora</i> var. <i>multiflora</i> Nutt.	9	
<i>Viola</i>	<i>Viola purpurea</i>	<i>Viola charlestonensis</i> M.S. Baker & J.C. Clausen ex Clokey	1	
<i>Viola</i>		<i>Viola</i> L.	1	
<i>Vitis</i>	<i>Vitis arizonica</i>	<i>Vitis arizonica</i> Engelm.	4	
<i>Wyethia</i>	<i>Wyethia arizonica</i>	<i>Wyethia arizonica</i> Gray	2	
<i>Yucca</i>	<i>Yucca baccata</i>	<i>Yucca baccata</i> Torr.	19	1
<i>Yucca</i>	<i>Yucca utahensis</i>	<i>Yucca elata</i> var. <i>utahensis</i> (McKelvey) Reveal	65	8

APPENDIX 3.2

Results of niche overlap significance tests comparing congeneric species of vascular plants recorded in the USGS-NPS vegetation characterization of Zion National Park, Utah (Cogan et al. 2004). Null model distributions of beta niche overlap (across habitats) and alpha niche overlap (across plots within habitats), using both presence and cover measures, were built through randomization and compared with observed overlap values. Prob=probabilities ranging from -1 (more segregated than expected) to 1 (more aggregated than expected). Sig=cases of statistically significant segregation (-) or aggregation (+). Families are listed in phylogenetic sequence of Fig. 3.1.

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Equisetaceae	<i>Equisetum arvense / hyemale</i>	1.00	+	0.88		0.55		0.55	
Equisetaceae	<i>Equisetum arvense / laevigatum</i>	0.98	+	1.00	+	0.00		0.00	
Equisetaceae	<i>Equisetum arvense / variegatum</i>	0.97	+	0.75		0.00		0.00	
Equisetaceae	<i>Equisetum hyemale / laevigatum</i>	0.86		0.90		0.00		0.00	
Equisetaceae	<i>Equisetum hyemale / variegatum</i>	0.81		0.65		0.00		0.00	
Equisetaceae	<i>Equisetum laevigatum / variegatum</i>	0.98	+	0.71		0.00		0.00	
Cupressaceae	<i>Juniperus osteosperma / scopulorum</i>	-1.00	-	-0.90		-0.93		-0.15	
Pinaceae	<i>Pinus edulis / monophylla</i>	-0.94		-0.46		-1.00	-	-0.98	-
Pinaceae	<i>Pinus edulis / ponderosa</i>	-1.00	-	-1.00	-	-0.55		-0.95	-
Pinaceae	<i>Pinus monophylla / ponderosa</i>	-1.00	-	-1.00	-	-0.75		-0.92	
Ephedraceae	<i>Ephedra nevadensis / viridis</i>	-0.45		-0.98	-	-0.19		-0.80	
Cyperaceae	<i>Carex geyeri / microptera</i>	-0.47		-0.47		0.00		0.00	
Cyperaceae	<i>Carex geyeri / nebrascensis</i>	0.00		0.00		0.00		0.00	
Cyperaceae	<i>Carex geyeri / occidentalis</i>	-0.19		-0.52		0.00		0.00	
Cyperaceae	<i>Carex geyeri / rossii</i>	0.13		0.00		0.00		0.00	
Cyperaceae	<i>Carex geyeri / utriculata</i>	0.00		0.00		0.00		0.00	
Cyperaceae	<i>Carex microptera / nebrascensis</i>	1.00	+	0.99	+	0.50		0.79	
Cyperaceae	<i>Carex microptera / occidentalis</i>	-0.42		0.94		0.91		1.00	+
Cyperaceae	<i>Carex microptera / rossii</i>	-0.99	-	-0.99	-	0.00		0.00	
Cyperaceae	<i>Carex microptera / utriculata</i>	1.00	+	0.99	+	0.53		0.53	
Cyperaceae	<i>Carex nebrascensis / occidentalis</i>	0.10		0.90		0.00		0.00	
Cyperaceae	<i>Carex nebrascensis / rossii</i>	-0.79		-0.79		0.00		0.00	
Cyperaceae	<i>Carex nebrascensis / utriculata</i>	0.97	+	0.99	+	0.00		0.00	
Cyperaceae	<i>Carex occidentalis / rossii</i>	-0.03		-0.84		0.45		0.45	
Cyperaceae	<i>Carex occidentalis / utriculata</i>	0.03		0.90		0.75		0.90	
Cyperaceae	<i>Carex rossii / utriculata</i>	-0.79		-0.79		0.00		0.00	
Cyperaceae	<i>Scirpus americanus / validus</i>	0.87		0.87		0.00		0.00	
Juncaceae	<i>Juncus arcticus / ensifolius</i>	1.00	+	1.00	+	0.41		0.53	
Juncaceae	<i>Juncus arcticus / longistylis</i>	1.00	+	1.00	+	0.21		0.39	
Juncaceae	<i>Juncus arcticus / tenuis</i>	1.00	+	1.00	+	0.15		0.29	
Juncaceae	<i>Juncus arcticus / torreyi</i>	1.00	+	0.97	+	0.00		0.00	

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Juncaceae	<i>Juncus ensifolius / longistylis</i>	1.00	+	1.00	+	0.97	+	0.97	+
Juncaceae	<i>Juncus ensifolius / tenuis</i>	1.00	+	0.99	+	0.97	+	0.91	
Juncaceae	<i>Juncus ensifolius / torreyi</i>	0.96	+	0.93		0.00		0.00	
Juncaceae	<i>Juncus longistylis / tenuis</i>	1.00	+	1.00	+	1.00	+	1.00	+
Juncaceae	<i>Juncus longistylis / torreyi</i>	1.00	+	1.00	+	0.00		0.00	
Juncaceae	<i>Juncus tenuis / torreyi</i>	1.00	+	0.98	+	0.00		0.00	
Juncaceae	<i>Luzula campestris / parviflora</i>	1.00	+	1.00	+	0.00		0.00	
Poaceae	<i>Aristida purpurascens / purpurea</i>	-0.87		-0.94		0.00		0.00	
Poaceae	<i>Muhlenbergia asperifolia / montana</i>	-0.91		-0.91		0.00		0.00	
Poaceae	<i>Muhlenbergia asperifolia / pauciflora</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia asperifolia / porteri</i>	-0.43		-0.43		0.00		0.00	
Poaceae	<i>Muhlenbergia asperifolia / racemosa</i>	0.90		0.88		0.00		0.00	
Poaceae	<i>Muhlenbergia asperifolia / thurberi</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia asperifolia / wrightii</i>	1.00	+	0.98	+	0.00		0.00	
Poaceae	<i>Muhlenbergia montana / pauciflora</i>	-0.24		-0.24		0.00		0.00	
Poaceae	<i>Muhlenbergia montana / porteri</i>	-0.90		-0.90		0.00		0.00	
Poaceae	<i>Muhlenbergia montana / racemosa</i>	-0.73		-0.73		0.00		0.00	
Poaceae	<i>Muhlenbergia montana / thurberi</i>	-0.34		-0.34		0.00		0.00	
Poaceae	<i>Muhlenbergia montana / wrightii</i>	-0.35		-0.35		0.00		0.00	
Poaceae	<i>Muhlenbergia pauciflora / porteri</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia pauciflora / racemosa</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia pauciflora / thurberi</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia pauciflora / wrightii</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia porteri / racemosa</i>	-0.07		-0.07		0.00		0.00	
Poaceae	<i>Muhlenbergia porteri / thurberi</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia porteri / wrightii</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia racemosa / thurberi</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Muhlenbergia racemosa / wrightii</i>	0.87		0.87		0.00		0.00	
Poaceae	<i>Muhlenbergia thurberi / wrightii</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Bouteloua barbata / gracilis</i>	-0.81		-0.71		0.00		0.00	
Poaceae	<i>Stipa hymenoides / lettermanii</i>	-0.99	-	-0.09		0.17		0.73	
Poaceae	<i>Stipa hymenoides / nelsonii</i>	-0.98	-	-0.97	-	0.00		0.00	
Poaceae	<i>Stipa hymenoides / speciosa</i>	-0.70		-0.84		-0.30		-0.24	
Poaceae	<i>Stipa lettermanii / nelsonii</i>	-0.18		-0.66		0.46		0.46	
Poaceae	<i>Stipa lettermanii / speciosa</i>	-0.99	-	-0.99	-	0.00		0.00	
Poaceae	<i>Stipa nelsonii / speciosa</i>	-0.47		-0.47		0.00		0.00	
Poaceae	<i>Poa fendleriana / pratensis</i>	-1.00	-	-1.00	-	-0.94		-0.95	-
Poaceae	<i>Poa fendleriana / secunda</i>	-0.11		0.87		-0.55		0.46	
Poaceae	<i>Poa pratensis / secunda</i>	-1.00	-	-0.98	-	-0.39		-0.39	
Poaceae	<i>Festuca occidentalis / octoflora</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Agrostis exarata / stolonifera</i>	1.00	+	1.00	+	0.00		0.00	
Poaceae	<i>Bromus anomalus / diandrus</i>	0.22		0.01		0.00		0.00	
Poaceae	<i>Bromus anomalus / inermis</i>	-0.89		-0.91		0.00		0.00	
Poaceae	<i>Bromus anomalus / rubens</i>	-0.45		-0.63		0.00		0.00	
Poaceae	<i>Bromus anomalus / tectorum</i>	-0.96	-	0.45		0.03		0.05	

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Poaceae	<i>Bromus diandrus / inermis</i>	-0.99	-	-0.99	-	-0.07		-0.01	
Poaceae	<i>Bromus diandrus / rubens</i>	-0.17		-0.37		0.00		0.00	
Poaceae	<i>Bromus diandrus / tectorum</i>	-0.98	-	-0.85		0.76		0.75	
Poaceae	<i>Bromus inermis / rubens</i>	-0.95	-	-0.99	-	0.00		0.00	
Poaceae	<i>Bromus inermis / tectorum</i>	-1.00	-	-1.00	-	-0.56		-0.65	
Poaceae	<i>Bromus rubens / tectorum</i>	-0.15		-0.27		0.77		0.27	
Poaceae	<i>Elymus canadensis / elymoides</i>	-0.99	-	-0.98	-	0.00		0.00	
Poaceae	<i>Elymus canadensis / glaucus</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Elymus canadensis / hispidus</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Elymus canadensis / lanceolatus</i>	-0.43		-0.65		0.00		0.00	
Poaceae	<i>Elymus canadensis / repens</i>	-0.57		-0.57		0.00		0.00	
Poaceae	<i>Elymus canadensis / smithii</i>	0.63		0.40		0.00		0.00	
Poaceae	<i>Elymus canadensis / spicatus</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Elymus elymoides / glaucus</i>	-0.71		0.03		0.00		0.00	
Poaceae	<i>Elymus elymoides / hispidus</i>	-0.66		-0.42		0.00		0.00	
Poaceae	<i>Elymus elymoides / lanceolatus</i>	-0.99	-	-0.97	-	0.38		0.95	+
Poaceae	<i>Elymus elymoides / repens</i>	-0.50		0.87		-0.58		-0.58	
Poaceae	<i>Elymus elymoides / smithii</i>	-1.00	-	-1.00	-	-0.36		-0.36	
Poaceae	<i>Elymus elymoides / spicatus</i>	0.92		0.93		0.48		0.91	
Poaceae	<i>Elymus glaucus / hispidus</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Elymus glaucus / lanceolatus</i>	-0.37		-0.36		0.00		0.00	
Poaceae	<i>Elymus glaucus / repens</i>	0.18		0.50		0.00		0.00	
Poaceae	<i>Elymus glaucus / smithii</i>	-0.32		-0.34		0.00		0.00	
Poaceae	<i>Elymus glaucus / spicatus</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Elymus hispidus / lanceolatus</i>	0.85		0.88		0.00		0.00	
Poaceae	<i>Elymus hispidus / repens</i>	-0.41		-0.14		0.00		0.00	
Poaceae	<i>Elymus hispidus / smithii</i>	0.40		0.49		0.00		0.00	
Poaceae	<i>Elymus hispidus / spicatus</i>	0.00		0.00		0.00		0.00	
Poaceae	<i>Elymus lanceolatus / repens</i>	-0.69		-0.68		0.00		0.00	
Poaceae	<i>Elymus lanceolatus / smithii</i>	-0.15		0.14		-0.10		-0.10	
Poaceae	<i>Elymus lanceolatus / spicatus</i>	0.24		-0.14		0.00		0.00	
Poaceae	<i>Elymus repens / smithii</i>	-0.91		-0.86		0.61		0.91	
Poaceae	<i>Elymus repens / spicatus</i>	0.97	+	0.87		0.00		0.00	
Poaceae	<i>Elymus smithii / spicatus</i>	-0.33		-0.33		0.00		0.00	
Ruscaceae	<i>Smilacina racemosa / stellata</i>	-0.38		-0.73		-0.24		-0.24	
Agavaceae	<i>Yucca baccata / utahensis</i>	-0.98	-	0.50		-0.43		0.55	
Ranunculaceae	<i>Clematis columbiana / ligusticifolia</i>	0.17		0.43		0.00		0.00	
Fagaceae	<i>Quercus gambelii / turbinella</i>	-1.00	-	-1.00	-	-0.98	-	-0.99	-
Rosaceae	<i>Cercocarpus intricatus / ledifolius</i>	-0.39		0.73		0.00		0.00	
Rosaceae	<i>Cercocarpus intricatus / montanus</i>	-0.93		0.62		-0.45		-0.35	
Rosaceae	<i>Cercocarpus ledifolius / montanus</i>	0.68		0.80		0.13		0.42	
Rosaceae	<i>Purshia mexicana / tridentata</i>	-0.69		0.85		-0.96	-	-0.95	
Rosaceae	<i>Amelanchier alnifolia / utahensis</i>	-1.00	-	-0.89		-0.97	-	-0.33	
Rhamnaceae	<i>Ceanothus fendleri / martinii</i>	0.83		0.35		0.00		0.00	

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Fabaceae	<i>Lupinus argenteus / concinnus</i>	0.42		0.62		0.00		0.00	
Fabaceae	<i>Lupinus argenteus / sericeus</i>	-0.76		-0.57		0.00		0.00	
Fabaceae	<i>Lupinus concinnus / sericeus</i>	0.00		0.00		0.00		0.00	
Fabaceae	<i>Lotus rigidus / utahensis</i>	0.18		-0.37		0.00		0.00	
Fabaceae	<i>Astragalus flavus / subcinereus</i>	0.00		0.00		0.00		0.00	
Fabaceae	<i>Trifolium gymnocarpon / longipes</i>	0.00		0.00		0.00		0.00	
Euphorbiaceae	<i>Chamaesyce albomarginata / fendleri</i>	-0.19		-0.19		0.00		0.00	
Euphorbiaceae	<i>Chamaesyce albomarginata / glyptosperma</i>	0.71		0.71		0.00		0.00	
Euphorbiaceae	<i>Chamaesyce albomarginata / parryi</i>	0.00		0.00		0.00		0.00	
Euphorbiaceae	<i>Chamaesyce fendleri / glyptosperma</i>	0.34		0.57		0.00		0.00	
Euphorbiaceae	<i>Chamaesyce fendleri / parryi</i>	0.19		0.19		0.00		0.00	
Euphorbiaceae	<i>Chamaesyce glyptosperma / parryi</i>	0.00		0.00		0.00		0.00	
Salicaceae	<i>Populus angustifolia / fremontii</i>	1.00	+	1.00	+	0.00		0.46	
Salicaceae	<i>Populus angustifolia / tremuloides</i>	-0.44		-0.44		0.00		0.00	
Salicaceae	<i>Populus fremontii / tremuloides</i>	-1.00	-	-1.00	-	0.00		0.00	
Salicaceae	<i>Salix eriocephala / exigua</i>	1.00	+	1.00	+	0.00		0.00	
Salicaceae	<i>Salix eriocephala / gooddingii</i>	0.99	+	0.90		0.00		0.00	
Salicaceae	<i>Salix eriocephala / lasiolepis</i>	0.88		0.88		0.00		0.00	
Salicaceae	<i>Salix eriocephala / lucida</i>	0.99	+	0.92		0.00		0.00	
Salicaceae	<i>Salix eriocephala / scouleriana</i> ¹	0.00				0.00			
Salicaceae	<i>Salix exigua / gooddingii</i>	1.00	+	1.00	+	0.07		0.00	
Salicaceae	<i>Salix exigua / lasiolepis</i>	1.00	+	1.00	+	0.00		0.00	
Salicaceae	<i>Salix exigua / lucida</i>	1.00	+	1.00	+	0.07		0.50	
Salicaceae	<i>Salix exigua / scouleriana</i> ¹	-0.54				0.00			
Salicaceae	<i>Salix gooddingii / lasiolepis</i>	0.99	+	0.92		0.00		0.00	
Salicaceae	<i>Salix gooddingii / lucida</i>	0.99	+	1.00	+	0.76		0.94	
Salicaceae	<i>Salix gooddingii / scouleriana</i> ¹	0.00				0.00			
Salicaceae	<i>Salix lasiolepis / lucida</i>	0.98	+	0.92		0.00		0.00	
Salicaceae	<i>Salix lasiolepis / scouleriana</i> ¹	0.00				0.00			
Salicaceae	<i>Salix lucida / scouleriana</i> ¹	0.00				0.00			
Brassicaceae	<i>Arabis holboellii / perennans</i>	0.73		0.73		0.00		0.00	
Brassicaceae	<i>Physaria chambersii / newberryi</i>	0.32		0.32		0.00		0.00	
Malvaceae	<i>Sphaeralcea ambigua / coccinea</i>	-0.09		0.49		0.00		0.00	
Malvaceae	<i>Sphaeralcea ambigua / grossulariifolia</i>	-0.09		-0.09		0.00		0.00	
Malvaceae	<i>Sphaeralcea coccinea / grossulariifolia</i>	-0.07		-0.24		0.00		0.00	
Sapindaceae	<i>Acer grandidentatum / negundo</i>	-1.00	-	-1.00	-	0.84		0.46	
Onagraceae	<i>Epilobium brachycarpum / latifolia</i>	0.76		0.76		0.00		0.00	
Onagraceae	<i>Oenothera caespitosa / flava</i>	0.00		0.00		0.00		0.00	
Onagraceae	<i>Oenothera caespitosa / howardii</i>	0.00		0.00		0.00		0.00	
Onagraceae	<i>Oenothera caespitosa / pallida</i>	0.00		0.00		0.00		0.00	
Onagraceae	<i>Oenothera flava / howardii</i>	0.00		0.00		0.00		0.00	
Onagraceae	<i>Oenothera flava / pallida</i>	-0.01		-0.01		0.00		0.00	
Onagraceae	<i>Oenothera howardii / pallida</i>	0.42		0.42		0.00		0.00	

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Polygonaceae	<i>Eriogonum corymbosum / fasciculatum</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum corymbosum / flavum</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum corymbosum / inflatum</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum corymbosum / microthecum</i>	0.01		0.12		0.00		0.00	
Polygonaceae	<i>Eriogonum corymbosum / ovalifolium</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum corymbosum / panguicense</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum corymbosum / racemosum</i>	-0.21		-0.21		0.00		0.00	
Polygonaceae	<i>Eriogonum corymbosum / umbellatum</i>	-0.55		-0.63		0.00		0.00	
Polygonaceae	<i>Eriogonum fasciculatum / flavum</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum fasciculatum / inflatum</i>	0.32		0.32		0.00		0.00	
Polygonaceae	<i>Eriogonum fasciculatum / microthecum</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum fasciculatum / ovalifolium</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum fasciculatum / panguicense</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum fasciculatum / racemosum</i>	-0.77		-0.77		0.00		0.00	
Polygonaceae	<i>Eriogonum fasciculatum / umbellatum</i>	-0.25		0.16		0.63		0.63	
Polygonaceae	<i>Eriogonum flavum / inflatum</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum flavum / microthecum</i>	0.98	+	0.97	+	0.00		0.00	
Polygonaceae	<i>Eriogonum flavum / ovalifolium</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum flavum / panguicense</i>	0.57		0.83		0.00		0.00	
Polygonaceae	<i>Eriogonum flavum / racemosum</i>	0.85		0.85		0.00		0.00	
Polygonaceae	<i>Eriogonum flavum / umbellatum</i>	1.00	+	0.98	+	0.00		0.00	
Polygonaceae	<i>Eriogonum inflatum / microthecum</i>	-0.72		-0.59		0.00		0.00	
Polygonaceae	<i>Eriogonum inflatum / ovalifolium</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum inflatum / panguicense</i>	0.28		0.05		0.00		0.00	
Polygonaceae	<i>Eriogonum inflatum / racemosum</i>	-0.33		-0.33		0.00		0.00	
Polygonaceae	<i>Eriogonum inflatum / umbellatum</i>	-0.71		-0.20		0.00		0.00	
Polygonaceae	<i>Eriogonum microthecum / ovalifolium</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum microthecum / panguicense</i>	0.45		0.98	+	0.00		0.00	
Polygonaceae	<i>Eriogonum microthecum / racemosum</i>	-0.13		0.33		0.00		0.00	
Polygonaceae	<i>Eriogonum microthecum / umbellatum</i>	0.68		0.75		0.00		0.00	
Polygonaceae	<i>Eriogonum ovalifolium / panguicense</i>	0.00		0.00		0.00		0.00	
Polygonaceae	<i>Eriogonum ovalifolium / racemosum</i>	-0.74		-0.74		0.00		0.00	
Polygonaceae	<i>Eriogonum ovalifolium / umbellatum</i>	-0.76		-0.77		0.00		0.00	
Polygonaceae	<i>Eriogonum panguicense / racemosum</i>	0.96	+	0.99	+	0.00		0.00	
Polygonaceae	<i>Eriogonum panguicense / umbellatum</i>	0.81		1.00	+	0.00		0.00	
Polygonaceae	<i>Eriogonum racemosum / umbellatum</i>	-0.29		0.25		0.99	+	0.97	+
Polygonaceae	<i>Rumex acetosella / hymenosepalus</i>	0.00		0.00		0.00		0.00	
Amaranthaceae	<i>Chenopodium album / fremontii</i>	-0.36		-0.36		0.00		0.00	
Caryophyllaceae	<i>Arenaria fendleri / macradenia</i>	0.49		0.96	+	0.04		0.60	
Caryophyllaceae	<i>Stellaria jamesiana / media</i>	0.00		0.00		0.00		0.00	
Cactaceae	<i>Echinocereus engelmannii / triglochidiatus</i>	1.00	+	1.00	+	-0.45		-0.31	
Cactaceae	<i>Opuntia basilaris / chlorotica</i>	-0.08		-0.10		0.00		0.00	
Cactaceae	<i>Opuntia basilaris / echinocarpa</i>	-0.79		-0.73		0.00		0.00	
Cactaceae	<i>Opuntia basilaris / erinacea</i>	0.28		-0.28		0.00		0.00	
Cactaceae	<i>Opuntia basilaris / macrorhiza</i>	0.73		0.18		-0.98	-	-0.95	

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Cactaceae	<i>Opuntia basilaris</i> / <i>phaeacantha</i>	0.67		0.83		-0.51		-0.38	
Cactaceae	<i>Opuntia basilaris</i> / <i>polyacantha</i>	0.92		0.68		0.99	+	0.98	+
Cactaceae	<i>Opuntia basilaris</i> / <i>whipplei</i>	0.11		-0.33		0.00		0.00	
Cactaceae	<i>Opuntia chlorotica</i> / <i>echinocarpa</i>	-0.20		-0.20		0.00		0.00	
Cactaceae	<i>Opuntia chlorotica</i> / <i>erinacea</i>	0.14		0.75		0.00		0.00	
Cactaceae	<i>Opuntia chlorotica</i> / <i>macrorhiza</i>	0.08		-0.01		0.95	+	0.59	
Cactaceae	<i>Opuntia chlorotica</i> / <i>phaeacantha</i>	-0.10		-0.12		0.00		0.00	
Cactaceae	<i>Opuntia chlorotica</i> / <i>polyacantha</i>	-0.30		0.51		0.00		0.00	
Cactaceae	<i>Opuntia chlorotica</i> / <i>whipplei</i>	0.08		0.04		0.00		0.00	
Cactaceae	<i>Opuntia echinocarpa</i> / <i>erinacea</i>	-0.48		-0.48		0.87		0.87	
Cactaceae	<i>Opuntia echinocarpa</i> / <i>macrorhiza</i>	-0.47		0.31		-0.31		-0.31	
Cactaceae	<i>Opuntia echinocarpa</i> / <i>phaeacantha</i>	-0.53		-0.56		0.00		0.00	
Cactaceae	<i>Opuntia echinocarpa</i> / <i>polyacantha</i>	-0.17		-0.23		0.00		0.00	
Cactaceae	<i>Opuntia echinocarpa</i> / <i>whipplei</i>	-0.14		0.62		0.00		0.00	
Cactaceae	<i>Opuntia erinacea</i> / <i>macrorhiza</i>	0.04		-0.10		-0.63		-0.94	
Cactaceae	<i>Opuntia erinacea</i> / <i>phaeacantha</i>	0.31		-0.32		0.84		0.15	
Cactaceae	<i>Opuntia erinacea</i> / <i>polyacantha</i>	-0.04		0.76		0.00		0.00	
Cactaceae	<i>Opuntia erinacea</i> / <i>whipplei</i>	0.98	+	0.17		0.94		0.00	
Cactaceae	<i>Opuntia macrorhiza</i> / <i>phaeacantha</i>	0.35		0.20		-0.93		-0.93	
Cactaceae	<i>Opuntia macrorhiza</i> / <i>polyacantha</i>	1.00	+	0.87		-0.69		-0.69	
Cactaceae	<i>Opuntia macrorhiza</i> / <i>whipplei</i>	0.44		0.68		-0.01		0.99	+
Cactaceae	<i>Opuntia phaeacantha</i> / <i>polyacantha</i>	0.87		0.86		0.00		0.00	
Cactaceae	<i>Opuntia phaeacantha</i> / <i>whipplei</i>	0.76		0.40		0.99	+	0.95	+
Cactaceae	<i>Opuntia polyacantha</i> / <i>whipplei</i>	0.52		0.50		0.00		0.00	
Polemoniaceae	<i>Phlox austromontana</i> / <i>hoodii</i>	1.00	+	1.00	+	-0.18		-0.18	
Polemoniaceae	<i>Phlox austromontana</i> / <i>longifolia</i>	-0.80		-0.83		0.00		0.00	
Polemoniaceae	<i>Phlox austromontana</i> / <i>pulvinata</i>	0.99	+	0.99	+	0.00		0.00	
Polemoniaceae	<i>Phlox hoodii</i> / <i>longifolia</i>	0.00		0.00		0.00		0.00	
Polemoniaceae	<i>Phlox hoodii</i> / <i>pulvinata</i>	0.84		0.84		0.00		0.00	
Polemoniaceae	<i>Phlox longifolia</i> / <i>pulvinata</i>	0.00		0.00		0.00		0.00	
Polemoniaceae	<i>Ipomopsis aggregata</i> / <i>congesta</i>	-0.49		0.90		0.00		0.00	
Ericaceae	<i>Arctostaphylos patula</i> / <i>pungens</i>	0.44		-0.14		0.67		-0.44	
Lamiaceae	<i>Mentha arvensis</i> / <i>piperita</i>	0.00		0.00		0.00		0.00	
Orobanchaceae	<i>Castilleja chromosa</i> / <i>flava</i>	0.37		0.37		0.00		0.00	
Orobanchaceae	<i>Castilleja chromosa</i> / <i>linariifolia</i>	-0.38		-0.38		0.00		0.00	
Orobanchaceae	<i>Castilleja flava</i> / <i>linariifolia</i>	0.50		0.50		0.00		0.00	
Orobanchaceae	<i>Cordylanthus kingii</i> / <i>parviflorus</i>	-0.34		-0.18		0.00		0.00	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>eatonii</i>	0.41		0.93		0.30		0.30	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>higginsii</i>	-0.40		0.83		0.00		0.00	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>humilis</i>	0.30		0.91		0.00		0.00	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>lentus</i>	0.96	+	0.58		0.00		0.00	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>linarioides</i>	0.80		0.03		0.00		0.00	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>pachyphyllus</i>	-0.81		-0.81		0.00		0.00	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>palmeri</i>	0.69		-0.06		-0.13		-0.04	
Plantaginaceae	<i>Penstemon caespitosus</i> / <i>rostriflorus</i>	0.47		0.68		0.00		0.00	

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Plantaginaceae	<i>Penstemon eatonii / higginsii</i>	-0.62		0.58		0.00		0.00	
Plantaginaceae	<i>Penstemon eatonii / humilis</i>	-0.33		0.49		0.00		0.00	
Plantaginaceae	<i>Penstemon eatonii / latus</i>	0.75		0.56		0.00		0.00	
Plantaginaceae	<i>Penstemon eatonii / linarioides</i>	0.16		-0.25		0.00		0.00	
Plantaginaceae	<i>Penstemon eatonii / pachyphyllus</i>	-0.89		-0.89		0.00		0.00	
Plantaginaceae	<i>Penstemon eatonii / palmeri</i>	-0.74		-0.72		0.86		0.89	
Plantaginaceae	<i>Penstemon eatonii / rostriflorus</i>	0.71		0.78		0.00		0.00	
Plantaginaceae	<i>Penstemon higginsii / humilis</i>	0.76		0.98	+	0.00		0.00	
Plantaginaceae	<i>Penstemon higginsii / latus</i>	-0.19		-0.19		0.00		0.00	
Plantaginaceae	<i>Penstemon higginsii / linarioides</i>	-0.44		-0.81		0.00		0.00	
Plantaginaceae	<i>Penstemon higginsii / pachyphyllus</i>	-0.66		-0.66		0.00		0.00	
Plantaginaceae	<i>Penstemon higginsii / palmeri</i>	-0.99	-	-0.96	-	0.00		0.00	
Plantaginaceae	<i>Penstemon higginsii / rostriflorus</i>	0.60		0.68		0.00		0.00	
Plantaginaceae	<i>Penstemon humilis / latus</i>	0.00		0.00		0.00		0.00	
Plantaginaceae	<i>Penstemon humilis / linarioides</i>	-0.37		-0.65		0.00		0.00	
Plantaginaceae	<i>Penstemon humilis / pachyphyllus</i>	-0.14		-0.14		0.00		0.00	
Plantaginaceae	<i>Penstemon humilis / palmeri</i>	-0.79		-0.72		0.00		0.00	
Plantaginaceae	<i>Penstemon humilis / rostriflorus</i>	0.75		0.75		0.00		0.00	
Plantaginaceae	<i>Penstemon latus / linarioides</i>	0.85		0.86		0.00		0.00	
Plantaginaceae	<i>Penstemon latus / pachyphyllus</i>	0.00		0.00		0.00		0.00	
Plantaginaceae	<i>Penstemon latus / palmeri</i>	1.00	+	0.87		0.00		0.00	
Plantaginaceae	<i>Penstemon latus / rostriflorus</i>	0.00		0.00		0.00		0.00	
Plantaginaceae	<i>Penstemon linarioides / pachyphyllus</i>	-0.40		0.45		0.00		0.00	
Plantaginaceae	<i>Penstemon linarioides / palmeri</i>	0.84		1.00	+	-0.07		0.00	
Plantaginaceae	<i>Penstemon linarioides / rostriflorus</i>	0.01		-0.32		0.00		0.00	
Plantaginaceae	<i>Penstemon pachyphyllus / palmeri</i>	-0.28		0.58		0.00		0.00	
Plantaginaceae	<i>Penstemon pachyphyllus / rostriflorus</i>	0.00		0.00		0.00		0.00	
Plantaginaceae	<i>Penstemon palmeri / rostriflorus</i>	-0.49		-0.36		0.00		0.00	
Oleaceae	<i>Fraxinus anomala / velutina</i>	-0.99	-	0.52		0.67		0.03	
Solanaceae	<i>Physalis hederifolia / heterophylla</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Cirsium arizonicum / calcareum</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Cirsium arizonicum / vulgare</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Cirsium arizonicum / wheeleri</i>	0.10		0.76		0.00		0.00	
Asteraceae	<i>Cirsium calcareum / vulgare</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Cirsium calcareum / wheeleri</i>	0.13		0.10		0.00		0.00	
Asteraceae	<i>Cirsium vulgare / wheeleri</i>	-0.11		-0.11		0.00		0.00	
Asteraceae	<i>Stephanomeria exigua / tenuifolia</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Senecio eremophilus / integerrimus</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Senecio eremophilus / multilobatus</i>	-0.91		-0.91		0.00		0.00	
Asteraceae	<i>Senecio eremophilus / spartioides</i>	-0.84		-0.89		0.00		0.00	
Asteraceae	<i>Senecio integrifolius / multilobatus</i>	-0.89		-0.89		0.00		0.00	
Asteraceae	<i>Senecio integrifolius / spartioides</i>	-0.36		0.10		0.00		0.00	
Asteraceae	<i>Senecio multilobatus / spartioides</i>	-1.00	-	-1.00	-	-0.32		-0.32	
Asteraceae	<i>Tetradymia axillaris / canescens</i>	0.42		0.79		0.00		0.00	

Family	Congeneric Pair	Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Asteraceae	<i>Artemisia campestris / dracunculus</i>	0.44		0.76		-0.08		0.03	
Asteraceae	<i>Artemisia campestris / filifolia</i>	-0.99	-	-0.98	-	0.00		0.00	
Asteraceae	<i>Artemisia campestris / ludoviciana</i>	-0.41		0.08		-0.84		-0.83	
Asteraceae	<i>Artemisia campestris / nova</i>	-0.70		-0.92		0.31		-0.11	
Asteraceae	<i>Artemisia campestris / tridentata</i>	-0.29		-0.93		0.71		0.66	
Asteraceae	<i>Artemisia dracunculus / filifolia</i>	-0.92		-0.41		0.66		0.42	
Asteraceae	<i>Artemisia dracunculus / ludoviciana</i>	-0.42		-0.11		0.30		0.98	+
Asteraceae	<i>Artemisia dracunculus / nova</i>	-0.70		-0.93		-0.07		-0.02	
Asteraceae	<i>Artemisia dracunculus / tridentata</i>	-0.08		0.36		0.14		0.53	
Asteraceae	<i>Artemisia filifolia / ludoviciana</i>	-0.99	-	-0.96	-	-0.50		-0.33	
Asteraceae	<i>Artemisia filifolia / nova</i>	-1.00	-	-1.00	-	0.00		0.00	
Asteraceae	<i>Artemisia filifolia / tridentata</i>	-1.00	-	0.41		-0.98	-	-0.98	-
Asteraceae	<i>Artemisia ludoviciana / nova</i>	-0.85		-0.62		-0.11		-0.34	
Asteraceae	<i>Artemisia ludoviciana / tridentata</i>	-0.45		-0.95		-0.97	-	-0.98	-
Asteraceae	<i>Artemisia nova / tridentata</i>	-0.98	-	-1.00	-	-0.84		-0.92	
Asteraceae	<i>Erigeron argentatus / divergens</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Erigeron argentatus / pumilus</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Erigeron argentatus / speciosus</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Erigeron argentatus / utahensis</i>	0.97	+	0.94		0.00		0.00	
Asteraceae	<i>Erigeron divergens / pumilus</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Erigeron divergens / speciosus</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Erigeron divergens / utahensis</i>	-0.08		-0.08		0.00		0.00	
Asteraceae	<i>Erigeron pumilus / speciosus</i>	0.51		0.40		0.00		0.00	
Asteraceae	<i>Erigeron pumilus / utahensis</i>	-0.25		-0.36		0.00		0.00	
Asteraceae	<i>Erigeron speciosus / utahensis</i>	-0.77		-0.77		0.00		0.00	
Asteraceae	<i>Haplopappus gracilis / canescens</i>	-0.36		-0.36		0.00		0.00	
Asteraceae	<i>Gutierrezia microcephala / sarotheiae</i>	-0.32		0.94		-1.00	-	-1.00	-
Asteraceae	<i>Chrysothamnus depressus / viscidiflorus</i>	-0.11		0.08		0.00		0.00	
Asteraceae	<i>Chrysothamnus nauseosus / parryi</i>	0.60		0.87		0.31		0.35	
Asteraceae	<i>Chrysothamnus nauseosus / linearifolius</i>	-0.84		-0.98	-	0.00		0.00	
Asteraceae	<i>Chrysothamnus parryi / linearifolius</i>	0.00		0.00		0.00		0.00	
Asteraceae	<i>Chaenactis douglasii / stevioides</i>	-0.32		-0.32		0.00		0.00	
Asteraceae	<i>Brickellia atracyloides / californica</i>	-0.39		-0.39		0.00		0.00	
Asteraceae	<i>Brickellia atracyloides / grandiflora</i>	-0.53		-0.67		0.00		0.00	
Asteraceae	<i>Brickellia atracyloides / longifolia</i>	0.23		-0.65		0.00		0.00	
Asteraceae	<i>Brickellia atracyloides / microphylla</i>	0.81		0.38		0.00		0.00	
Asteraceae	<i>Brickellia californica / grandiflora</i>	0.44		0.75		0.00		0.00	
Asteraceae	<i>Brickellia californica / longifolia</i>	-0.58		-0.50		0.00		0.00	
Asteraceae	<i>Brickellia californica / microphylla</i>	-0.21		0.00		0.00		0.00	
Asteraceae	<i>Brickellia grandiflora / longifolia</i>	-0.92		-0.92		0.00		0.00	
Asteraceae	<i>Brickellia grandiflora / microphylla</i>	-0.27		0.00		0.00		0.00	
Asteraceae	<i>Brickellia longifolia / microphylla</i>	0.94		0.98	+	0.00		0.00	
Apiaceae	<i>Osmorhiza depauperata / occidentalis</i>	0.39		0.77		0.00		0.00	
Caprifoliaceae	<i>Symporicarpos longiflorus / oreophilus</i>	-0.84		-0.80		0.00		0.00	

¹Cover-based overlap could not be calculated for *Salix scouleriana* because it lacked cover values in the dataset used for this analysis

APPENDIX 3.3

Results of niche overlap significance tests comparing phylogenetic sister groups (genus and above) for vascular plants recorded in the USGS-NPS vegetation characterization of Zion National Park, Utah (Cogan et al. 2004). Null model distributions of beta niche overlap (across habitats) and alpha niche overlap (across plots within habitats), using both presence and cover measures, were built through randomization and compared with observed overlap values. Prob=probabilities ranging from -1 (more segregated than expected) to 1 (more aggregated than expected). Sig=cases of statistically significant segregation (-) or aggregation (+), i.e. $p \leq 0.05$ for a two-tailed test. Note: This nested sequence of sister groups follows the phylogenetic trees in Fig. 1 and 3-6; sister groups comprised of additive or subtractive combinations of taxa are separated by a slash.

Sister Group Pair	Beta Niche				Alpha Niche			
	Presence		Cover		Presence		Cover	
	Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Moniliformopses / Spermatophyta	-1.00	-	-1.00	-	0.00		-0.29	
<i>Equisetum</i> / Filicales	-0.89		-0.96	-	0.13		0.46	
<i>Pteridium</i> / Eupolypods	0.08		0.30		0.00		0.00	
<i>Asplenium</i> / <i>Polypodium</i>	0.00		0.00		0.00		0.00	
Gymnosperms / Magnoliophyta	-1.00	-	-1.00	-	0.00		-1.00	-
Coniferales / <i>Ephedra</i>	-1.00	-	-0.98	-	0.87		0.67	
Cupressaceae / Pinaceae	-0.99	-	-1.00	-	1.00	+	0.99	+
<i>Cupressus</i> / <i>Juniperus</i>	-0.73		-0.83		0.00		0.00	
<i>Abies</i> / <i>Pinus</i> + <i>Pseudotsuga</i>	-1.00	-	-1.00	-	0.03		0.65	
<i>Pinus</i> / <i>Pseudotsuga</i>	-1.00	-	-1.00	-	-0.20		0.37	
Liliopsida / eudicots	-0.95	-	-1.00	-	0.00		-0.16	
commelinids + Asparagales / <i>Calochortus</i>	0.34		0.18		0.00		-0.03	
commelinids / Asparagales	-0.96	-	-0.67		-0.95	-	-0.71	
<i>Tradescantia</i> / Poales	-0.05		0.01		0.13		0.30	
<i>Typha</i> / Cyperaceae + Juncaceae + Poaceae	-0.88		-0.17		0.00		0.87	
Cyperaceae + Juncaceae / Poaceae	-1.00	-	-1.00	-	-0.17		0.43	
Cyperaceae / Juncaceae	-1.00	-	0.99	+	0.95	+	0.97	+
<i>Carex</i> + <i>Scirpus</i> / <i>Eleocharis</i>	0.16		1.00	+	0.58		0.94	
<i>Carex</i> / <i>Scirpus</i>	-0.76		1.00	+	0.22		0.87	
<i>Juncus</i> / <i>Luzula</i>	1.00	+	1.00	+	0.65		0.70	

Sister Group Pair		Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Poaceae (identified to genus) / Poaceae (not identified to genus)		-0.75		0.32		0.00		0.69	
Aristidoideae + Arundinoideae + Chloridoideae + Panicoideae / Pooideae		-1.00	-	-0.82		0.95		-0.09	
Aristidoideae + Arundinoideae + Chloridoideae / Panicoideae		-0.94		-0.30		0.62		0.88	
Aristida / Arundinoideae + Chloridoideae		-0.67		0.08		0.99	+	-0.04	
<i>Phragmites</i> / Cynodonteae		0.09		-0.01		0.00		0.00	
<i>Muhlenbergia</i> + <i>Hilaria</i> + <i>Bouteloua</i> + <i>Erioneuron</i> / <i>Sporobolus</i> + <i>Distichlis</i>		-0.90		0.22		0.44		0.80	
<i>Muhlenbergia</i> + <i>Hilaria</i> + <i>Bouteloua</i> / <i>Erioneuron</i>		0.05		0.10		0.00		-0.15	
<i>Muhlenbergia</i> + <i>Hilaria</i> / <i>Bouteloua</i>		-1.00	-	-0.83		-0.09		0.38	
<i>Muhlenbergia</i> / <i>Hilaria</i>		-0.99	-	-0.95	-	-0.88		-0.88	
<i>Sporobolus</i> / <i>Distichlis</i>		0.17		0.09		0.00		0.00	
Andropogoneae / <i>Cenchrus</i>		0.00		0.00		0.00		0.00	
<i>Andropogon</i> + <i>Schizachyrium</i> / <i>Sorghastrum</i>		0.94		0.94		0.00		0.00	
<i>Andropogon</i> / <i>Schizachyrium</i>		0.00		0.00		0.00		0.00	
Stipeae / Poeae + Aveneae + Bromeae + Triticeae		-0.79		-0.98	-	-0.37		0.18	
Stipeae (identified to genus) / Stipeae (not identified to genus)		0.58		0.94		0.26		0.55	
<i>Achnatherum</i> + <i>Nassella</i> / <i>Hesperostipa</i>		-0.87		0.99	+	0.03		0.84	
<i>Achnatherum</i> / <i>Nassella</i>		-1.00	-	-0.98	-	0.41		0.84	
Poeae + Aveneae / Bromeae + Triticeae		-1.00	-	-0.70		-0.77		0.75	
Poeae / Aveneae		-0.98	-	-0.99	-	0.86		0.33	
<i>Phleum</i> + <i>Poa</i> + <i>Puccinellia</i> / <i>Dactylis</i> + <i>Festuca</i>		0.44		0.75		0.14		0.26	
<i>Phleum</i> + <i>Poa</i> / <i>Puccinellia</i>		-0.04		0.10		0.00		0.86	
<i>Phleum</i> / <i>Poa</i>		-0.02		0.15		0.00		0.78	
<i>Dactylis</i> / <i>Festuca</i>		-0.63		-0.56		0.00		0.00	
<i>Agrostis</i> + <i>Polypogon</i> + <i>Calamagrostis</i> / <i>Koeleria</i>		-0.95		-0.98	-	0.00		0.00	
<i>Agrostis</i> + <i>Polypogon</i> / <i>Calamagrostis</i>		-0.41		-0.41		0.00		0.00	
<i>Agrostis</i> / <i>Polypogon</i>		-0.37		-0.37		0.00		0.00	
<i>Bromus</i> / Triticeae		-0.99	-	-0.83		0.70		0.21	
<i>Hordeum</i> + <i>Agropyron</i> / <i>Elymus</i>		0.58		0.65		0.65		0.04	
<i>Hordeum</i> / <i>Agropyron</i>		0.00		0.00		0.00		0.00	

Sister Group Pair	Beta Niche				Alpha Niche			
	Presence		Cover		Presence		Cover	
	Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
<i>Sisyrinchium</i> / Alliaceae + Ruscaceae + Agavaceae	-0.08		-0.24		0.61		0.78	
<i>Allium</i> / Ruscaceae + Agavaceae	-0.67		-0.98	-	-0.15		-0.17	
<i>Maianthemum</i> / <i>Yucca</i>	-1.00	-	-1.00	-	0.23		0.09	
Ranunculales / core eudicots	-1.00	-	-1.00	-	0.00		1.00	+
<i>Mahonia</i> / Ranunculaceae	-1.00	-	-0.35		-0.11		0.92	
<i>Clematis</i> / <i>Aquilegia</i> + <i>Thalictrum</i>	0.33		0.80		0.81		0.47	
<i>Aquilegia</i> / <i>Thalictrum</i>	0.33		-0.26		0.00		0.00	
rosids + Saxifragales / Santalales + Caryophyllales + asterids	-0.82		-1.00	-	0.47		-1.00	-
rosids / <i>Heuchera</i>	-1.00	-	-0.96	-	0.00		0.22	
rosids - Vitales / <i>Vitis</i>	-0.65		0.43		0.00		0.90	
fabids / malvids	-1.00	-	-1.00	-	0.95	+	0.93	
fabids - Zygophyllales / Zygophyllales	0.41		-0.01		0.00		-0.25	
Fagales + Rosales + Fabales / Celastrales + Malpighiales	-1.00	-	-1.00	-	0.05		-0.47	
Fagales + Rosales / Fabaceae	-1.00	-	-1.00	-	-0.06		0.83	
Fagales / Rosales	-0.76		-0.95	-	1.00	+	0.55	
<i>Betula</i> / <i>Quercus</i>	-1.00	-	-1.00	-	0.54		-0.42	
Rosaceae / Elaeagnaceae + Rhamnaceae + Ulmaceae	-0.81		-0.94		0.39		-0.31	
Dryadoideae / Rosoideae + Spiraeoideae	-0.92		-0.58		0.91		1.00	+
<i>Cercocarpus</i> / <i>Purshia</i>	-0.60		0.65		0.64		-0.26	
Potentilleae / Spiraeoideae	-1.00	-	-0.01		0.58		-0.92	
<i>Ivesia</i> + Potentilla / <i>Rosa</i>	-0.51		0.36		0.00		0.00	
<i>Ivesia</i> / Potentilla	0.00		0.00		0.00		0.00	
Pyreae + Spiraeae + Kerreae / <i>Prunus</i>	-1.00	-	-1.00	-	-0.08		0.68	
Pyreae + Spiraeae / Coleogyne	-1.00	-	-1.00	-	-0.11		-0.36	
Pyreae / Spiraeae	-0.26		-0.42		-0.28		0.64	
<i>Amelanchier</i> / <i>Peraphyllum</i>	-0.38		0.95	+	-0.11		0.09	
<i>Holodiscus</i> / <i>Petrophytum</i>	-0.14		-0.14		0.42		0.42	

Sister Group Pair		Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
Elaeagnaceae + Rhamnaceae / <i>Celtis</i>		-0.91		-0.76		0.53		0.87	
Elaeagnaceae / <i>Ceanothus</i>		-1.00	-	-1.00	-	0.00		0.00	
<i>Elaeagnus</i> / <i>Shepherdia</i>		-0.99	-	-0.87		0.00		0.00	
Fabaceae (identified to genus) / Fabaceae (not identified to genus)		-0.49		-0.63		0.00		-0.11	
<i>Prosopis</i> / Papilionoideae		0.10		0.22		0.00		0.26	
Genisteeae + Thermopsideae / Amorpheae + Psoraleeae + Hologalegina		-1.00	-	-0.82		0.96	+	0.98	+
<i>Lupinus</i> / Thermopsis		0.20		0.39		0.00		0.00	
Amorpheae / Psoraleeae + Hologalegina		-1.00	-	-1.00	-	0.62		-0.08	
<i>Dalea</i> / <i>Psorothamnus</i>		0.18		-0.16		0.00		0.00	
<i>Psoralidium</i> / Hologalegina		-0.03		-0.35		0.29		0.56	
<i>Lotus</i> / Galegeae + Trifolieae + Fabeae		-0.98	-	-0.95	-	0.98	+	0.34	
Galegeae / Trifolieae + Fabeae		-0.79		-0.56		-0.46		-0.28	
<i>Astragalus</i> / <i>Oxytropis</i>		-0.75		-0.80		0.00		0.00	
Trifolieae clade 1 / Trifolieae clade 2 + Fabeae		-1.00	-	-1.00	-	-0.08		-0.63	
<i>Medicago</i> / <i>Melilotus</i>		1.00	+	1.00	+	0.13		0.13	
<i>Trifolium</i> / Fabeae		-0.99	-	-0.98	-	0.28		0.70	
<i>Lathyrus</i> / <i>Vicia</i>		-0.97	-	-0.94		0.56		0.54	
<i>Pachystima</i> / Malpighiales		-1.00	-	-1.00	-	0.88		0.14	
Euphorbiaceae + Linaceae / Salicaceae + Violaceae		-1.00	-	-1.00	-	0.23		-0.40	
Euphorbiaceae / <i>Linum</i>		0.06		-0.03		-0.13		-0.13	
<i>Tragia</i> / Euphorbieae		0.16		0.11		0.00		0.00	
<i>Chamaesyce</i> / Euphorbia		-0.76		-0.80		-0.44		-0.44	
Salicaceae / <i>Viola</i>		-0.93		-0.99	-	0.00		0.00	
<i>Populus</i> / <i>Salix</i>		0.92		0.86		0.96	+	-0.09	
<i>Tribulus</i> / Krameria		0.00		0.00		0.00		0.00	
Brassicaceae + Malvales + Sapindales / Geraniales + Myrtales		-0.96	-	-1.00	-	0.04		-0.12	
Brassicaceae + Malvales / Sapindales		-1.00	-	-1.00	-	-0.53		0.25	
Brassicaceae / Sphaeralcea		-0.62		-0.19		0.16		-0.26	

Sister Group Pair		Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
	Brassicaceae (identified to genus) / Brassicaceae (not identified to genus)	-0.94		-0.83		0.25		0.73	
	Arabideae + Sisymbrieae + Schizopetaleae + Cardamineae + Lepidieae + Descurainieae + Physarieae / <i>Erysimum</i>	0.13		-0.29		0.68		0.75	
	Arabideae + Sisymbrieae + Schizopetaleae + Cardamineae + Lepidieae + Descurainieae / <i>Physaria</i>	0.26		-0.19		-0.14		-0.23	
	Arabideae + Sisymbrieae + Schizopetaleae + Cardamineae / Lepidieae + Descurainieae	-0.64		-0.62		-0.13		-0.13	
	Arabideae + Sisymbrieae + Schizopetaleae / <i>Nasturtium</i>	-0.23		0.09		0.00		0.00	
	Arabideae / Sisymbrieae + Schizopetaleae	-0.78		-0.97	-	-0.13		-0.13	
	<i>Arabis</i> / <i>Draba</i>	1.00	+	1.00	+	-0.04		-0.04	
	<i>Sisymbrium</i> / Schizopetaleae	-0.77		-0.38		0.00		0.00	
	<i>Streptanthus</i> / <i>Stanleya</i>	-0.35		-0.35		0.00		0.00	
	<i>Lepidium</i> / Descurainia	-0.30		-0.30		0.00		0.00	
	Anardiaceae / <i>Acer</i>	-1.00	-	-1.00	-	-0.07		-0.47	
	<i>Rhus</i> / <i>Toxicodendron</i>	0.80		0.84		0.00		0.00	
	Geraniaceae / Onagraceae	-0.92		-0.93		0.18		0.18	
	<i>Erodium</i> / <i>Geranium</i>	-0.56		-0.56		0.00		0.00	
	<i>Epilobium</i> / Onagreae	-0.72		-0.72		0.00		0.00	
	<i>Gayophytum</i> / <i>Oenothera</i>	-0.64		-0.64		0.00		0.00	
	Santalaceae / Caryophyllales + asterids	-0.94		0.25		0.09		0.95	
	<i>Phoradendron</i> / <i>Comandra</i>	-0.16		-0.28		0.25		0.27	
	Caryophyllales / asterids	-1.00	-	-1.00	-	0.82		0.54	
	Polygonaceae + Tamaricaceae / Amaranthaceae + Caryophyllaceae + Nyctaginaceae + Cactaceae + Portulacaceae	-1.00	-	-0.96	-	0.98	+	1.00	+
	Polygonaceae / <i>Tamarix</i>	-1.00	-	-1.00	-	0.38		0.71	
	<i>Eriogonum</i> / Polygonoideae	-0.95	-	-0.96	-	0.79		0.74	
	<i>Polygonum</i> / <i>Rumex</i>	-0.97	-	-0.97	-	0.00		0.00	
	Amaranthaceae + Caryophyllaceae / Nyctaginaceae + Cactaceae + Portulacaceae	-1.00	-	-0.28		0.97	+	0.96	+
	Amaranthaceae / Caryophyllaceae	-1.00	-	-1.00	-	0.22		-0.24	
	<i>Atriplex</i> + <i>Chenopodium</i> + <i>Grayia</i> / <i>Salsola</i>	0.10		0.41		0.00		0.00	
	<i>Atriplex</i> + <i>Chenopodium</i> / <i>Grayia</i>	0.83		0.85		0.39		-0.08	
	<i>Atriplex</i> / <i>Chenopodium</i>	-0.97	-	-0.99	-	0.38		0.13	

Sister Group Pair		Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
	<i>Arenaria + Stellaria</i> / Caryophyllaceae (not identified to genus)	-0.87		-0.52		0.00		0.00	
	<i>Arenaria</i> / <i>Stellaria</i>	-0.83		-0.79		0.00		0.00	
	Nyctaginaceae / Cactaceae + Portulacaceae	-0.97	-	-0.82		0.90		0.72	
	<i>Abronia</i> / <i>Allionia</i> + <i>Mirabilis</i>	-0.58		-0.58		0.00		0.00	
	<i>Allionia</i> / <i>Mirabilis</i>	0.99	+	0.99	+	0.66		0.66	
	Cactaceae / <i>Claytonia</i>	-1.00	-	-1.00	-	-0.01		-0.01	
	<i>Echinocereus</i> / <i>Opuntia</i>	-0.98	-	-1.00	-	0.98	+	0.68	
	<i>Fendlerella</i> / asterids - Cornales	-0.32		-0.66		0.00		-0.66	
	Ericales / lamiids + campanulids	-1.00	-	-1.00	-	-0.84		-1.00	-
	Polemonioideae / Ericaceae	0.95		0.40		0.99	+	0.09	
	Phlocideae / Gilieae + Loeselieae	-0.46		0.83		-0.19		-0.03	
	<i>Phlox</i> + <i>Leptosiphon</i> / Linanthus	-0.79		-0.85		0.00		0.00	
	<i>Phlox</i> / <i>Leptosiphon</i>	0.77		0.75		0.00		0.00	
	<i>Gilia</i> / <i>Ipomopsis</i>	0.97	+	0.89		0.00		0.00	
	<i>Arctostaphylos</i> / Monotropoideae	-0.84		-0.95	-	-0.10		0.43	
	<i>Pterospora</i> / <i>Chimaphila</i>	0.00		0.00		0.00		0.00	
	lamiids / campanulids	-0.52		-0.96	-	0.97	+	0.46	
	Lamiales + Solanales + Gentianales / Boraginaceae	-0.25		0.84		0.58		-0.23	
	Lamiales + Solanales / Gentianales	-0.99	-	-0.95		0.65		0.67	
	Lamiales / Solanales	-0.99	-	0.13		0.05		0.88	
	Lamiaceae + Orobanchaceae + Verbenaceae + Scrophulariaceae + Plantaginaceae / <i>Fraxinus</i>	-0.83		-1.00	-	0.27		-0.47	
	Lamiaceae + Orobanchaceae + Verbenaceae + Scrophulariaceae / Plantaginaceae	-0.88		-0.93		0.53		0.97	+
	Lamiaceae + Orobanchaceae + Verbenaceae / <i>Verbascum</i>	-0.76		-0.68		0.61		0.81	
	Lamiaceae + Orobanchaceae / <i>Verbena</i>	-0.34		-0.58		0.00		0.00	
	Mentheae / Rhinantheae	0.02		0.44		-0.28		-0.22	
	<i>Agastache</i> + <i>Dracocephalum</i> + <i>Mentha</i> + <i>Monardella</i> / <i>Salvia</i>	-0.98	-	-0.96	-	0.00		0.00	
	<i>Agastache</i> + <i>Dracocephalum</i> / <i>Mentha</i> + <i>Monardella</i>	-0.84		-0.84		0.00		0.00	
	<i>Agastache</i> / <i>Dracocephalum</i>	0.00		0.00		0.00		0.00	
	<i>Mentha</i> / <i>Monardella</i>	0.00		0.00		0.00		0.00	

Sister Group Pair		Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
	<i>Castilleja / Cordylanthus</i>	-0.25		-0.45		0.86		0.82	
	Antirrhineae + Plantagineae / Cheloneae	-0.98	-	-0.97	-	-0.06		-0.05	
	<i>Linaria / Plantago</i>	-0.11		-0.11		0.00		0.00	
	<i>Collinsia / Penstemon</i>	-0.51		-0.69		0.23		0.26	
	Convolvulaceae / Solanaceae	0.50		0.36		0.24		0.76	
	<i>Convolvulus / Ipomoea</i>	0.84		0.89		0.00		0.00	
	<i>Nicotiana / Solanoideae</i>	0.17		0.10		0.00		0.00	
	Physaleae + Solaneae + Datureae / <i>Lycium</i>	-0.97	-	-0.85		0.59		0.67	
	Physaleae + Solaneae / <i>Datura</i>	-0.16		-0.16		0.71		0.71	
	<i>Physalis / Solanum</i>	0.00		0.00		0.00		0.00	
	Apocynaceae + Gentianaceae / Rubioideae	0.22		-0.64		0.00		0.00	
	Apocynaceae / <i>Swertia</i>	0.36		0.21		0.00		0.00	
	<i>Apocynum / Asclepias</i>	0.00		0.00		0.00		0.00	
	<i>Kelloggia / Galium</i>	0.32		-0.60		0.00		0.00	
	Boraginaceae (identified to genus) / Boraginaceae (not identified to genus)	0.09		-0.19		0.00		0.00	
	Boraginoideae / Hydrophylloideae	-0.85		0.52		-0.92		-0.92	
	<i>Cryptantha + Plagiobothrys / Mertensia</i>	-0.99	-	-0.97	-	0.00		0.00	
	<i>Cryptantha / Plagiobothrys</i>	-0.42		-0.51		0.00		0.00	
	<i>Hydrophyllum / Phacelia</i>	-0.78		-0.83		0.00		0.00	
	Asteraceae / Apiales + Dipsacales	-1.00	-	-1.00	-	-0.84		-1.00	-
	Asteraceae (identified to genus) / Asteraceae (not identified to genus)	-0.45		-0.71		0.18		0.53	
	<i>Cirsium / Cichorioideae + Asteroideae</i>	-0.90		-0.60		0.47		-0.20	
	Chicorieae / Asteroideae	-1.00	-	-0.99	-	0.92		0.64	
	<i>Agoseris + Stephanomeria + Lygodesmia + Crepis + Taraxacum + Hypochaeris + Lactuca / Tragopogon</i>	-0.78		-0.27		-0.04		-0.14	
	<i>Agoseris + Stephanomeria + Lygodesmia / Crepis + Taraxacum + Hypochaeris + Lactuca</i>	-0.65		-0.48		0.00		0.00	
	<i>Agoseris + Stephanomeria / Lygodesmia</i>	0.84		0.58		0.00		0.00	
	<i>Agoseris / Stephanomeria</i>	0.41		0.42		0.00		0.00	

Sister Group Pair		Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
	<i>Crepis + Taraxacum + Hypochaeris / Lactuca</i>	0.32		0.32		0.82		0.82	
	<i>Crepis + Taraxacum / Hypochaeris</i>	-0.18		-0.18		0.00		0.00	
	<i>Crepis / Taraxacum</i>	0.52		0.52		0.00		0.00	
	<i>Senecioneae / Asteroideae - Senecioneae</i>	-0.87		0.93		0.81		-0.26	
	<i>Senecio / Tetradymina</i>	-0.85		-0.90		0.36		0.46	
	Anthemideae + Astereae + Gnaphalieae / Plucheeae + Helenieae + Bahieae + Chaenactideae + Eupatorieae + Perityleae + Madieae + Heliantheae	-0.96	-	-0.76		0.94		-0.53	
	Anthemideae + Astereae / <i>Antennaria</i>	-0.83		-0.84		0.79		0.85	
	Anthemideae / Astereae	-0.72		-1.00	-	0.99	+	0.43	
	<i>Achillea / Artemisia + Sphaeromeria</i>	-1.00	-	-1.00	-	0.29		-0.96	-
	<i>Artemisia / Sphaeromeria</i>	-0.07		-0.07		-0.16		0.35	
	<i>Baccharis / Astereae - Baccharis</i>	-1.00	-	-0.99	-	0.50		0.55	
	<i>Erigeron + Chrysopsis + Chaetopappa + Townsendia + Symphyotrichum + Eurybia + Machaeranthera + Gutierrezia / Solidago + Petradoria + Chrysanthmnus + Hesperodoria + Ericameria</i>	-0.72		-1.00	-	-0.26		-0.71	
	<i>Erigeron + Chrysopsis + Chaetopappa + Townsendia + Symphyotrichum + Eurybia + Machaeranthera / Gutierrezia</i>	-1.00	-	-1.00	-	-0.60		-0.95	-
	<i>Erigeron + Chrysopsis + Chaetopappa + Townsendia / Symphyotrichum + Eurybia + Machaeranthera</i>	0.21		0.08		0.77		0.93	
	<i>Erigeron + Chrysopsis + Chaetopappa / Townsendia</i>	0.79		-0.12		0.80		0.52	
	<i>Erigeron + Chrysopsis / Chaetopappa</i>	-0.40		0.17		0.26		0.81	
	<i>Erigeron / Chrysopsis</i>	-0.90		-0.56		0.93		0.73	
	<i>Asterinae spp. + Symphyotrichum / Eurybia + Machaeranthera</i>	0.59		0.13		-0.23		0.37	
	<i>Asterinae spp. / Symphyotrichum</i>	-0.24		-0.24		0.00		0.00	
	<i>Eurybia / Machaeranthera</i>	-0.22		-0.22		0.00		0.00	
	<i>Solidago + Petradoria + Chrysanthmnus + Hesperodoria / Ericameria</i>	-0.96	-	0.05		-0.02		-0.06	
	<i>Solidago + Petradoria / Chrysanthmnus + Hesperodoria</i>	-0.98	-	-0.78		0.00		0.66	
	<i>Solidago / Petradoria</i>	0.97	+	1.00	+	0.00		0.00	
	<i>Chrysanthmnus / Hesperodoria</i>	-0.71		-0.39		0.00		0.00	

Sister Group Pair		Beta Niche				Alpha Niche			
		Presence		Cover		Presence		Cover	
		Prob	Sig	Prob	Sig	Prob	Sig	Prob	Sig
	<i>Pluchea</i> / Helenieae + Bahieae + Chaenactideae + Eupatorieae + Madieae + Heliantheae	-0.90		0.91		0.00		0.00	
	Helenieae / Bahieae + Chaenactideae + Eupatorieae + Madieae + Heliantheae	0.07		-0.31		0.00		0.00	
	<i>Baileya</i> / <i>Gaillardia</i>	0.00		0.00		0.00		0.00	
	Bahieae + Chaenactideae / Eupatorieae + Madieae + Heliantheae	-0.41		-0.95	-	-0.59		-0.93	
	<i>Hymenopappus</i> / <i>Chaenactis</i>	0.02		0.02		-0.36		-0.36	
	Eupatorieae + Madieae / Heliantheae	-0.77		-0.90		-0.76		-0.67	
	<i>Brickellia</i> / <i>Arnica</i>	-0.03		-0.23		0.00		0.00	
	<i>Ambrosia</i> + <i>Helianthus</i> + <i>Viguiera</i> + <i>Geraea</i> + <i>Balsamorhiza</i> + <i>Wyethia</i> / <i>Helianthella</i>	-0.03		-0.48		0.82		0.96	+
	<i>Ambrosia</i> + <i>Helianthus</i> + <i>Viguiera</i> + <i>Geraea</i> / <i>Balsamorhiza</i> + <i>Wyethia</i>	0.40		0.91		0.00		0.00	
	<i>Ambrosia</i> / <i>Helianthus</i> + <i>Viguiera</i> + <i>Geraea</i>	-0.46		-0.46		0.00		0.00	
	<i>Helianthus</i> + <i>Viguiera</i> / <i>Geraea</i>	-0.43		-0.43		0.00		0.00	
	<i>Helianthus</i> / <i>Viguiera</i>	-0.34		-0.34		0.00		0.00	
	<i>Balsamorhiza</i> / <i>Wyethia</i>	0.98	+	0.43		0.79		0.81	
	Apiaceae / Dipsacales	-0.75		0.36		0.95		0.86	
	<i>Lomatium</i> / <i>Osmorhiza</i>	0.94		0.85		0.28		0.61	
	<i>Sambucus</i> / <i>Symporicarpos</i>	-0.76		-0.86		0.00		0.00	